



The Abdus Salam  
International Centre  
for Theoretical Physics



*Conference on Atomistic Simulations of Biomolecules:  
towards a Quantitative Understanding of Life Machinery  
(6-10 March 2017)*

# **Dynamics of ordered counterions in the ion-hydrate shell of DNA double helix**

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Bogolyubov Institute for Theoretical Physics, NAS of Ukraine  
Biophysics of Macromolecules Lab

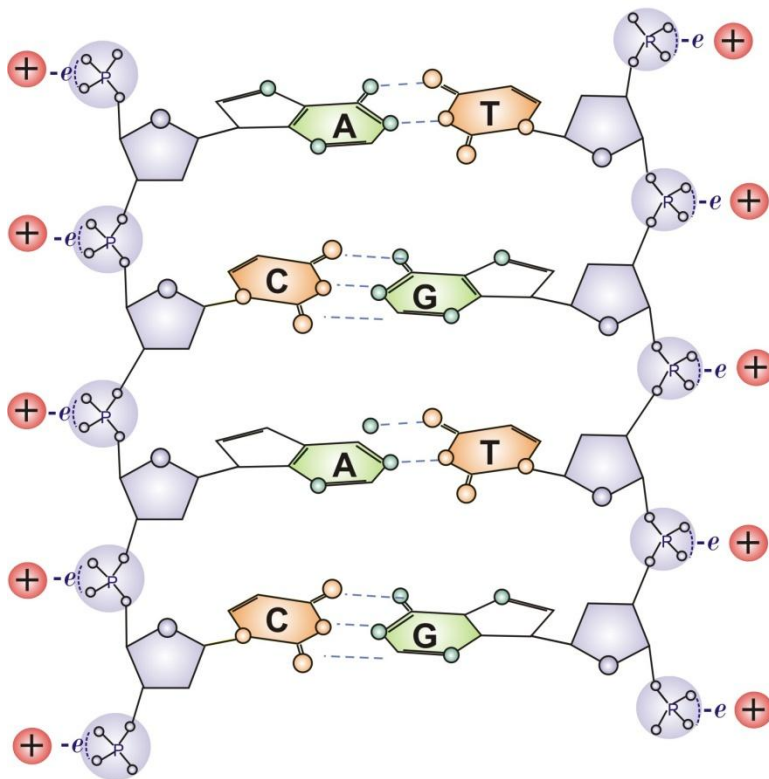
06.03.2017

# Outline

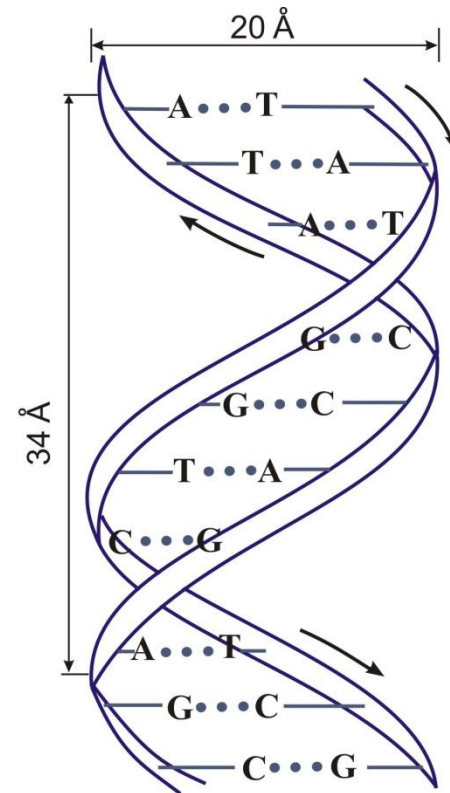
- 1. Ion-hydrate environment in the structure formation of DNA double helix.**  
water molecules in the hydration shells of DNA double helix;  
counterion atmosphere around DNA macromolecule.
- 2. Counterion vibrations in the low-frequency spectra of DNA.**  
model of conformational vibrations of DNA with counterions;  
manifestations of counterion dynamics in the low-frequency spectra.
- 3. MD simulations of counterion dynamics around DNA double helix.**  
characteristic binding sites of  $\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Cs}^+$  and  $\text{Mg}^{2+}$  with DNA;  
dynamics of counterions, tethered to DNA;  
dynamics of counterion dissociation;
- 4. Conclusions.**

# DNA double helix (1953)

## Structure elements of DNA



## DNA double helix



James Watson  
1928



Francis Crick  
1916 - 2004



Maurice Wilkins  
1916-2004



Rosalind  
Franklin

1920 - 1958

*Watson J.D., Crick F.H.C. A structure of deoxyribose nucleic acid. Nature. 171, 737-738 (1953).*

*Wilkins M.H.F., Stokes A.R., Wilson H.R. Molecular structure of deoxypentose nucleic acid. Nature. 171, 738-740 (1953).*

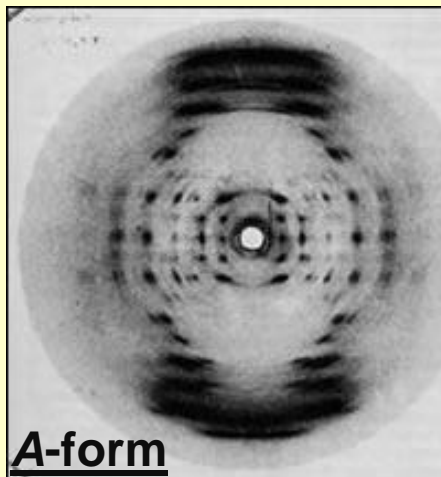
*Franklin R.E., Gosling R.G., Molecular configuration in sodium thymonucleate. Nature. 171, 740-741 (1953).*

# Different forms of DNA double helix

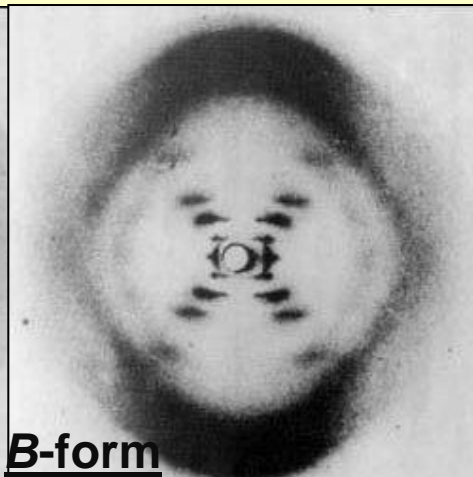


Rosalind Franklin (1920 – 1958)

X-ray diffraction images of DNA (Photo #51)



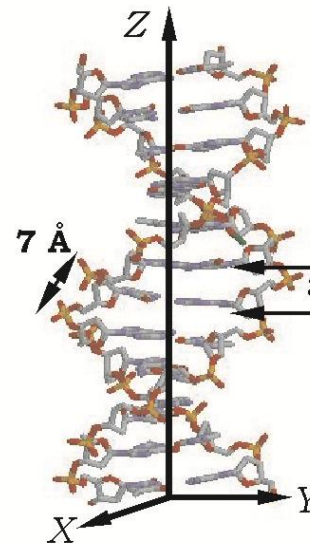
A-form



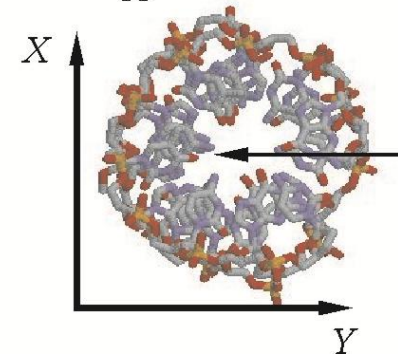
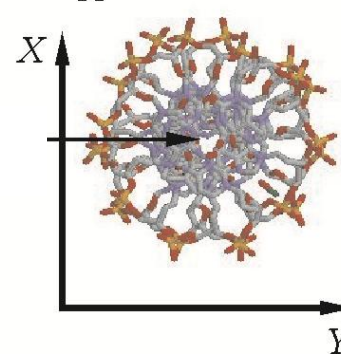
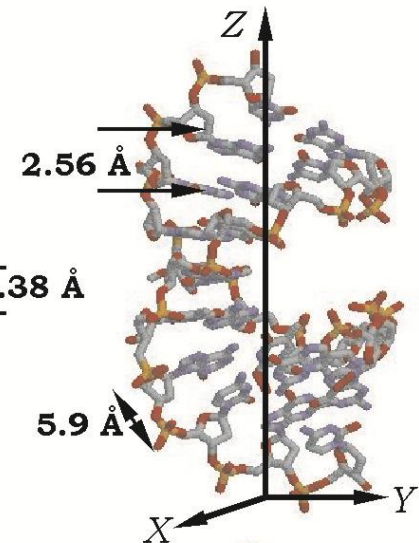
B-form

Franklin R.E., Gosling R.G., Molecular configuration in sodium thymonucleate. *Nature*. 171, 740-741 (1953).

**B-form**



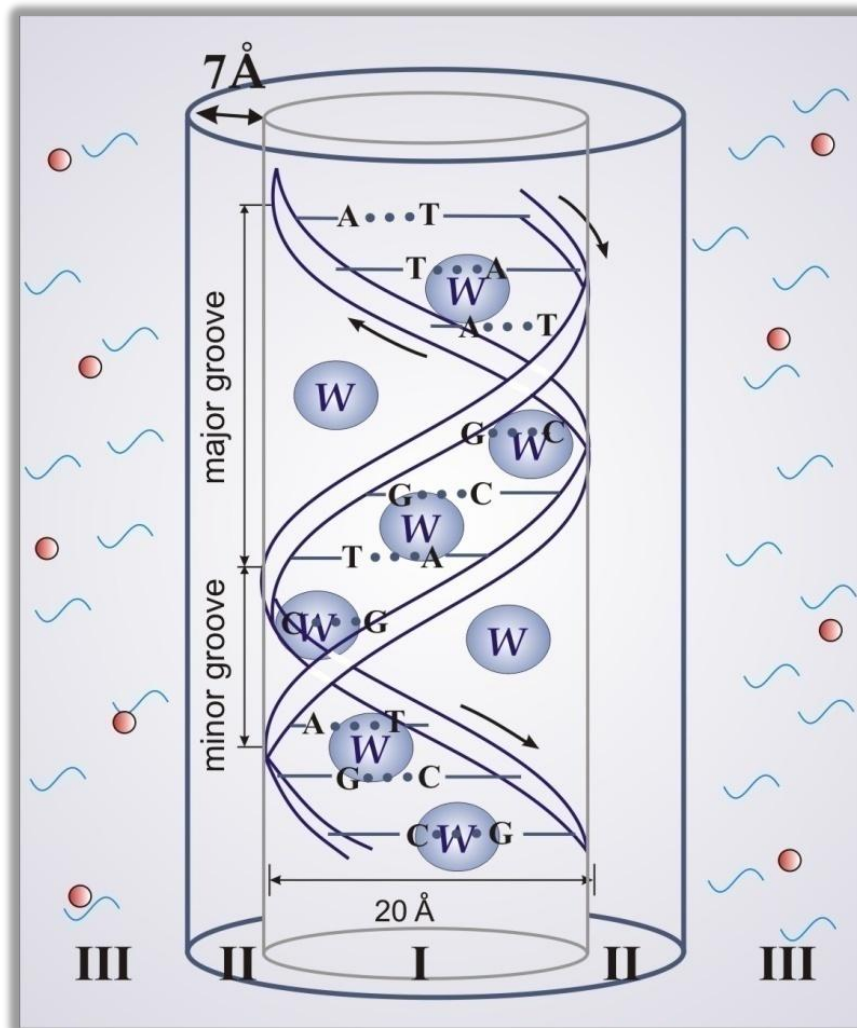
**A-form**



NDB bd0007

Tereshko V., Minasov G., Egli M., *J.Am.Chem.Soc.* 121, 470 (1999).

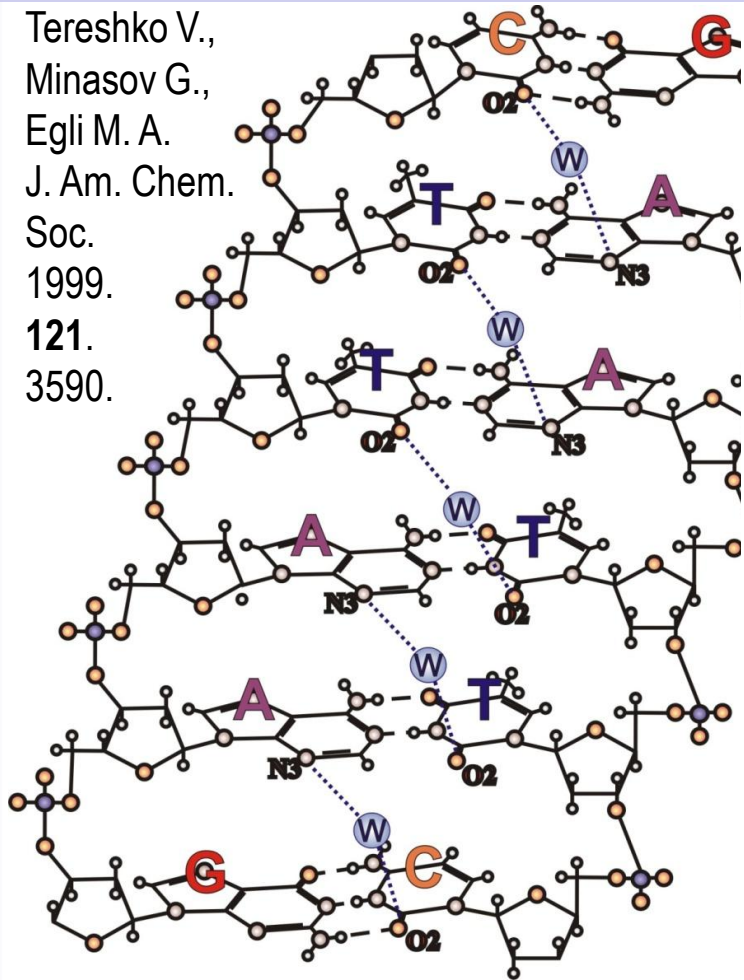
# Structure of the ion-hydrate shell



I – the first hydration shell (20w/bp);  
II – the second hydration shell (30w/bp);  
III – bulk water.

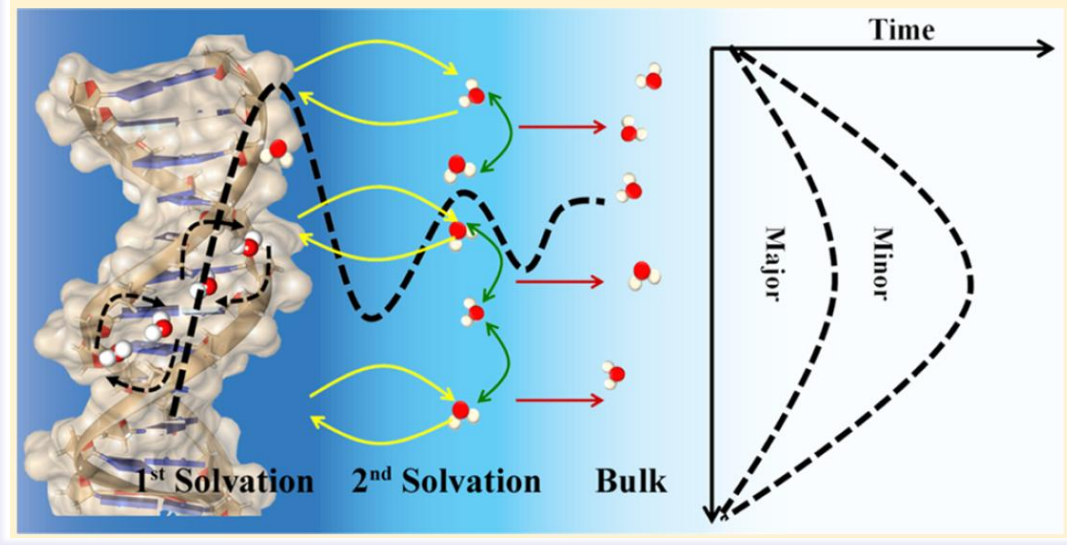
# Hydration of DNA grooves

Tereshko V.,  
Minasov G.,  
Egli M. A.  
J. Am. Chem.  
Soc.  
1999.  
121.  
3590.

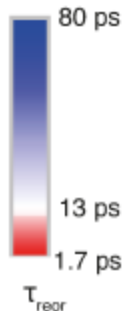
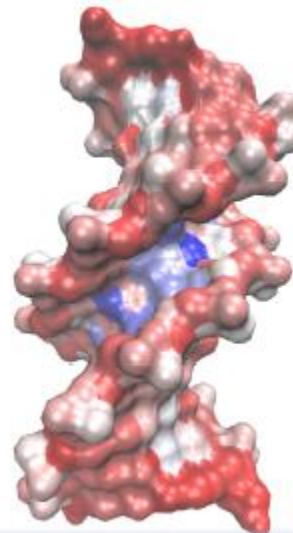


Drew H.R., Wing R.M., Takano T., Broka C.,  
Takana S., Itakura K., Dickerson R.E., *Proc.  
Natl. Acad. Sci. USA*, **78**,2179,1981.

D. Saha, S. Supekar, A. Mukherjee. "Distribution of Residence Time of Water around DNA Base Pairs: Governing Factors and the Origin of Heterogeneity" *J. Phys. Chem. B* 2015, 119, 11371–11381



E. Duboué-Dijon, A.C. Fogarty, J.T. Hynes, D. Laage. "Dynamical disorder in the DNA hydration shell" *J Am Chem Soc* 2016.



The assumption that hydration shell dynamics is much faster than DNA dynamics is thus not valid. Biomolecular conformational fluctuations are essential to facilitate the water motions and accelerate the hydration dynamics in confined groove sites.

