

Conference on Atomistic Simulations of Biomolecules