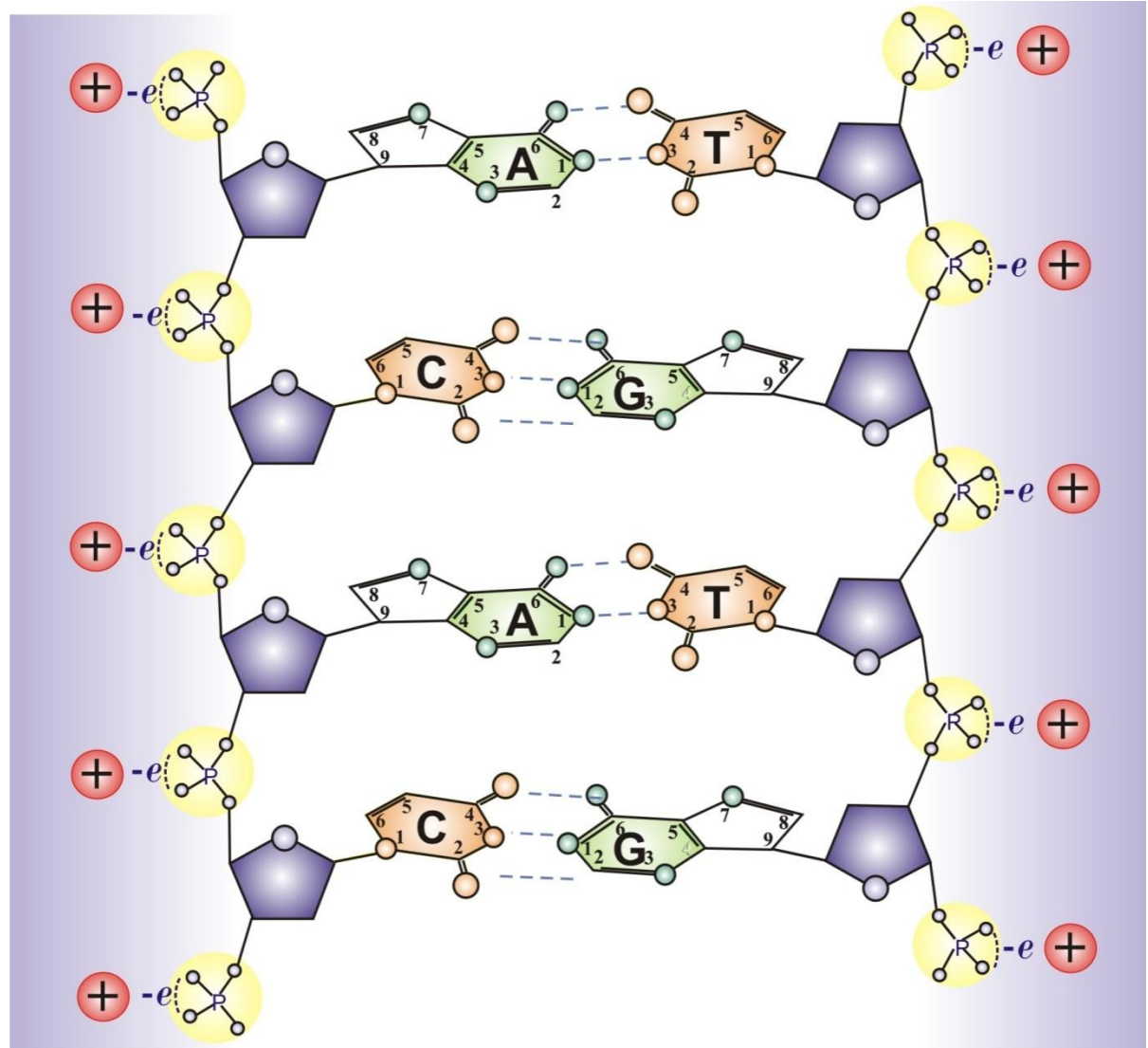
# Stabilization of the double helix by counterions

Under the natural conditions the phosphate groups are neutralized by metal counterions:

- $\text{Na}^+$
- $\text{K}^+$
- $\text{Mg}^{2+}$
- .....

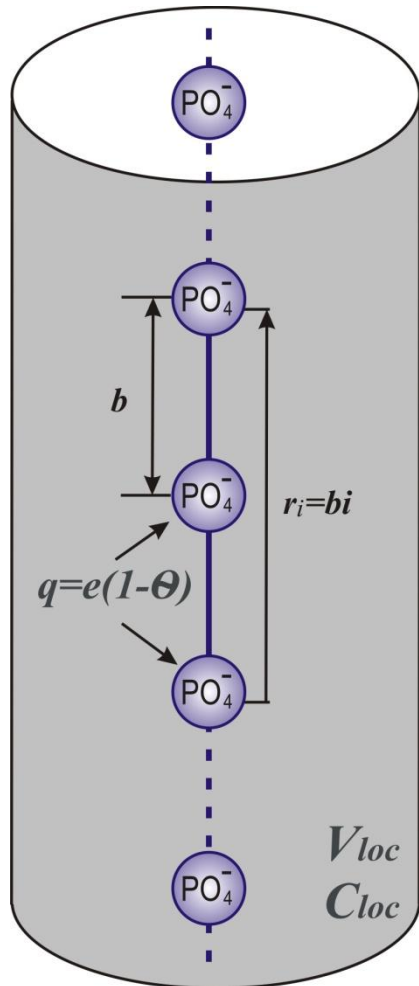
*Metals in a human body*

- $\text{Na}^+$   
100 (g/70 kg)
- $\text{K}^+$   
140(g/70 kg)



# Manning counterion condensation theory

Manning G.S.  
*Quant.Rev.Biophys.*  
 11, 179 (1978).



## Energy of the polyelectrolyte:

$$G = G_{rep} + G_{mix};$$

### Electrostatic contribution:

$$G_{rep} = \frac{(1-\theta)^2 e^2}{4\pi\epsilon\epsilon_0} \sum_{ij} \frac{\exp(-r_{ij} / r_D)}{r_{ij}};$$

### Entropy contribution:

$$G_{mix} = kTN\theta \ln \frac{C_{loc}}{C};$$

$$\frac{dg}{d\theta} = 0, \quad \frac{l_B}{b} (1-\theta) - 1 = 0; \quad \theta = 1 - \frac{b}{l_B}. \quad l_B = \frac{e^2}{4\pi\epsilon\epsilon_0 kT};$$

not charges  
correlated

$$b > l_B$$

$$b < l_B$$

Condensation  
of counterions

Form	$l_B/b$	$C_{loc}(\text{M})$	$L (\text{\AA})$	$\theta$
<b>B-DNA</b>	4.2	1.18	7.4	0.76
<b>A-DNA</b>	5.4	2.07	5.9	0.82



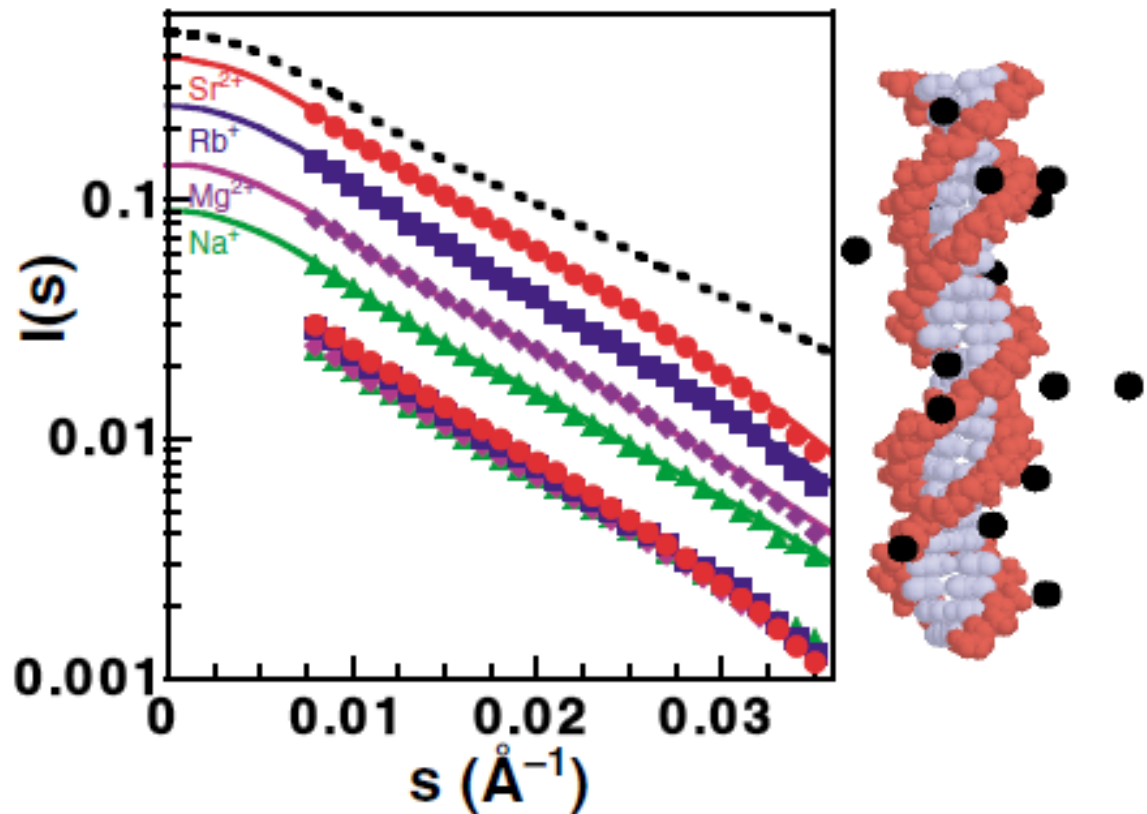
# Localization of counterions around DNA: experiments

## Counterion coating

The experimental data show that the counterions form the cloud around the macromolecule. The data agree with the calculations of distributions of the ions within the framework of nonlinear PB equation.

Das R., et. al.  
*Phys.Rev.Lett.* **90**, 188103 (2003).

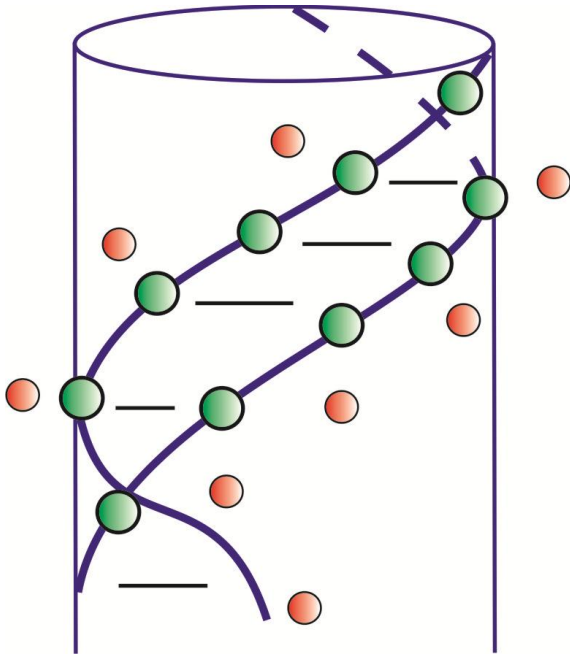
Small angle X-ray scattering profiles for DNA in presence of different counterions (0.4M).



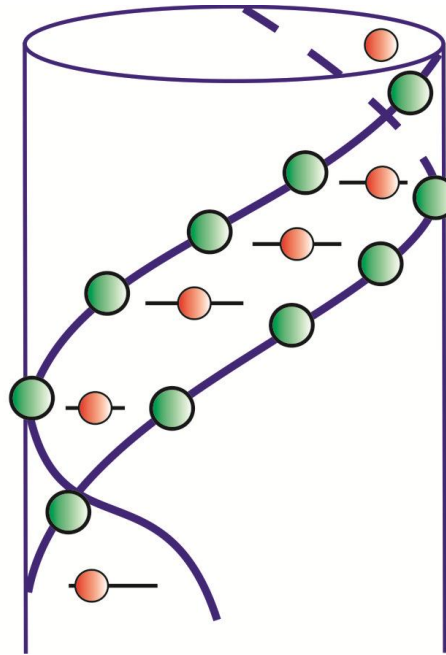
Actual curves (bottom) are shown, as well as offset curves (top), to aid visual comparison with calculations for DNA without modeled counterions (dashed line) and with NLPB ion atmospheres (solid lines).

# Ion-phosphate lattice of DNA

Single-stranded neutralization

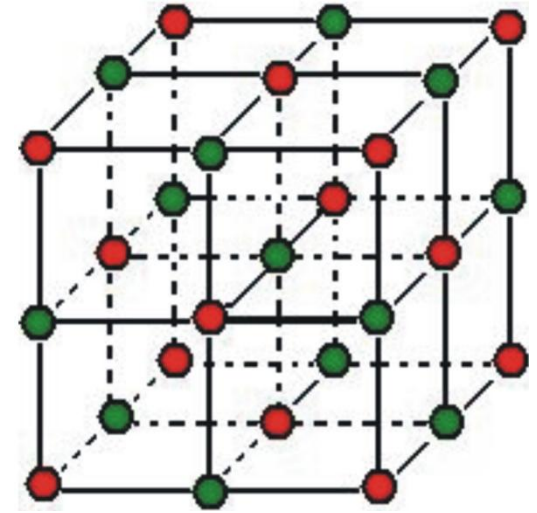


Cross-stranded neutralization



$\text{Na}^+$ ,  $\text{K}^+$ ,  $\text{Rb}^+$ ,  $\text{Cs}^+$ ,  $\text{Mg}^{2+}$

NaCl Crystal



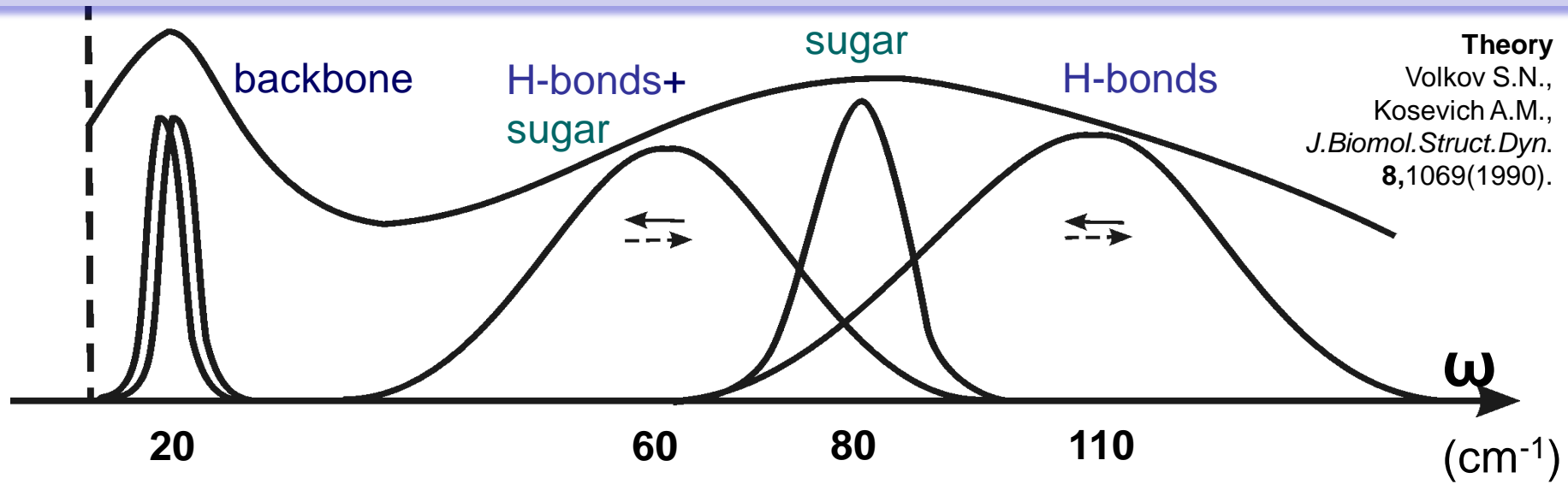
Vibrations in the ionic crystals  $<200 \text{ cm}^{-1}$ .

C. Kittel *Introduction to solid state physics* (1954).

S.M. Perepelytsya, S.N. Volkov,  
*Ukr. J. Phys.* **49**, 1182 (2004).  
*Eur. Phys. J. E.* **24**, 261 (2007).

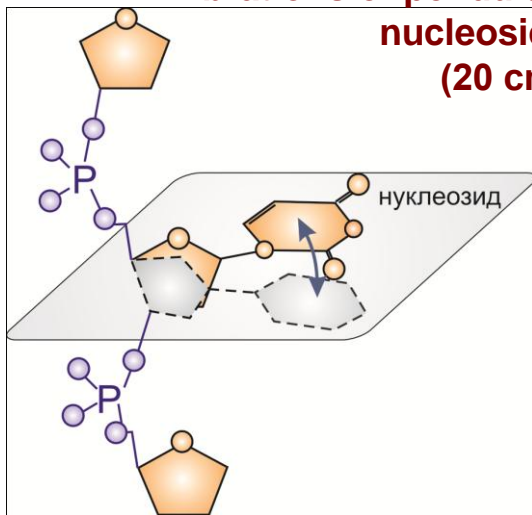
The goal is to study the manifestations of ion-phosphate lattice vibrations in the low-frequency spectra of DNA double helix.

# DNA low-frequency spectra

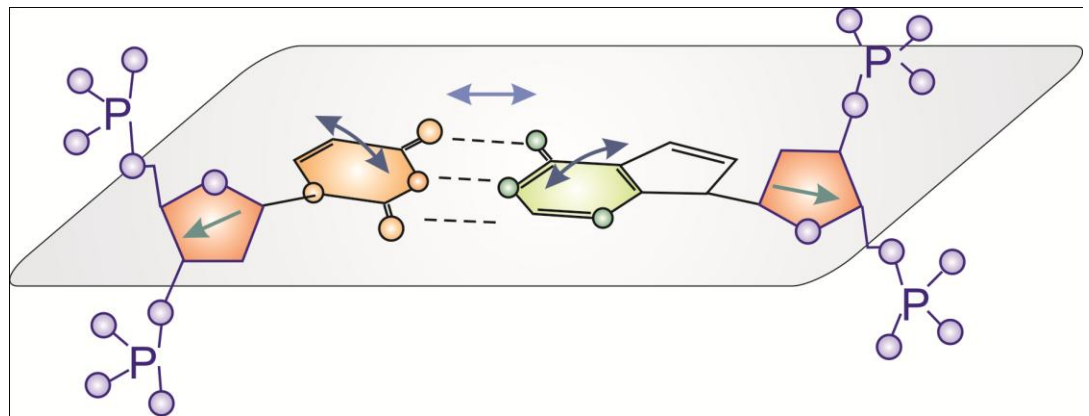


## Conformational vibrations of DNA double helix

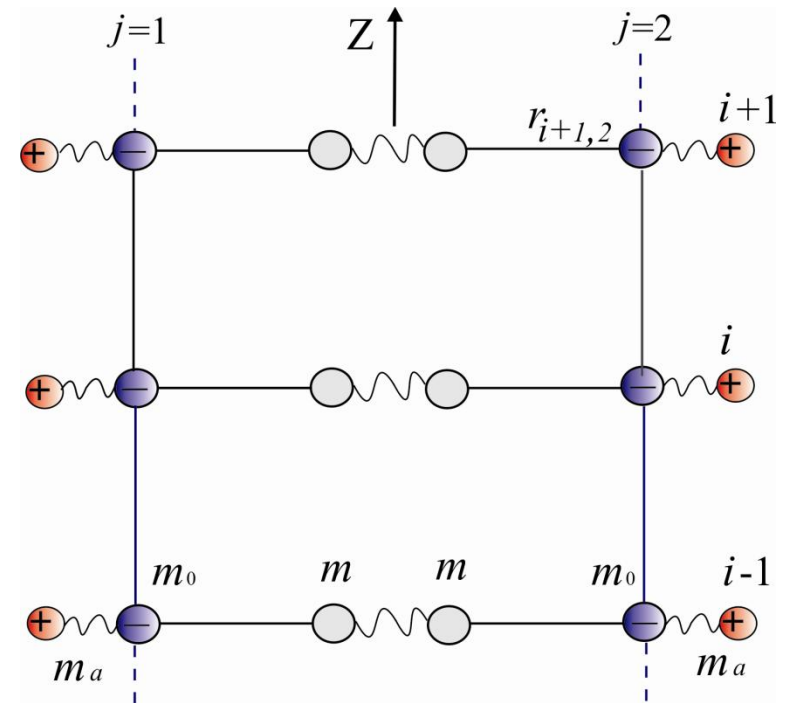
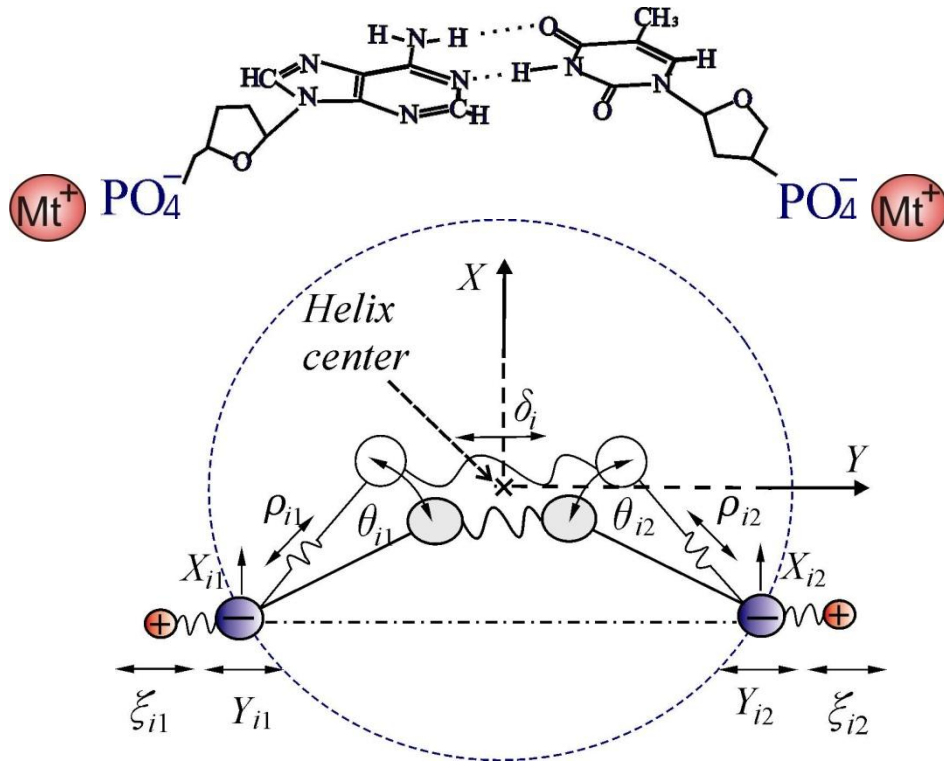
**Vibrations of pendulum-nucleosides (20 cm<sup>-1</sup>)**



**H-bond stretching vibrations (60, 80 and 110 cm<sup>-1</sup>)**

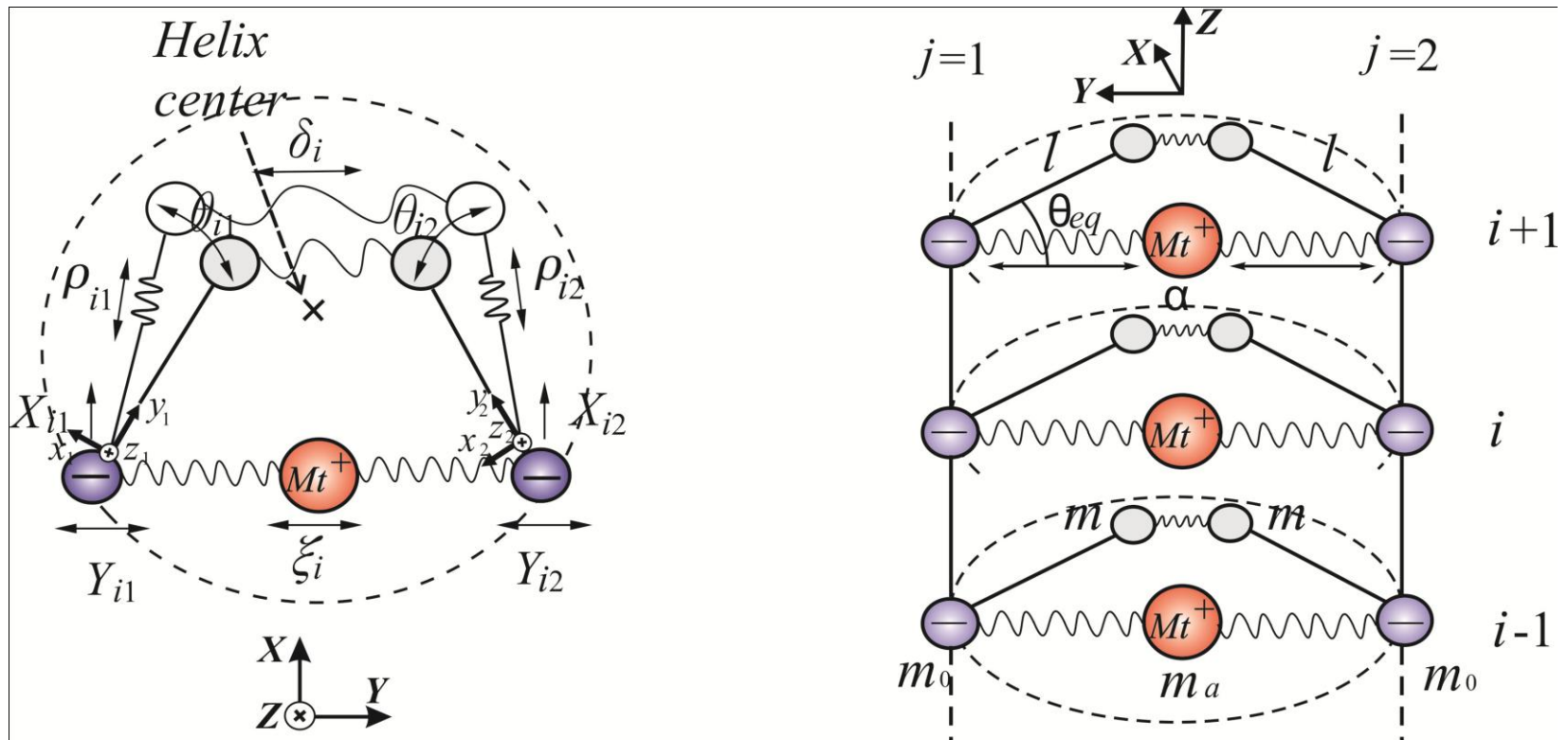


# Model of DNA conformational vibrations



**Ререлытська С.М., Волков С.Н. Ukr. J. Phys., 49, 1072 (2004).**  
**Перепелица С.Н., Волков С.Н. Біофізичний вісник (2005).**  
**Ререлытська С.М., Волков С.Н. Eur. Phys. J. E. (2007).**

# The model for counterion in the minor groove of the double helix



S.M. Perepelytsya, S.N. Volkov, *Ukr. J. Phys.* **55**, 1182-1188 (2010).  
 S.M. Perepelytsya S.N. Volkov, *J. Molec. Liq.* **5**, 113-119 (2011).

# Vibrational energy

$$E = \sum_i (E_i^h + E_i^c + E_{i,i+1}),$$

Energy of the monomer link:

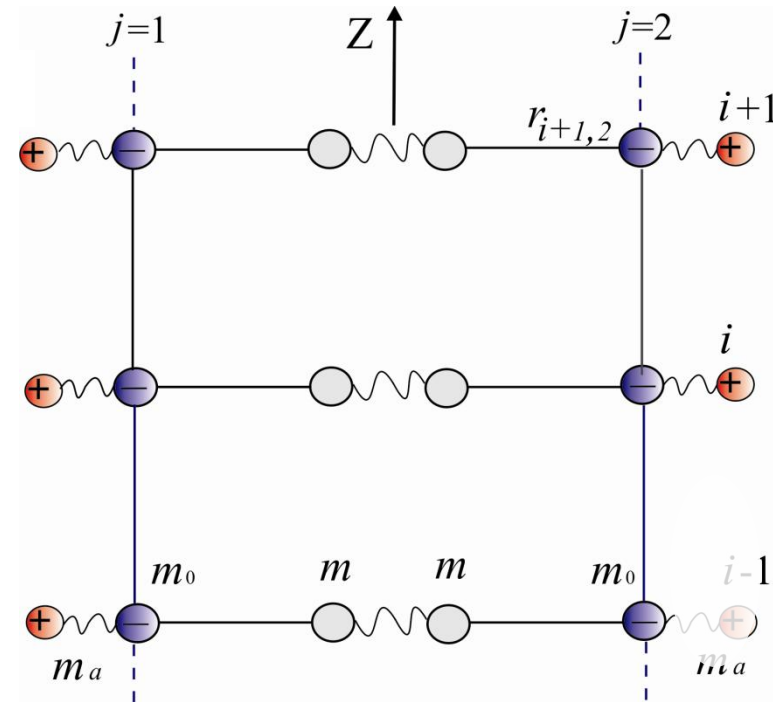
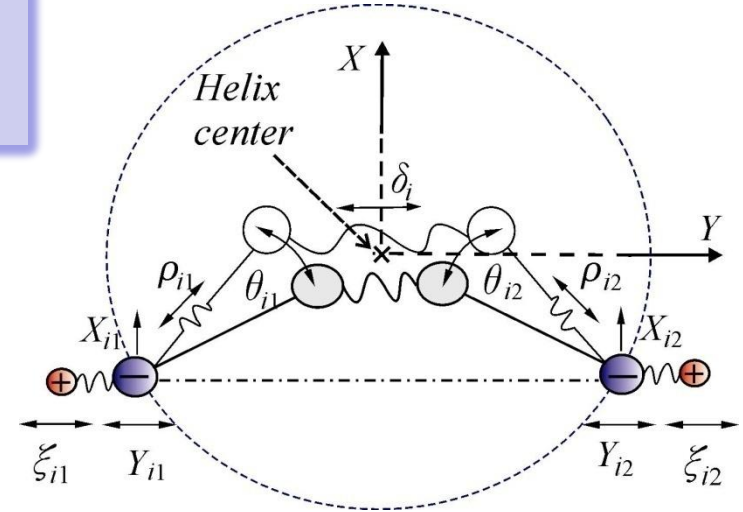
$$E_i^h = \sum_{j=1}^2 (K_{ij} + U_{ij}).$$

Kinetic energy of  $i$ -th link:

$$K_i = \frac{1}{2} \sum_{j=1}^2 \left[ M (\dot{X}_{ij}^2 + \dot{Y}_{ij}^2) + m (\dot{\rho}_{ij}^2 + l^2 \dot{\theta}_{ij}^2 + 2la \dot{\theta}_{ij} \dot{Y}_{ij} + 2b \dot{\rho}_{ij} \dot{Y}_{ij} - 2a \dot{X}_{ij} \dot{\rho}_{ij} + 2lb \dot{X}_{ij} \dot{\theta}_{ij}) \right].$$

Potential energy:

$$U_i = \frac{1}{2} \alpha \delta_i^2 + \frac{1}{2} \sum_{j=1}^2 (\sigma \rho_{ij}^2 + \beta \theta_{ij}^2).$$



# Energy of counterion vibrations

Single-stranded  
neutralization:

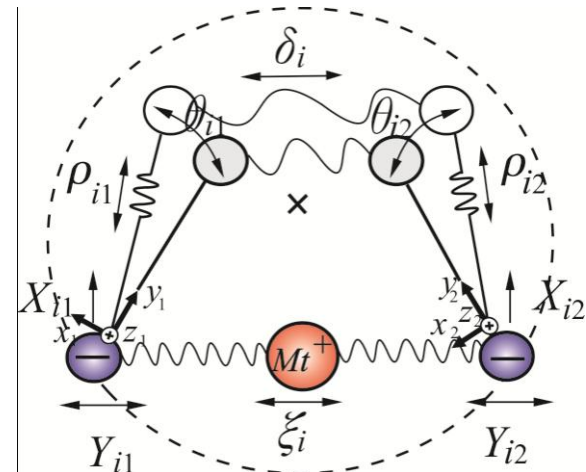
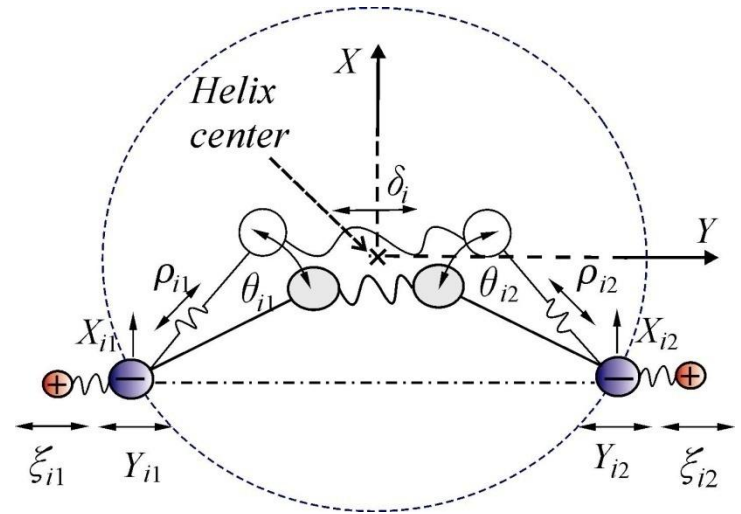
$$E_i^{\text{sn}} = \frac{1}{2} \sum_{j=1}^2 \left[ m_a (\dot{\xi}_{ij} + \dot{Y}_{ij})^2 + \gamma \xi_{ij}^2 \right].$$

Cross-stranded  
neutralization:

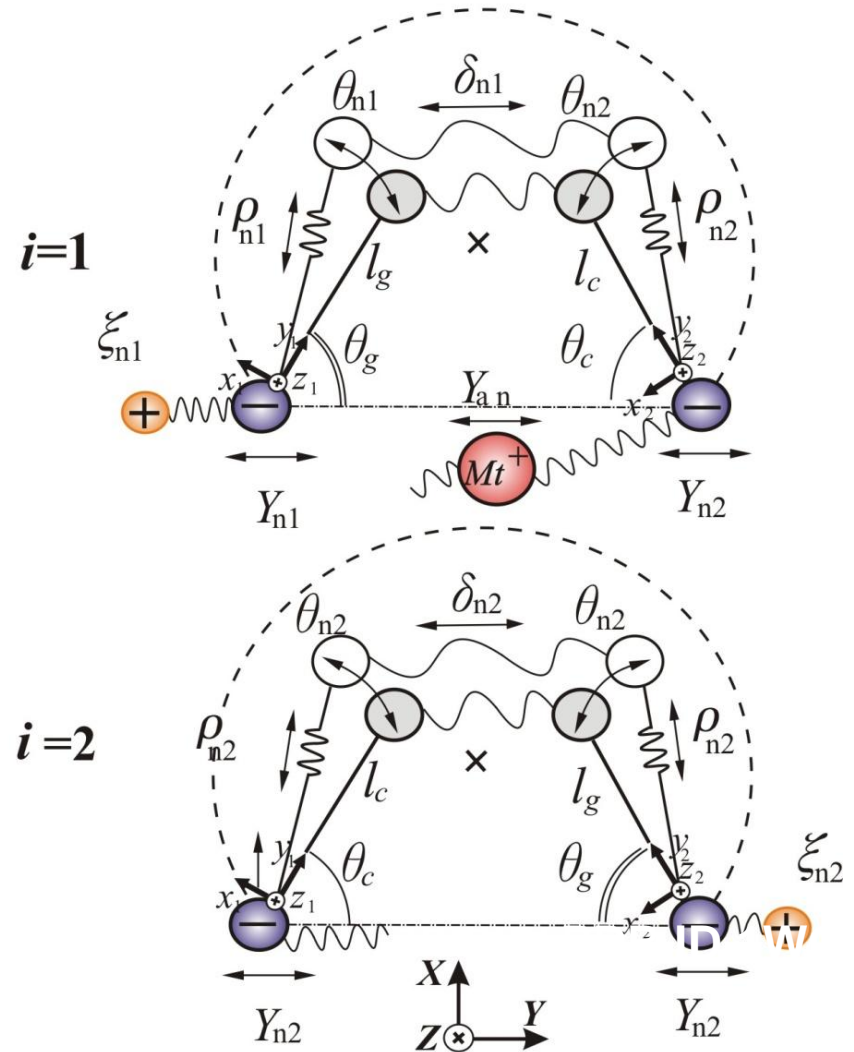
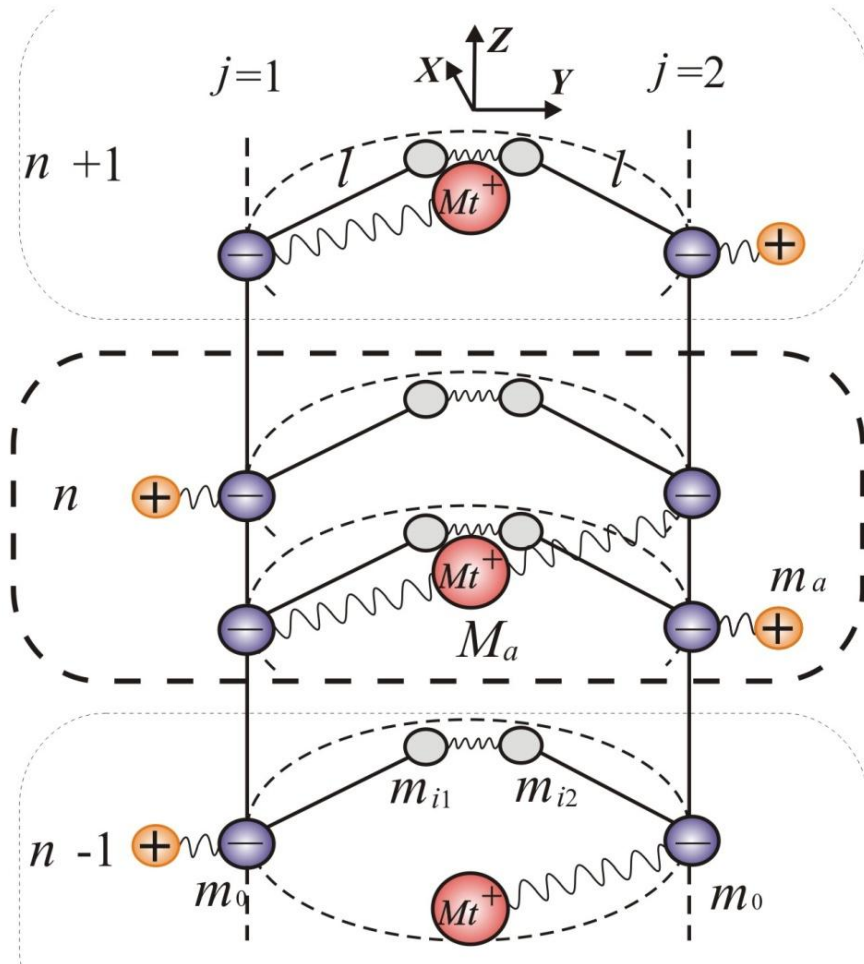
$$E_i^{\text{cn}} = \frac{1}{2} \sum_{j=1}^2 \left[ m_a \dot{\xi}^2 + \gamma (Y_{ij} + (-1)^j \xi_i)^2 \right].$$

Interaction along the helix:

$$E_{i,i+1} = U(X', Y', \theta', \rho', \xi').$$



# Model for the left-handed Z-DNA





# Vibrational energy for Z-DNA

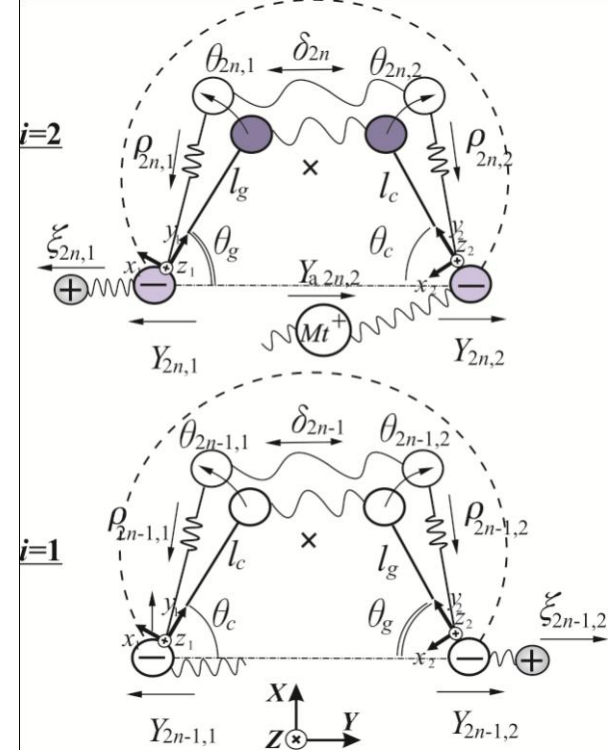
Kinetic energy

Potential energy

$$E = K + U;$$

$$K = \sum_n \left[ \sum_{j=1}^2 (K_{2n,j}^h + K_{2n-1,j}^h) + K_{2n}^I + K_{2n-1}^I \right];$$

$$U = \sum_n (U_{2n} + U_{2n-1} + U_{2n,2n-1} + U_{2n}^{Ion} + U_{2n-1}^{Ion}).$$



$$K_{2n,j}^h = \frac{1}{2} \left[ M_{2nj} \dot{X}_{2nj}^2 + m_{2nj} (\dot{\rho}_{2nj}^2 + l_{2nj}^2 \dot{\theta}_{2nj}^2 + 2b_{2nj} \dot{X}_{2nj} \dot{\rho}_{2nj} + 2l_{s2nj} \dot{X}_{2nj} \dot{\theta}_{2nj}) \right];$$

$$K_{2n}^I = M_a \dot{X}_{a2n}^2 + m_a (\dot{X}_{2n1} + \dot{\xi}_{2n1})^2; \quad K_{2n-1}^I = m_a (\dot{X}_{2n-1,2} + \dot{\xi}_{2n-1,2})^2.$$

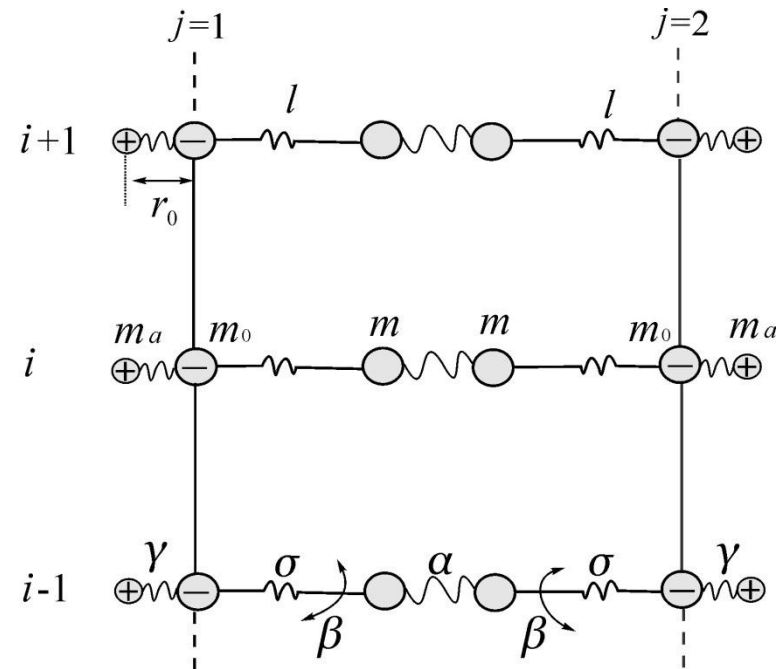
$$U_{2n} = \frac{1}{2} \left[ \alpha (X_{2n1} + X_{2n2} + l_{s2n1} \theta_{2n1} + l_{s2n2} \theta_{2n2} + b_{2n1} \rho_{2n1} + b_{2n2} \rho_{2n2})^2 + \sum_{j=1}^2 (\beta_{2nj} \theta_{2nj}^2 + \sigma_{2nj} \rho_{2nj}^2) \right];$$

$$U_n^I = \frac{\gamma_1}{2} [(X_{a2n} - X_{2n2})^2 + (X_{a2n} + X_{2n-1,1})^2] + \frac{\gamma_2}{2} (\xi_{2n1}^2 + \xi_{2n-1,2}^2).$$

# Parameters of the models

## Structure parameters

Nucleoside	$l$ (Å)	$\theta_{eq}$ (°)	$m$ (a. o. M.)
Adenosine	5,3	28	203
Thymidine	4,8	32	194
Guanosine	5,5	23	219
Cytosine	4,7	30	179
Average	4,9	27,5	199



## Force constants

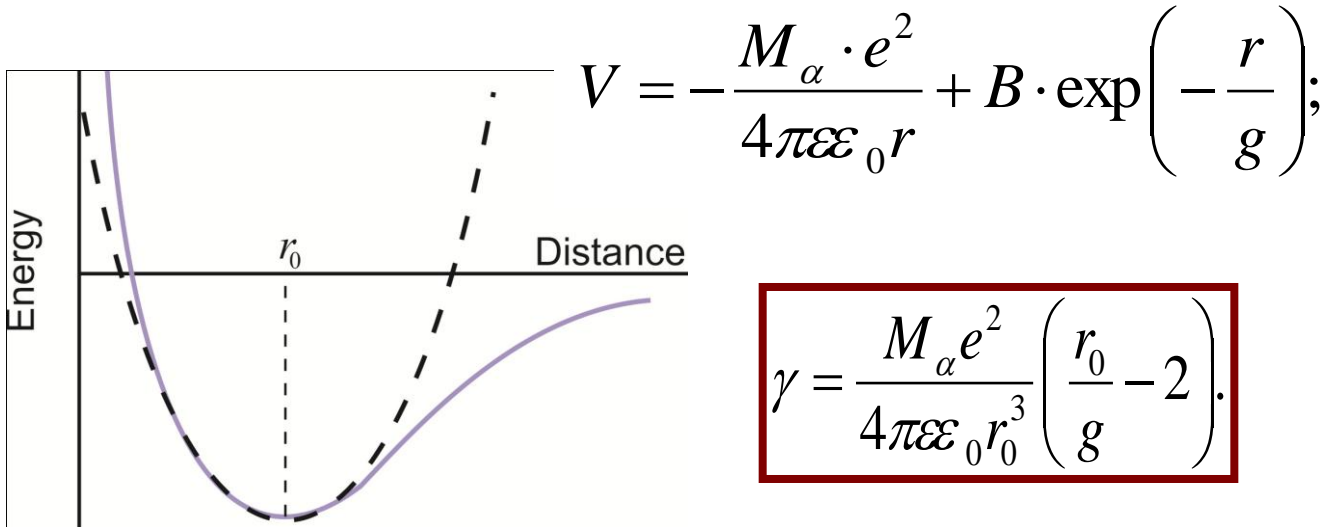
DNA form	$\alpha$ (kcal/molÅ <sup>2</sup> )	$\sigma$ (kcal/molÅ <sup>2</sup> )	$\beta$ (kcal/molÅ <sup>2</sup> )	$\gamma$ (kcal/molÅ <sup>2</sup> )
B-form	80 ±5	43 ±5	40 ±8	?
A-form	18	22	46	

Volkov S.N., Kosevich A.M., Wainreb G.E. Biopolimery i Kletka. 5, 32-39 (1989).

Volkov S.N., Kosevich A.M. J. Biomolec. Struct. Dyn., 8, 1069 (1990).

# Constant of ion-phosphate vibrations

Potential with the Born-Mayer repulsion:



**Values:**

$$\gamma = 30 \div 60$$

(kcal/molÅ<sup>2</sup>)

**The Madelung constant:**

$$M_{\alpha} = r_0 \sum_{i,j} \left( \frac{1}{r_{ij}} - \frac{1}{d_{ij}} \right) + \frac{r_0}{d_{i1}}$$

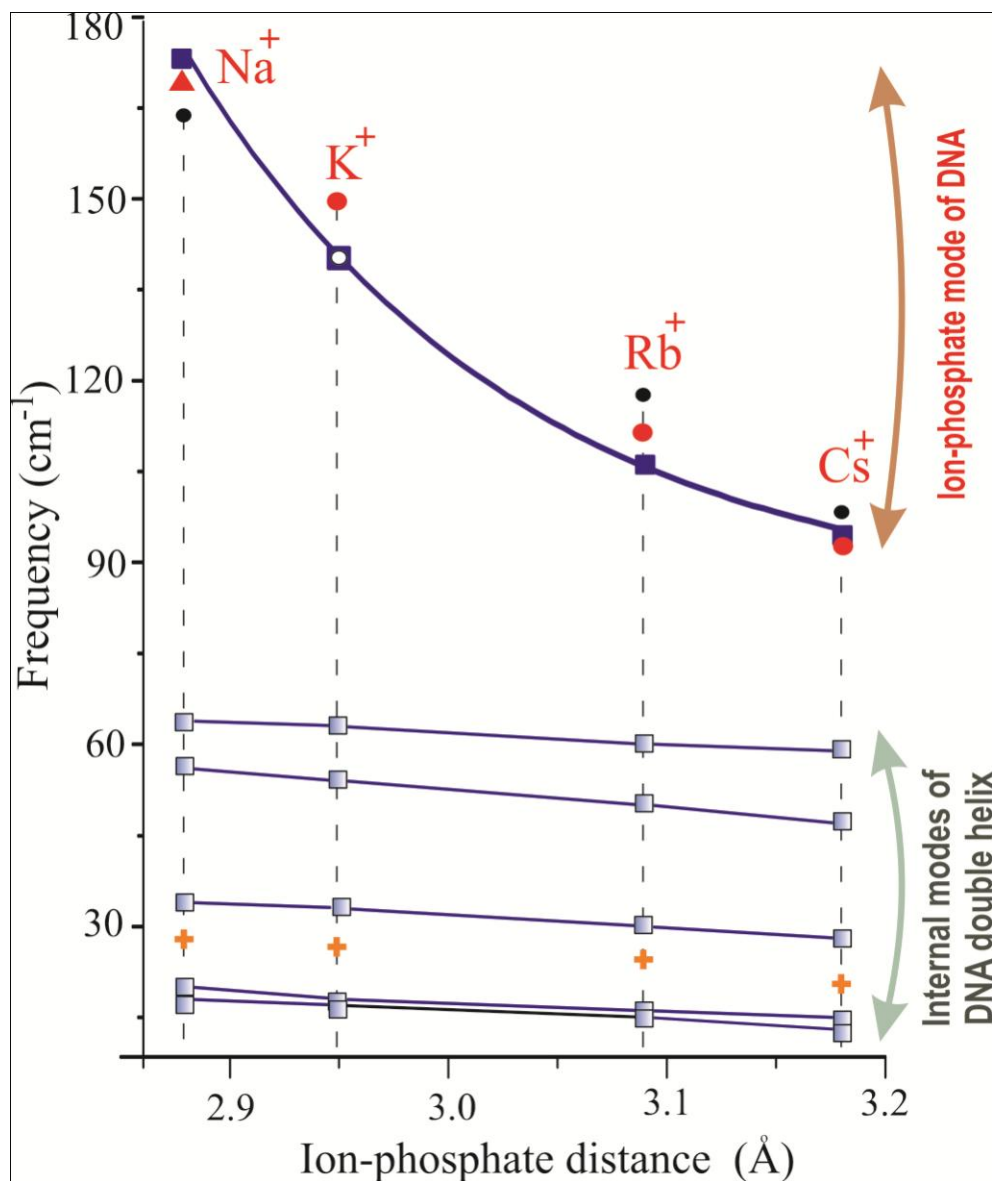
$$M_{\alpha} \approx 1 \div 1,3$$

**Dielectric constant:**

$$\epsilon = n^2 \frac{M_{\alpha} (r_0 / g - 2)}{M_{\alpha} (r_0 / g - 2) - 8\pi r_0^3 / V}.$$

$$\epsilon = 2.3 \div 2.6$$

# Ion-phosphate vibrations in the low-frequency spectra of DNA



Atomic weight of metals (a.u.m.)

Na <sup>+</sup>	K <sup>+</sup>	Rb <sup>+</sup>	Cs <sup>+</sup>
23	39	86	133

## Theory

**Perepelytsya S.M., Volkov S.N.**

*Ukr. J. Phys.*, 49, 1072 (2004).

*Eur. Phys. J. E.* 24, 261 (2007).

## Experiment

**Wittlin A. et al.** *Phys. Rev. A*, 34, 493 (1986).

**Weidlich T. et al.**

*Biopolymers*, 30, 477 (1990).

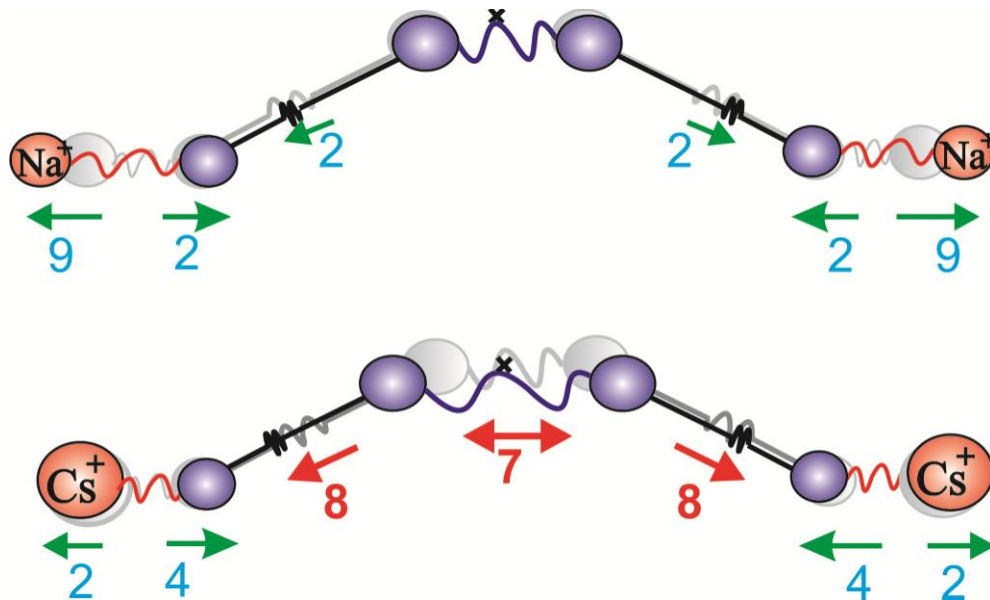
**Powell J. W. et al.**

*Phys. Rev. A*, 35, 3929 (1987).

In the IR low-frequency spectra of DNA the modes of ion-phosphate vibrations have been found.

# Influence of heavy counterions

Scheme of structural motions in nucleotide pair  
(amplitudes in pm)



Na-DNA  
182 cm<sup>-1</sup>

Cs-DNA  
118 cm<sup>-1</sup>

*Perepelytsya S.M., Volkov S.N. Eur. Phys. J. E (2007)*

# Intensities in the Raman low-frequency spectra of DNA

**Semiclassical approach:**

Volkenshtein M.V., Eliashevich M.A., Stepanov B.I., *Vibrations of Molecules. Volume 2.* (Moscow, 1949).

**Semiclassical approach**

**+**

**Model of DNA conformational vibrations with counterions**

$$J_n \approx \frac{3\kappa J_0 (\nu_0 - \nu_n)^4}{1 - \exp(-h\nu_n/kT)} \left[ \left( \sum_{j=1}^2 A_j^n \right)^2 + \left( \sum_{j=1}^2 B_j^n \right)^2 \right]$$

$$A_j^n = \left[ (b_{jyy} - b_{jxx}) \sin 2\theta_{eq} + 2(-1)^j b_{jxy} \cos 2\theta_{eq} \right] \cdot \left( \tilde{\theta}_j^n + \frac{\tilde{\rho}_j^n}{l} \right),$$

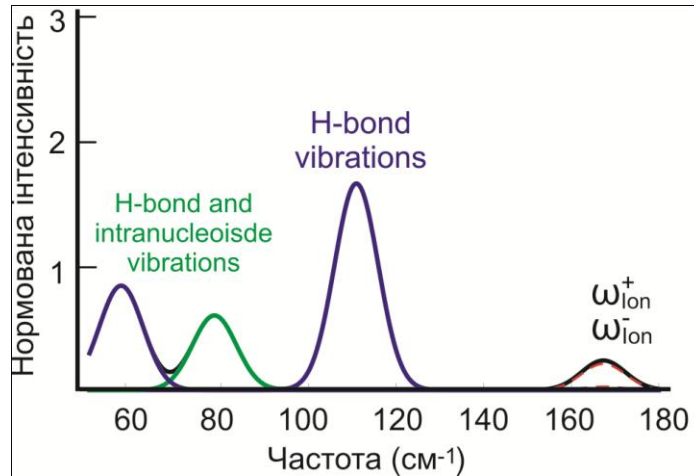
$$B_j^n = \left[ (b_{jyy} - b_{jxx}) (-1)^j \cos 2\theta_{eq} - 2b_{jxy} \sin 2\theta_{eq} \right] \cdot \left( \tilde{\theta}_j^n + \frac{\tilde{\rho}_j^n}{l} \right).$$

**Perepelytsya S.M., Volkov S.N. *Biophys.Bull.* **23(2)**, 5 (2009).**

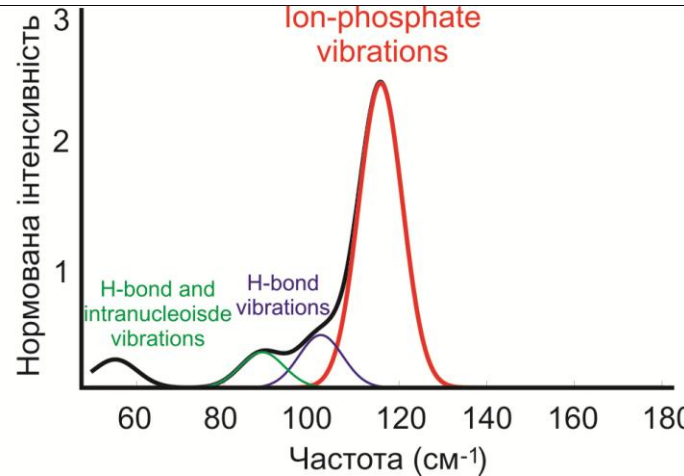
**Perepelytsya S.M., Volkov S.N. *Eur. Phys. J. E.* **31**, 201 (2010).**

# Observation of DNA ion-phosphate vibrations

## Na-DNA



## Cs -DNA



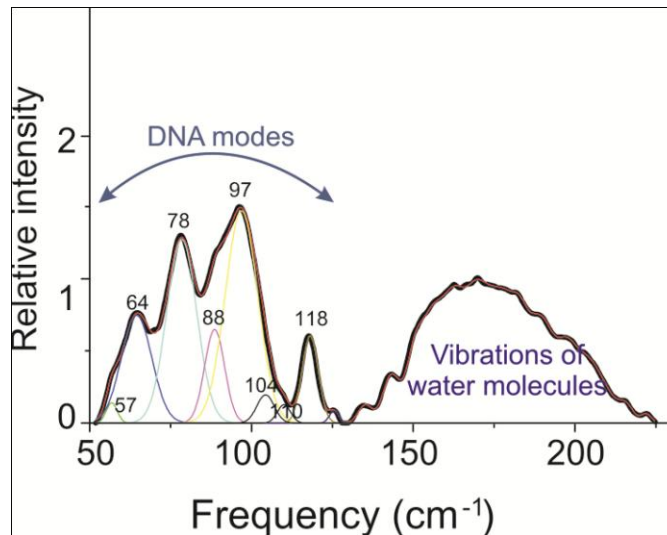
## Theory

Approach for calculation of intensities in DNA low-frequency Raman spectra.

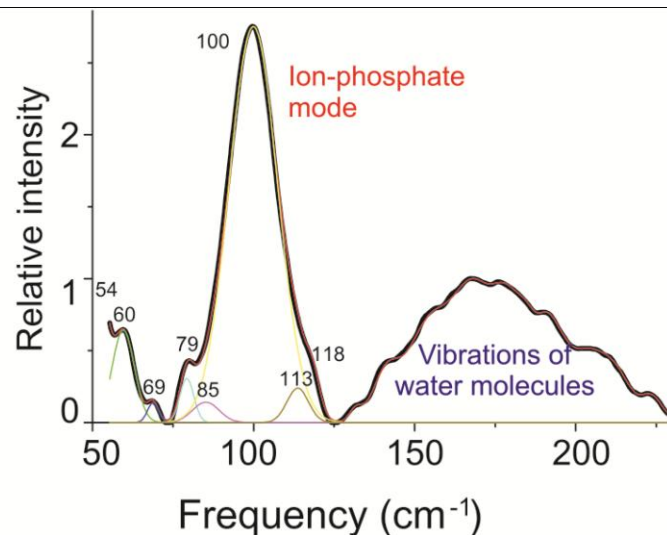
**Perepelytsya S.M., Volkov S.N.**

Eur. Phys. J. E. **31**, 201 (2010).  
Biophys. Bull. (2009).

## Na-DNA



## Cs -DNA

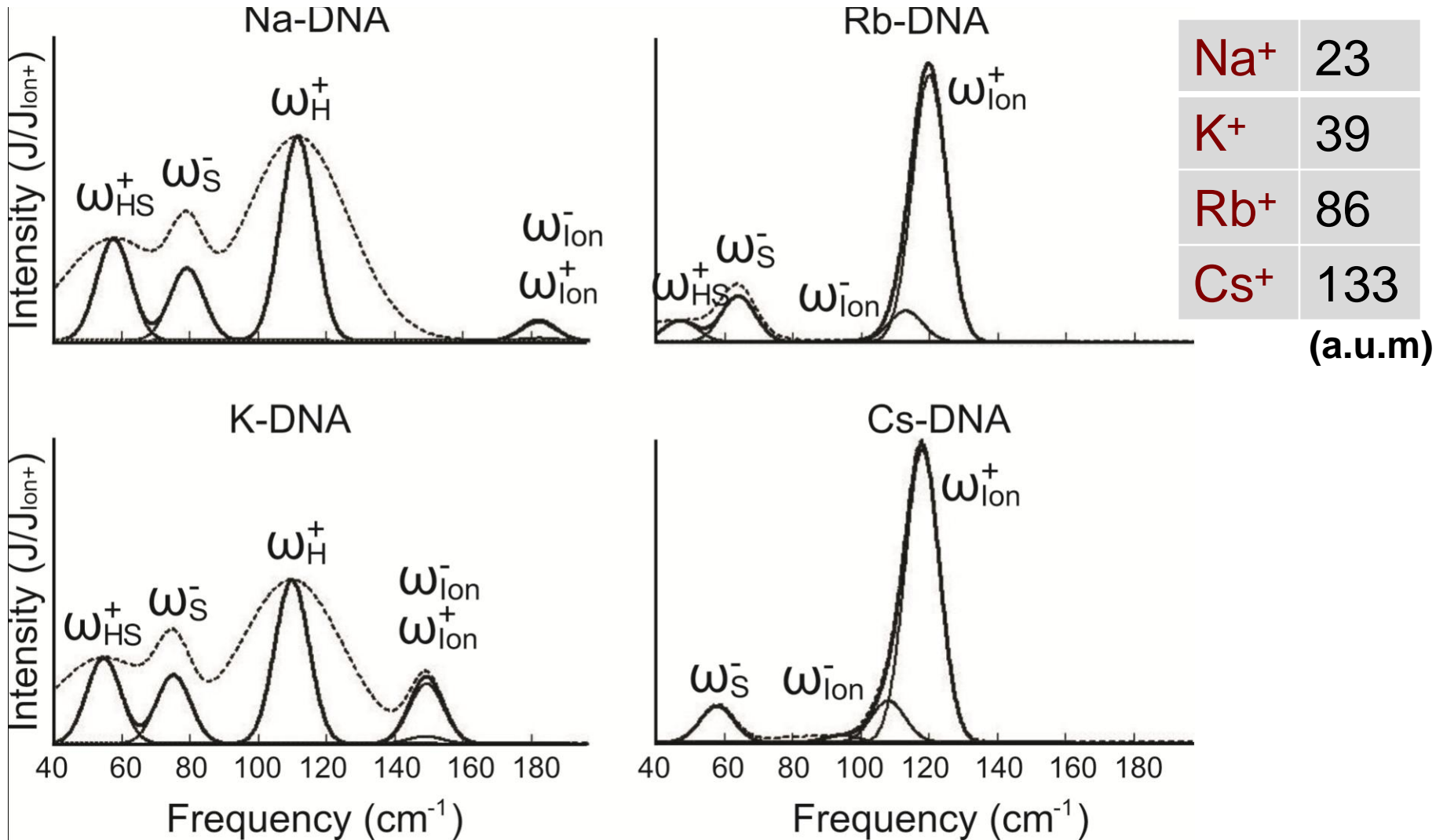


## Experiment

Raman spectra of DNA water solutions with Na<sup>+</sup> and Cs<sup>+</sup> counterions

**Bulavin L.A., et al.**  
*Reports of NASU*, **9** (2007).  
arXiv:0805.0696v1

# The calculated Raman spectra of *B*-DNA with alkali metal counterions

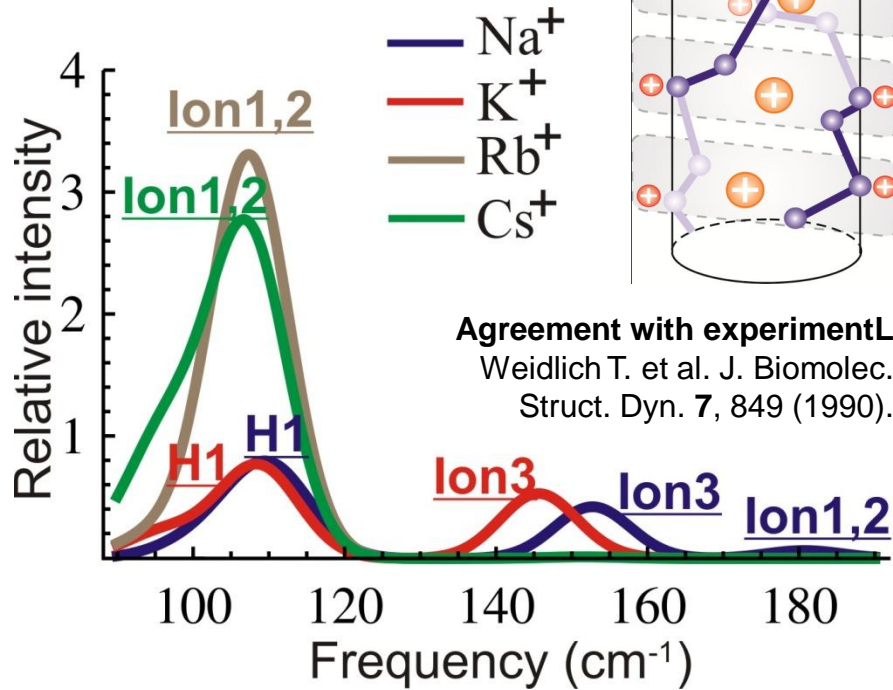




# Comparison of Z- and B-DNA low-frequency spectra

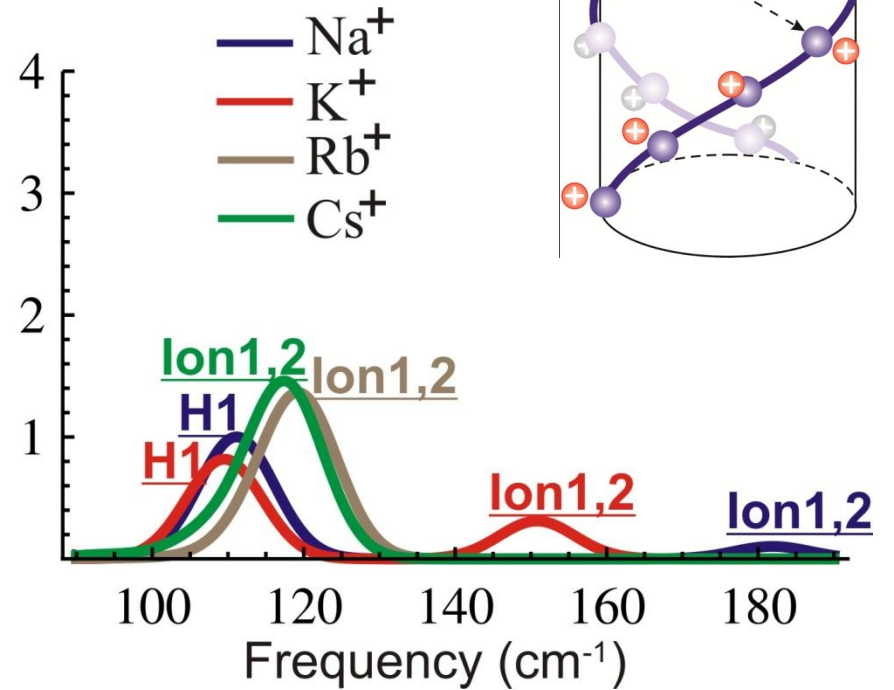
## Z-DNA

S.M. Perepelytsya, S.N. Volkov,  
*Ukr. J. Phys.* **58**, 554 (2013).  
*J. Phys.: C. Ser.* **438**, 012013 (2013).



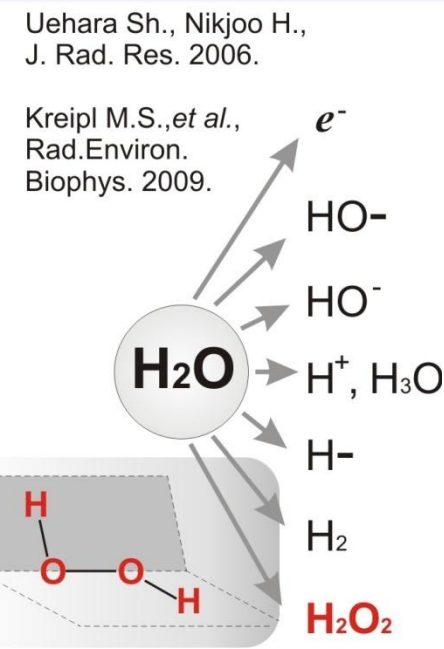
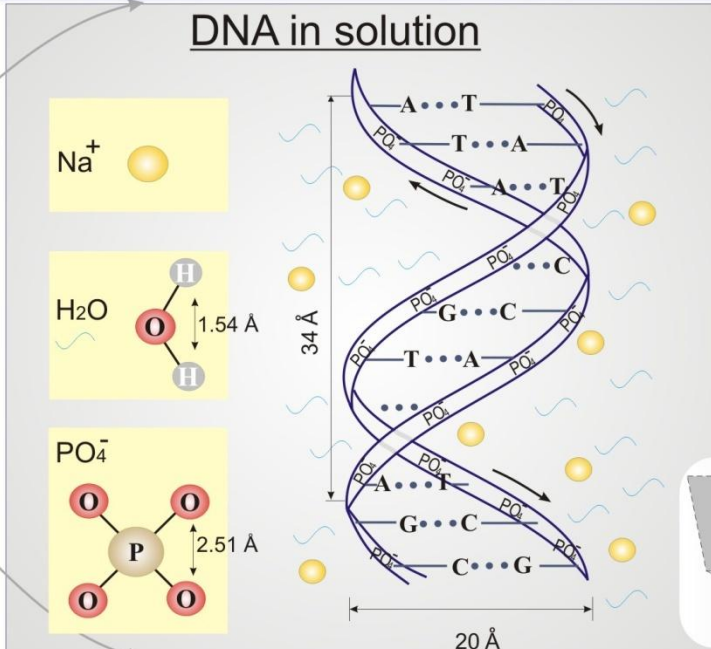
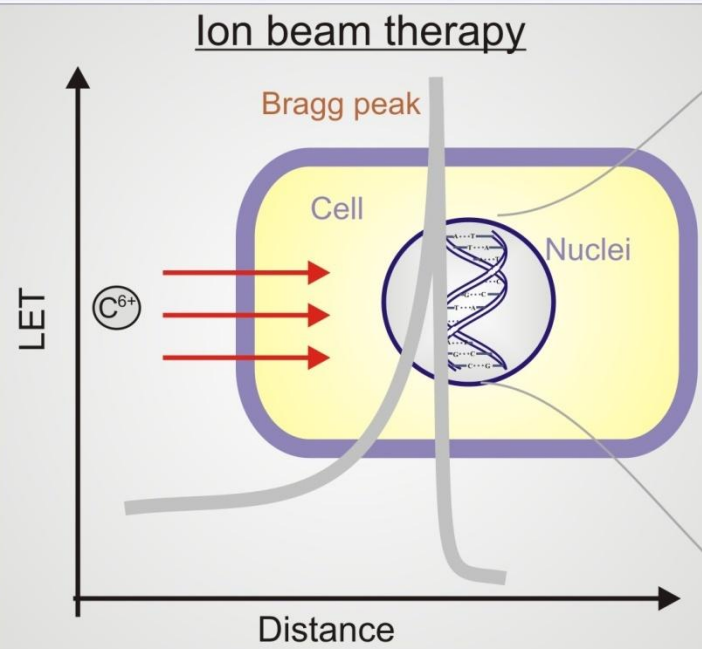
## B-DNA

S.M. Perepelytsya, S.N. Volkov,  
*Eur. Phys. J. E.* **31**, 201 (2010).  
*J. Molec. Liq.* **5**, 113 (2011).



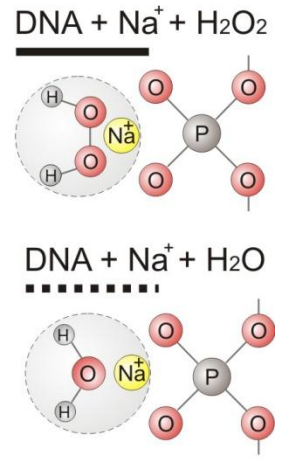
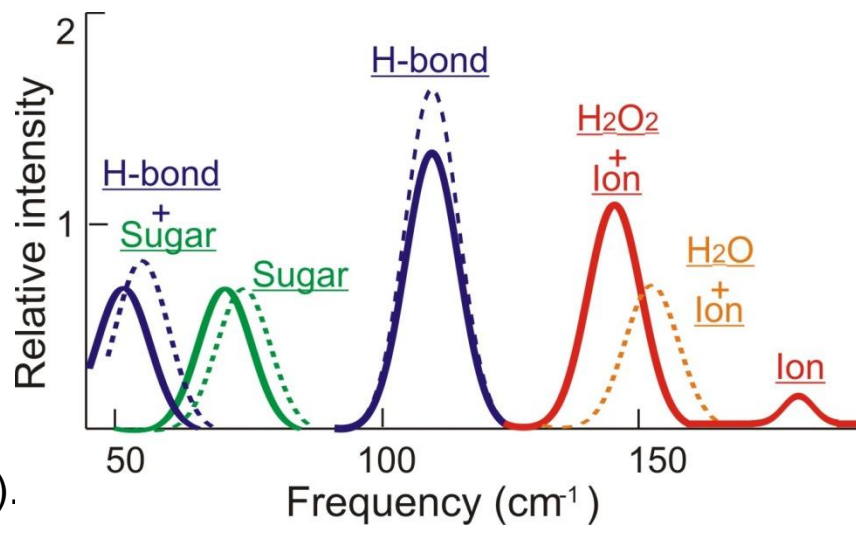
For the left-handed DNA (Z-form) the mode of ion-phosphate vibrations was found near  $150 \text{ cm}^{-1}$  that characterizes the vibrations of  $\text{Mg}^{2+}$  counterions with respect to the phosphate groups in the minor groove of the macromolecule.

# Low-frequency spectra of DNA with H<sub>2</sub>O<sub>2</sub>



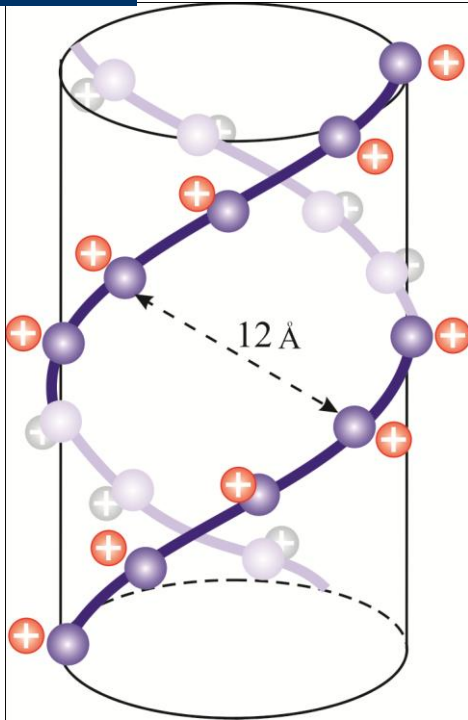
## Low-frequency spectra of DNA with H<sub>2</sub>O<sub>2</sub> and ions

D.V. Piatnytskyi,  
O.O. Zdorevskyyi,  
S.M. Perepelytsya,  
S.N. Volkov,  
Eur. Phys. J. D., **69**, 255 (2015).

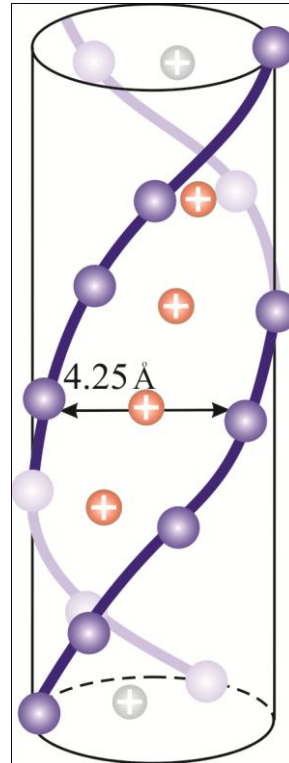


# Spectroscopic manifestations of counterion ordering around DNA double helix

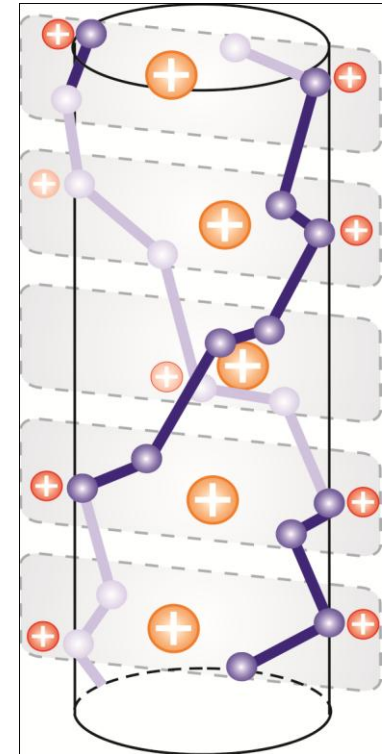
## B-DNA



## C- and D-DNA



## Z-DNA



**S.M. Perepelytsya, S.N. Volkov,**  
*Eur. Phys. J. E.* **24**, 261 (2007).  
*Eur. Phys. J. E.* **31**, 201 (2010).

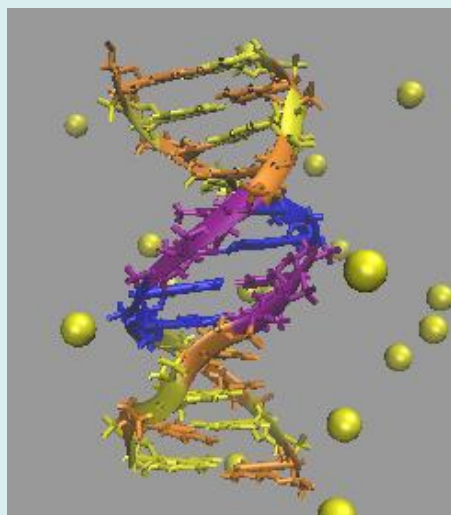
**S.M. Perepelytsya, S.N. Volkov,**  
*Ukr. J. Phys.* **55**, 1182 (2010).  
*J. Molec. Liq.* **5**, 113 (2011).

**S.M. Perepelytsya, S.N. Volkov,**  
*Ukr. J. Phys.* **58**, 554 (2013).  
*J. Phys.: C. Ser.* **438**, 012013 (2013).

Existence of lattice-like ordering of counterions is confirmed by observation of the ion-phosphate vibrations in DNA low-frequency spectra ( $200 \text{ cm}^{-1}$ ).

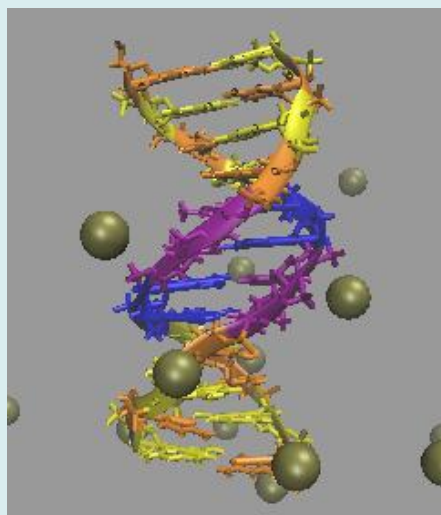
# Modeling of DNA with counterions: Classical MD simulations

## Na-DNA



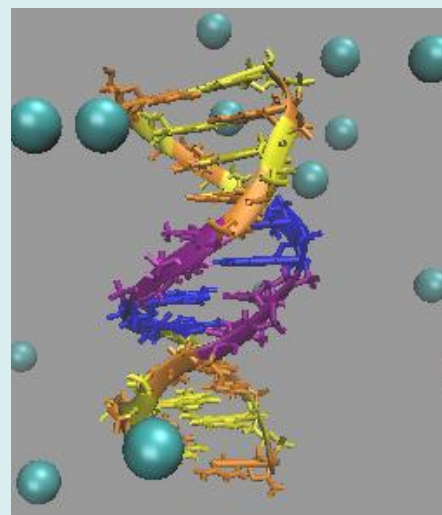
Sodium – 22  
Water – 7912

## K-DNA



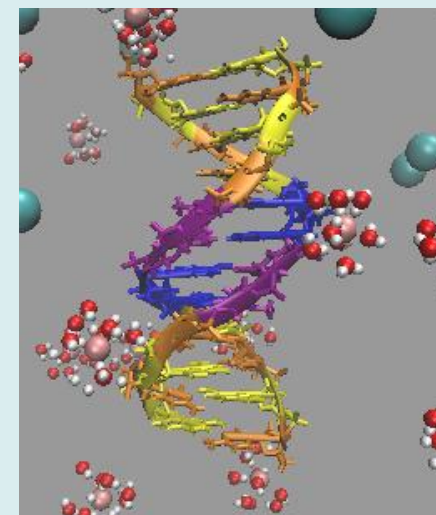
Potassium – 22  
Water – 7912

## Cs-DNA



Cesium – 22  
Water – 7912

## Mg-DNA



Magnesium – 18  
Chlorine – 14  
Water – 7902

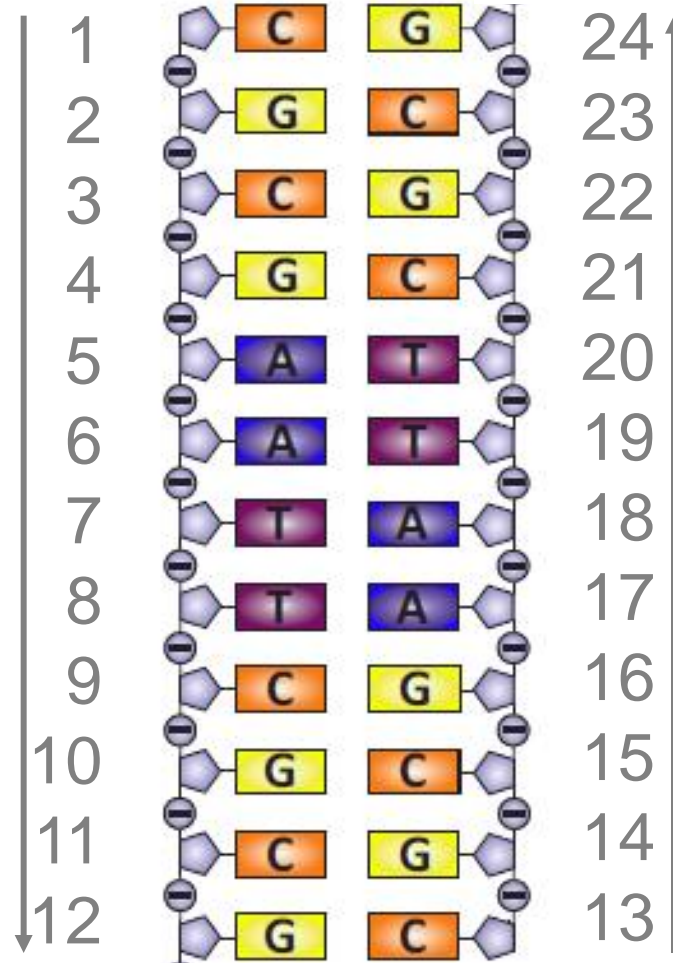
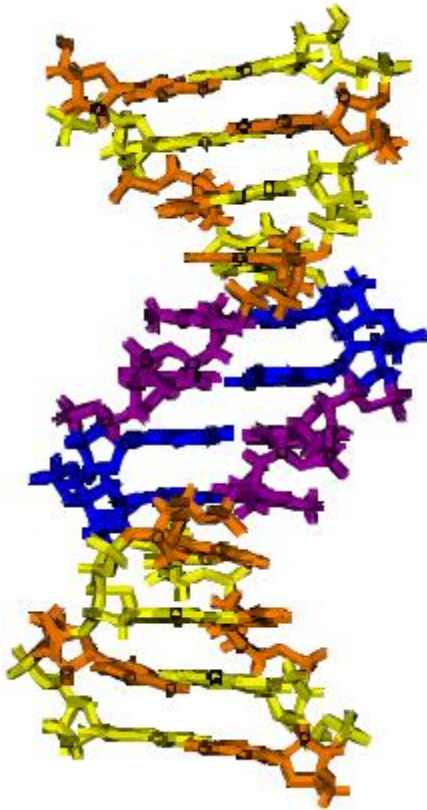
**Box size 64x64x64 Å<sup>3</sup>; simulation trajectory >100 ns.**

**Software packages:**  
**VMD NAMD**  
**Force field: CHARMM27**

Humphrey, W., Dalke, A. and Schulten, K., 'VMD – Visual Molecular Dynamics', *J. Molec. Graphics* 1996, 14.1, 33-38.  
Phillips J.C. *et al.* *J. Comp. Chem.* 26, 1781 (2005).

# Drew-Dickerson dodecamer (CGCGAATTCGCG)

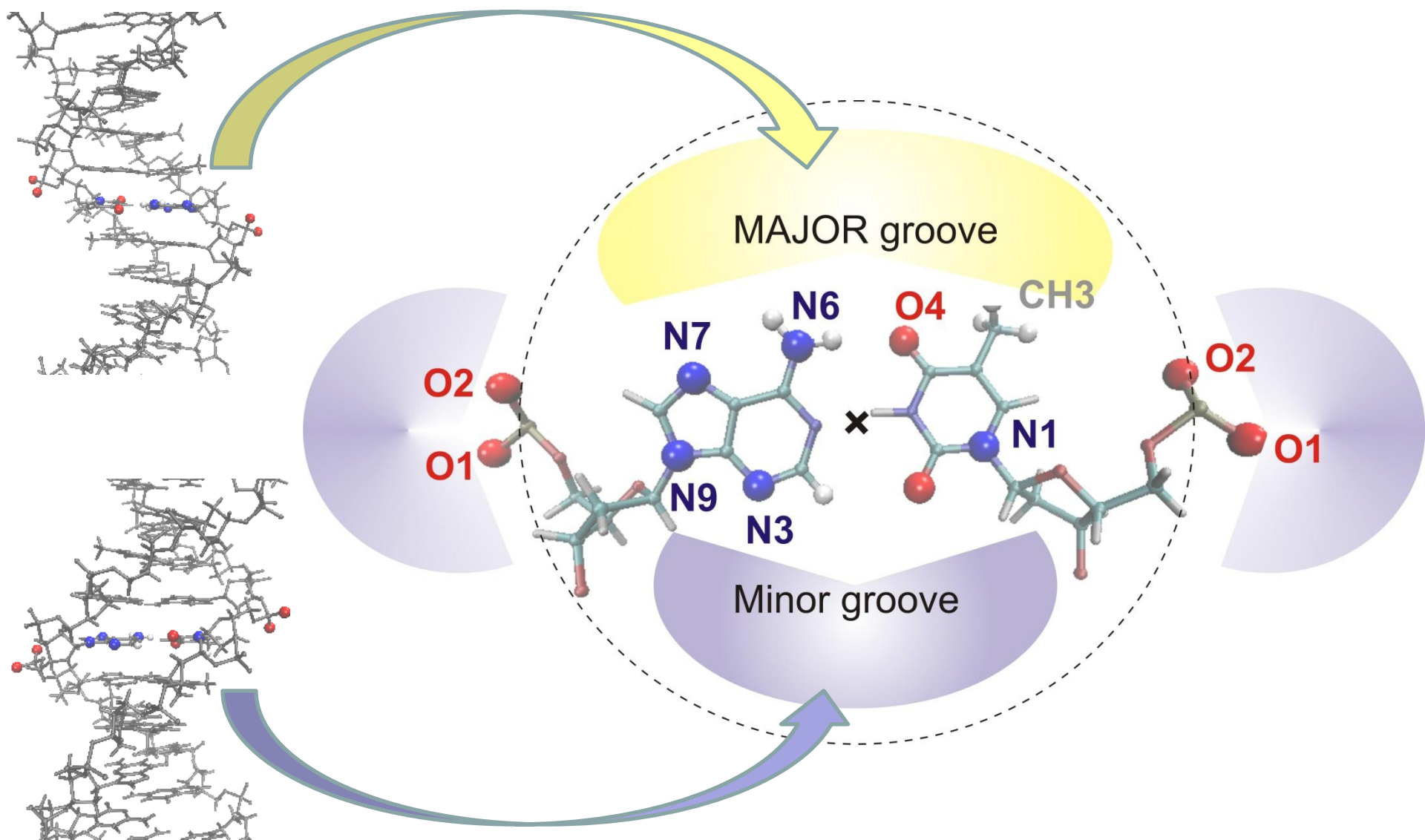
X-ray  
structure  
1BNA



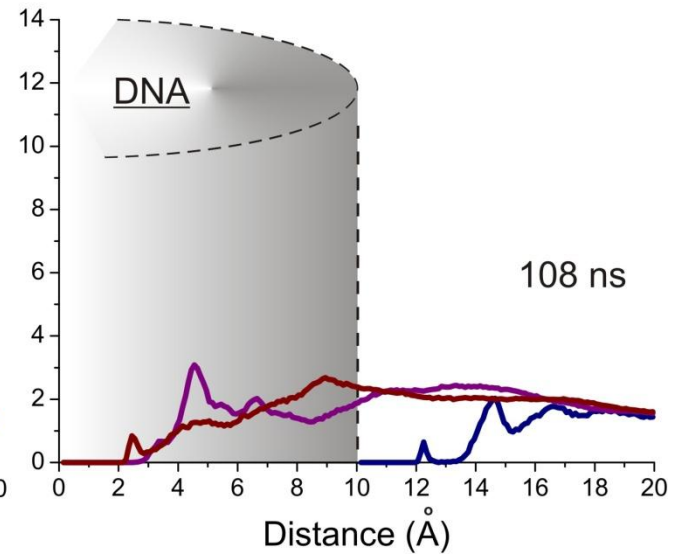
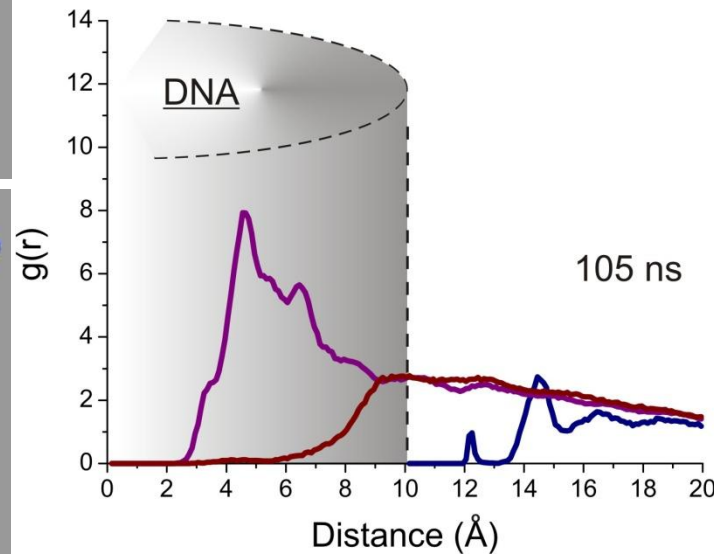
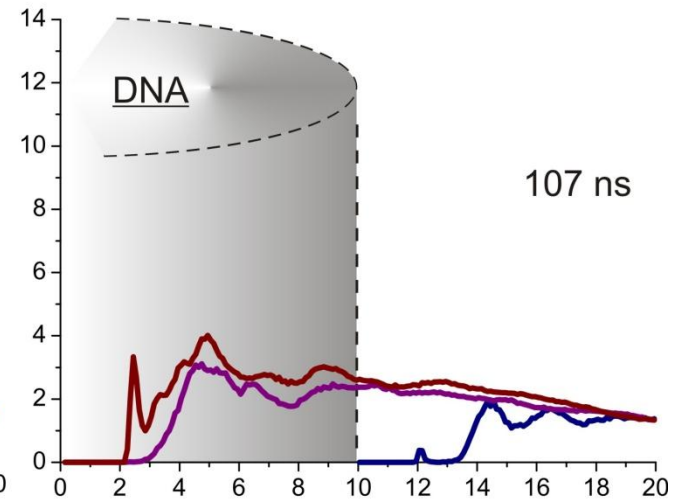
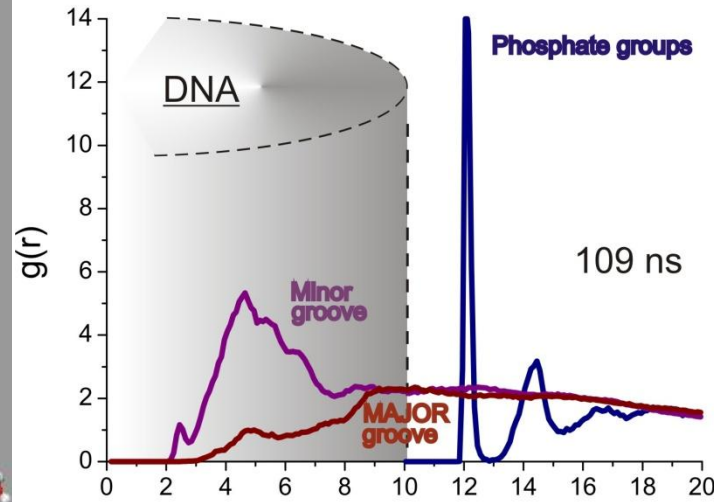
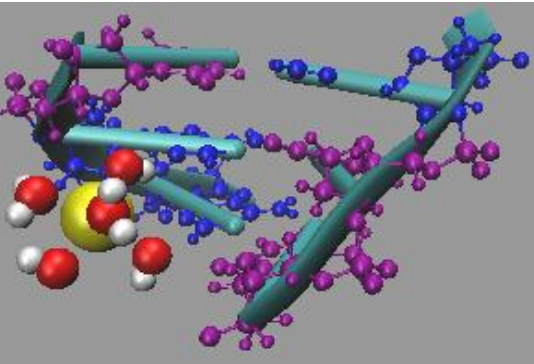
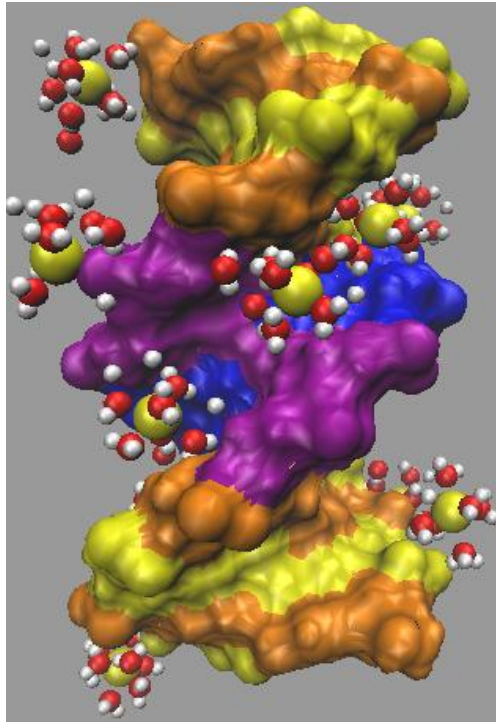
Schematic  
structure

Drew H.R., Wing R.M., Takano T., Broka C., Takana S., Itakura K., Dickerson R.E.,  
*Proc. Natl. Acad. Sci. USA*, **78**,2179-2183 (1981).

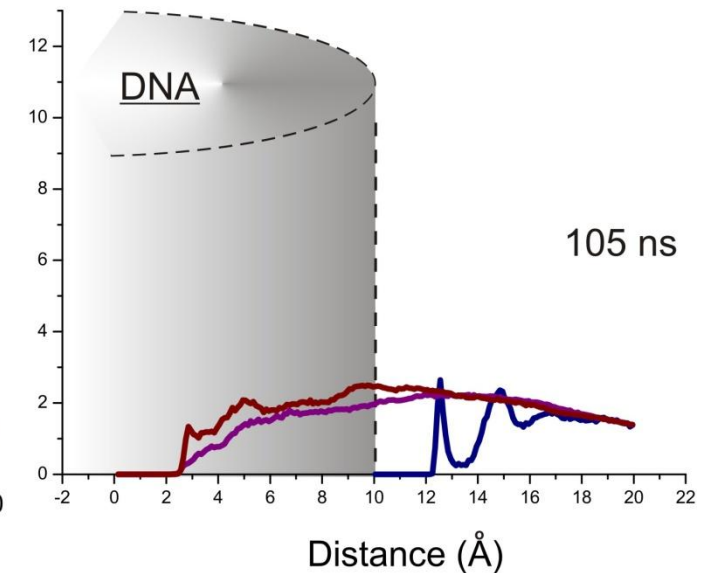
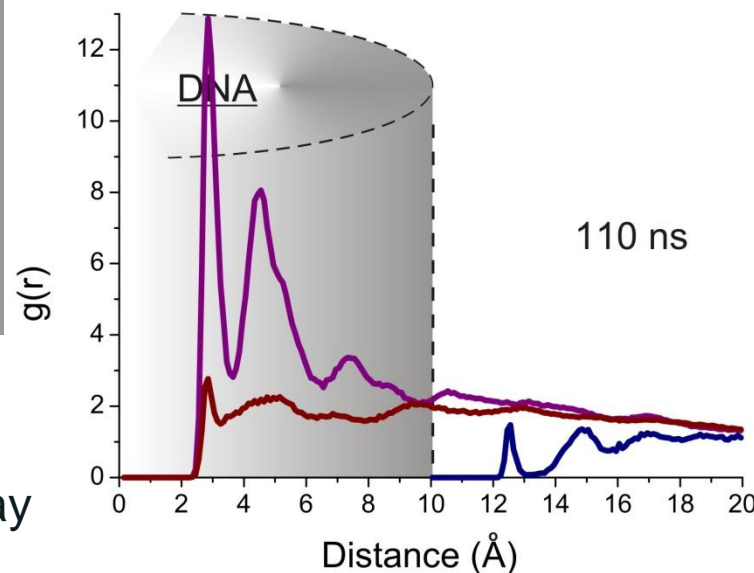
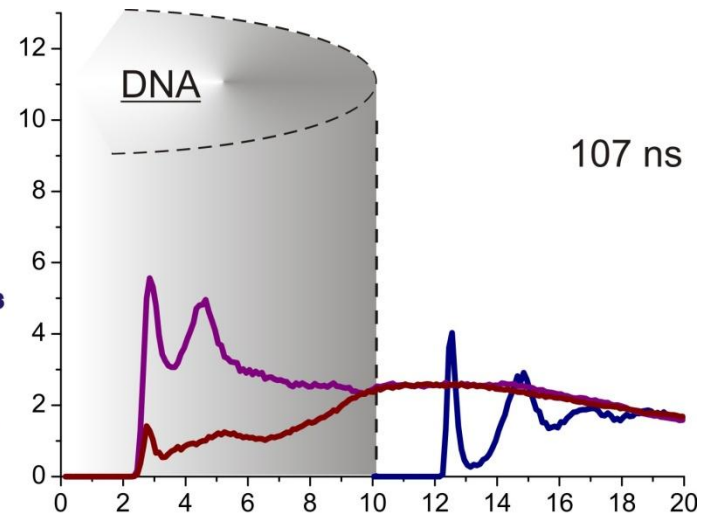
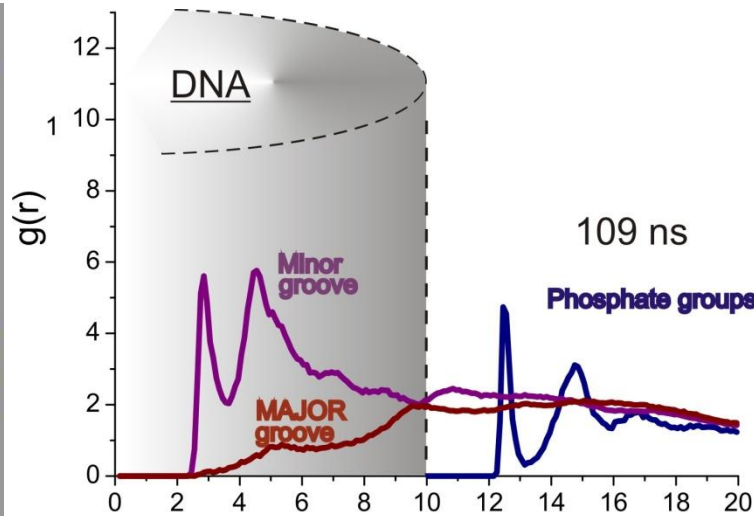
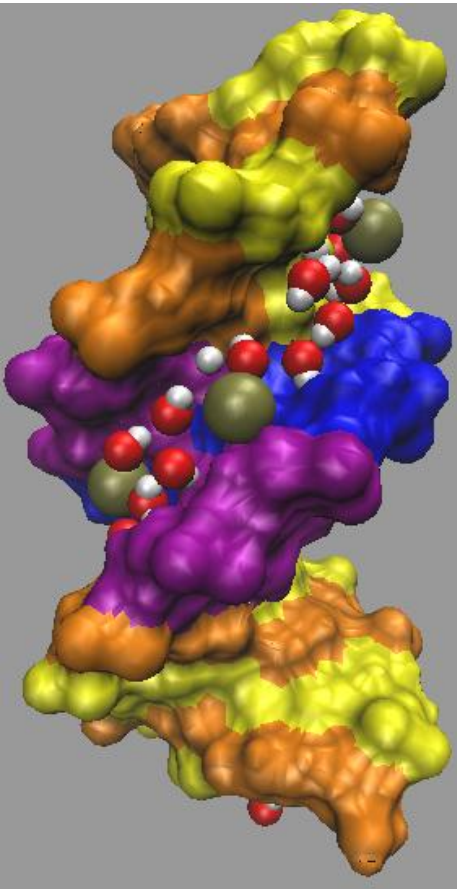
# Radial distribution functions



# Distribution of ions in Na-DNA system



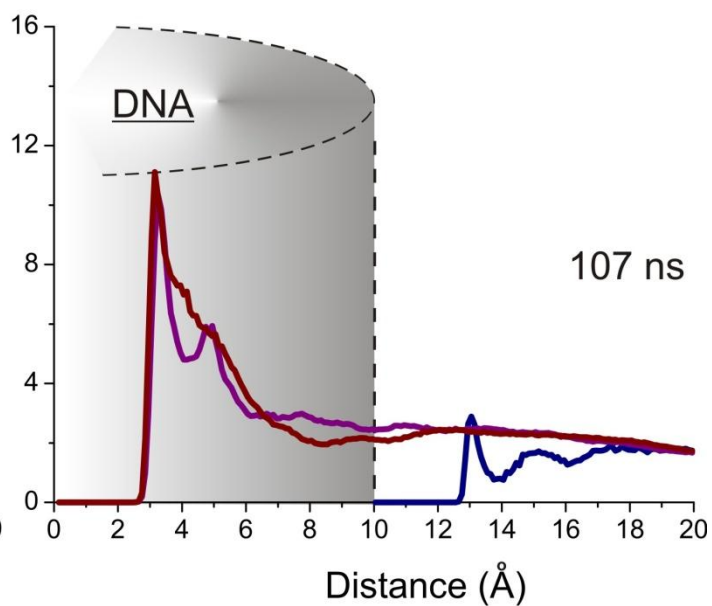
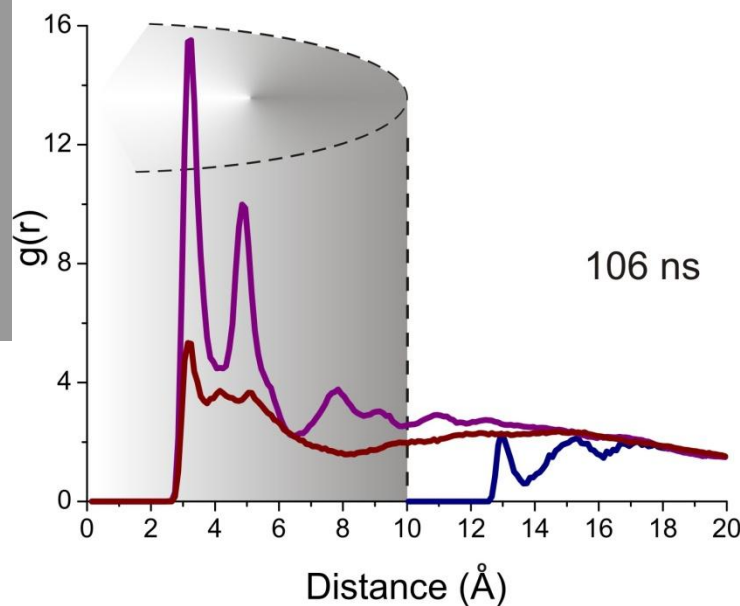
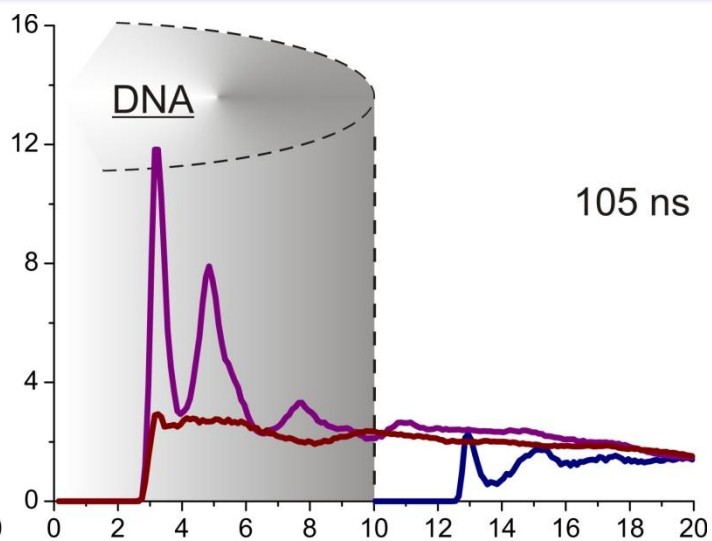
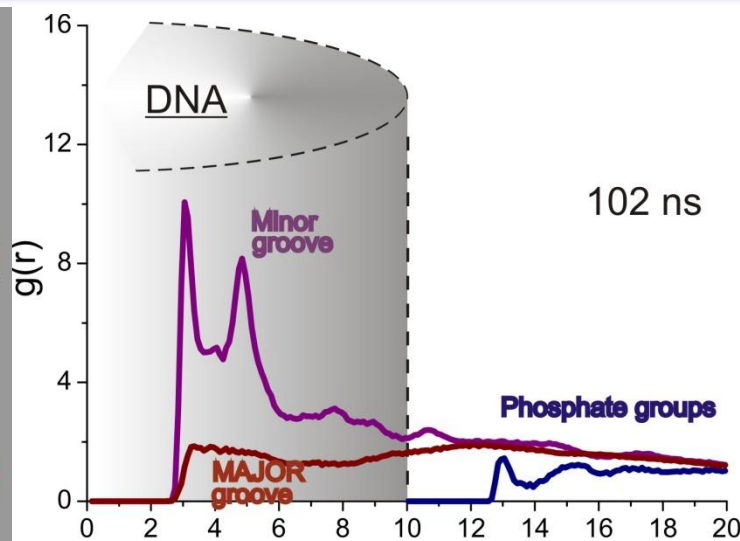
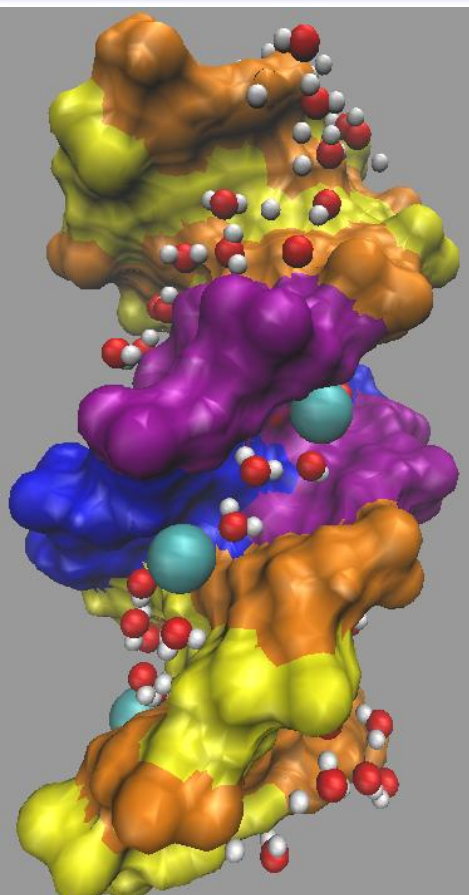
# Distribution of ions in K-DNA system



K<sup>+</sup> in the minor groove. Localization inside the groove may reach about 0.3 ns.

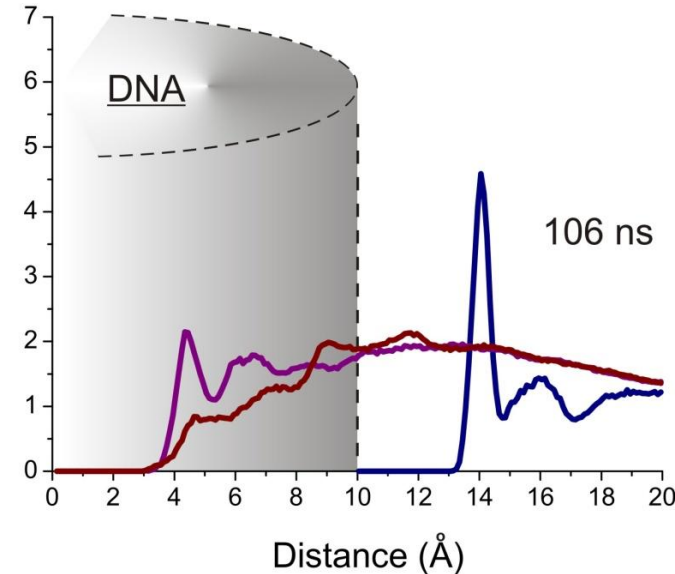
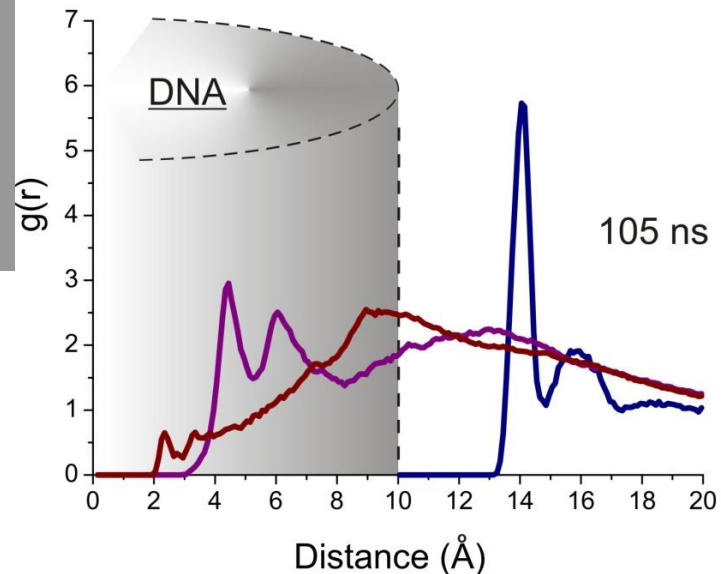
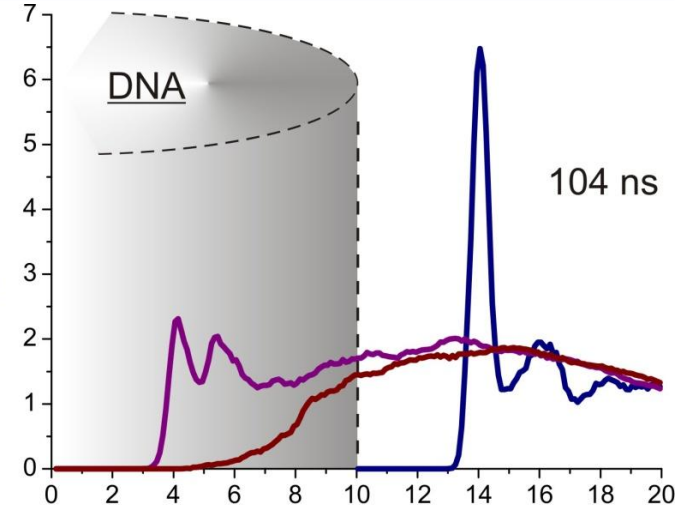
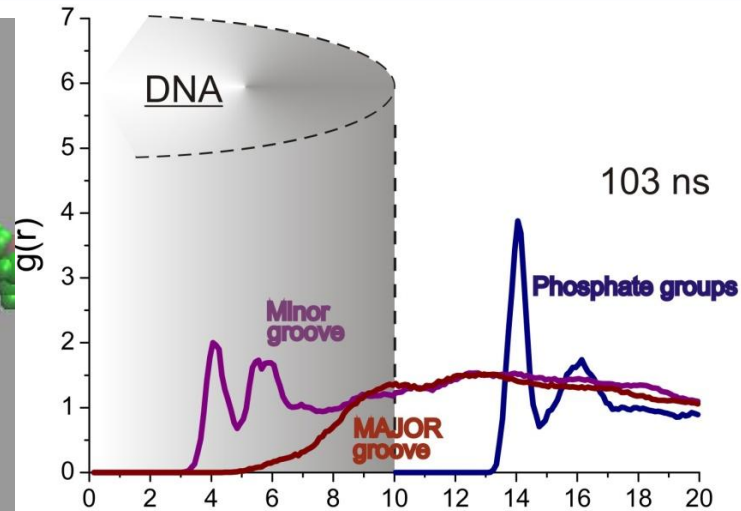
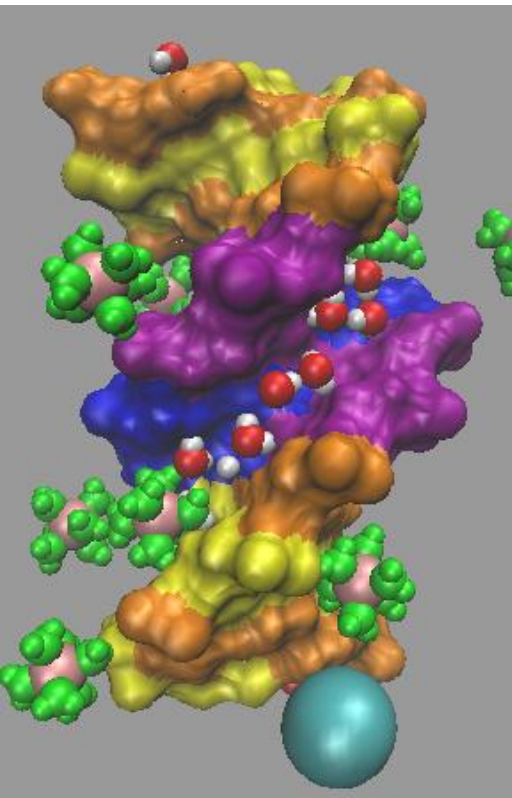


# Distribution of ions in Cs-DNA system



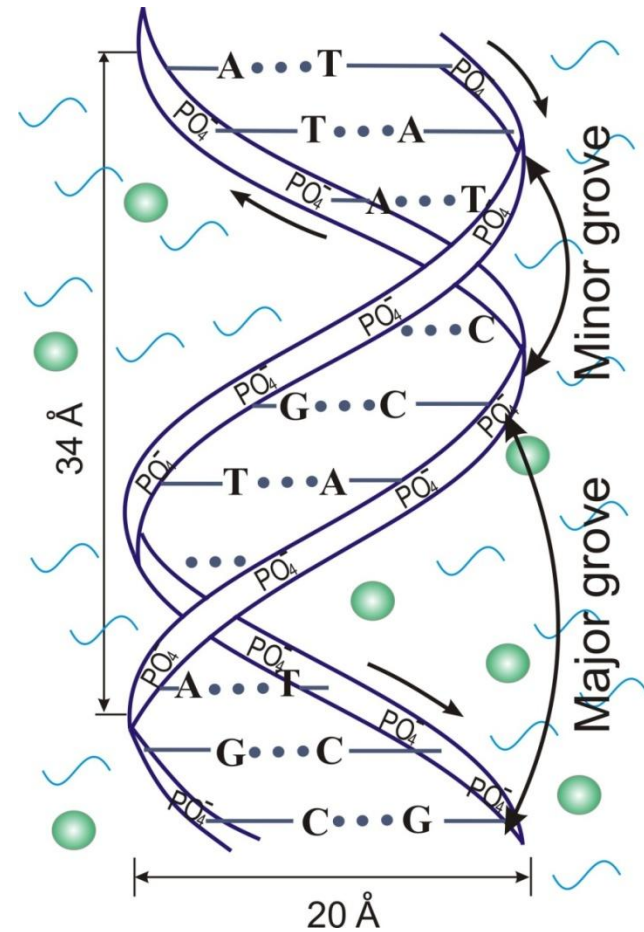
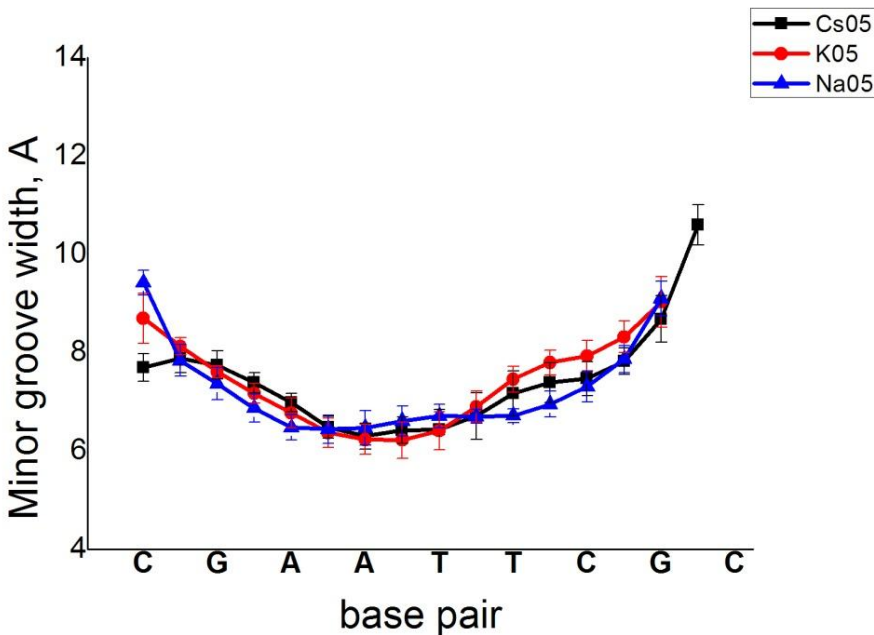
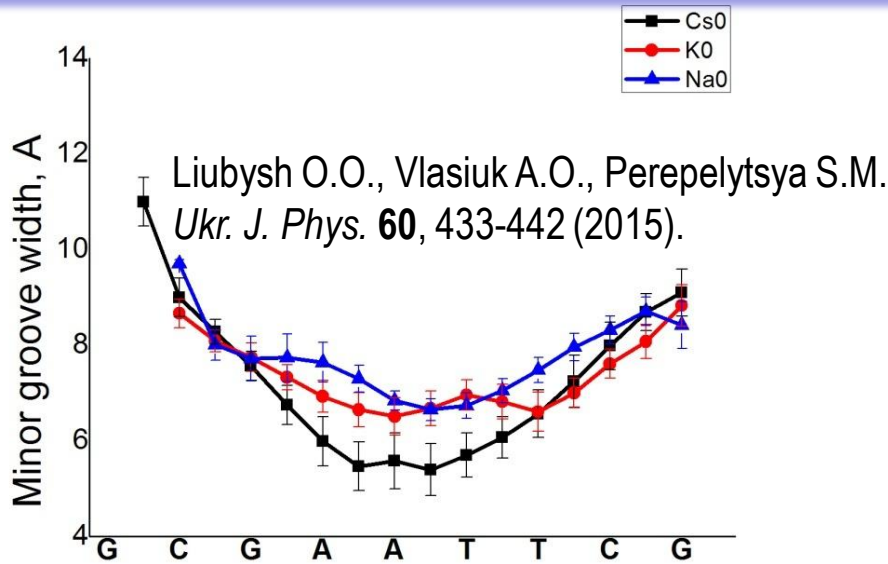
Cs<sup>+</sup> in the minor groove. Localization inside the groove may reach > 1ns.

# Distribution of ions in Mg-DNA system



Hydrated  $Mg^{2+}$  ions are localized in the top edge of the minor groove and near the phosphates ( $t > 1$  ns).

# Width of DNA minor groove

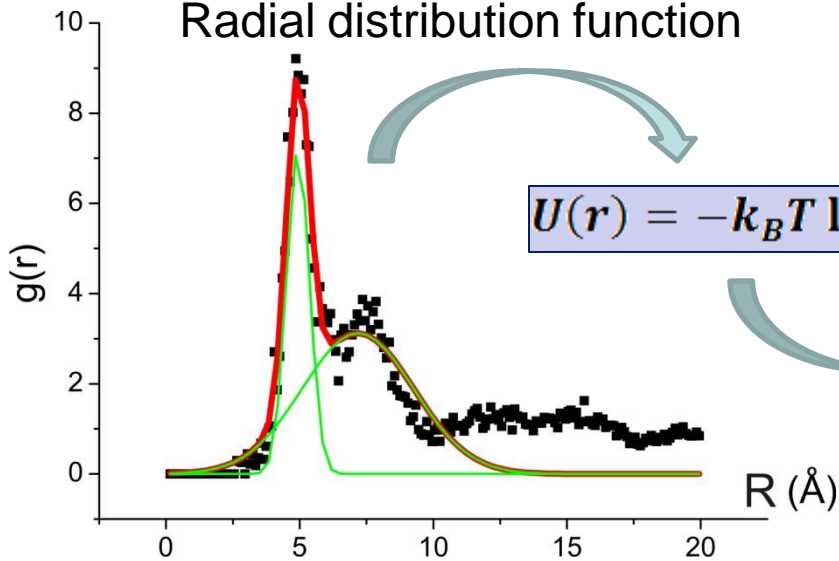


## Curves+

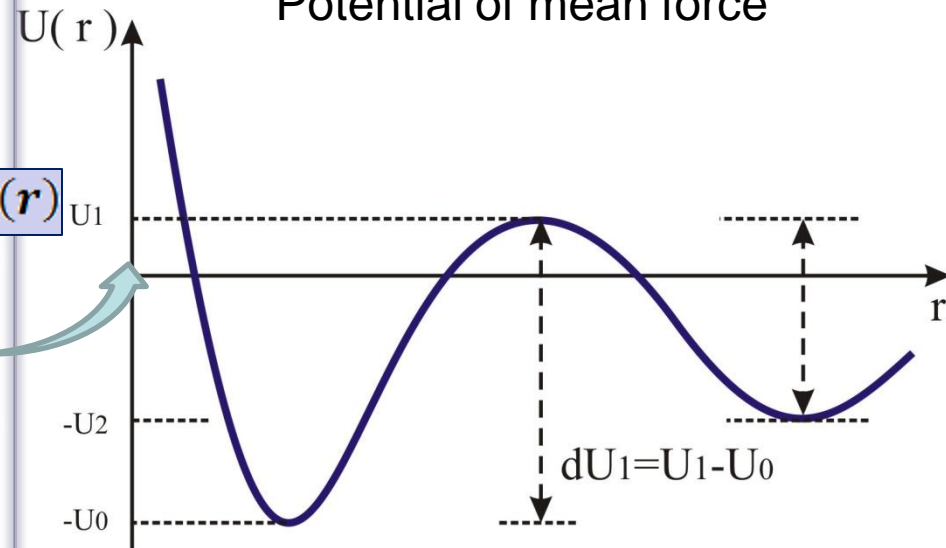
Lavery R., Moakher M., Maddocks J.H.,  
 Petkeviciute D., Zakrzewska K., *Nucleic Acids Res.*,  
**37**, 5917-5929 (2009).

# Estimation of the potential of mean force

Radial distribution function



Potential of mean force



Fitting of the RDF shape:

$$g(r) = g_0 + \frac{A_1}{w_1 \sqrt{\frac{\pi}{2}}} e^{-\frac{2(r-r_{01})^2}{w_1^2}} + \dots$$

$$U(r)|_{r_{01}} \approx U_1 + \frac{\gamma_1}{2} (r - r_{01})^2 =$$

$$-g_0 - \frac{A_1}{w_1 \sqrt{\frac{\pi}{2}}} + \frac{2}{w_1^2} (r - r_{01})^2$$

$\frac{w}{2}$  is the standard deviation.

Estimation of average frequency of vibrations in the potential well:

$$\nu_1 = \frac{1}{2\pi} \sqrt{\frac{\gamma_1}{m}} = \frac{1}{\pi w_1}$$

Estimation of average residence time of the ion in the potential well:

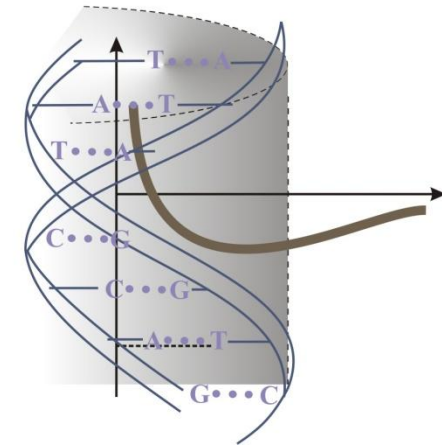
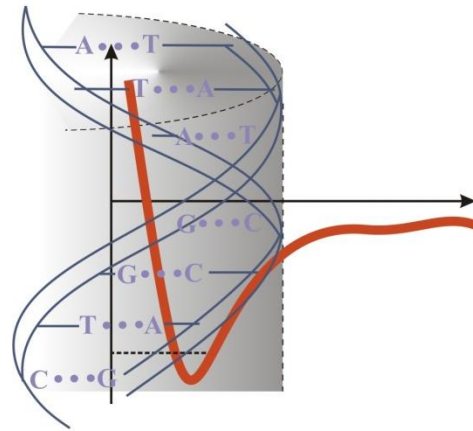
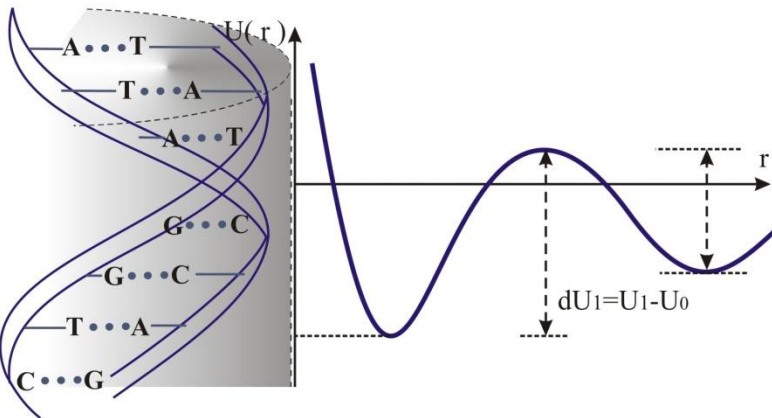
$$\tau = \tau_0 e^{\frac{\Delta U}{k_B T}}$$

# Residence time of counterions and frequencies of vibrations

Phosphate groups

Minor groove

MAJOR groove



		Na-DNA	K-DNA	Cs-DNA	Mg-DNA
Phosphate group	$\omega$ , THz	5.5	3.7	1.7	1.4
	$\tau$ , ps	122	15	32	400
Minor groove	$\omega$ , THz	0.94	2.8	1.5	0.8
	$\tau$ , ps	429	142	2052	800
Well 1	$R_1$ , Å	2.35	2.65	3.10	--
Well 2	$R_2$ , Å	4.40	4.70	4.29	4.20

# Conclusions

1. The counterions form a dynamical ordered structure around the DNA double helix that has a properties of the lattice of ionic crystal.
2. The manifestations of the counterion ordering are found in the DNA low-frequency spectra ( $<200 \text{ cm}^{-1}$ ) as the modes of ion-phosphate vibrations.
3. MD simulation data show:
  - sodium counterions are localized from the outside of the double helix and at the top edge of the minor groove; the counterions interact with the atoms of the phosphate groups directly or via water molecules;
  - potassium counterions may be localized at the both external or internal compartments of the double helix; the counterions interact with the atoms of the double helix directly or via water molecules;
  - cesium counterions penetrate deeply inside the minor groove of the double helix and interact directly with the atoms of basses; the ions spend rather long time inside the minor groove ( $> 1 \text{ ns}$ ) and may form a regular structure at this time scale;
  - the ions in the major groove of the double helix are free to move;
  - the frequencies of counterion vibrations corresponding to the potential wells characterizes the dynamics of the ions in bonded states; the estimated frequencies of vibrations of the counterions qualitatively agree with the previous calculations and experimental data.

# Acknowledgments

## Laboratory of Biophysics of Macromolecules



Prof. S.N.Volkov  
(Head of the Lab.)

Dr. D.V.Piatnytskyi

Ms. P.P.Kanevska

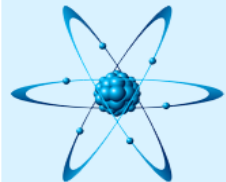
Prof. E.S.Kryachko

O.O.Zdorevskyi  
(Ph.D. stud.)

UKRAINIAN  
NATIONAL  
GRID

## Computational Facilities

Memorandum of  
Understanding  
between UW-CHTC  
and BCC



Grid computing cluster of the  
Bogolyubov Institute for Theoretical Physics



*Thank you for  
your attention!*

Bogolyubov Institute for  
Theoretical Physics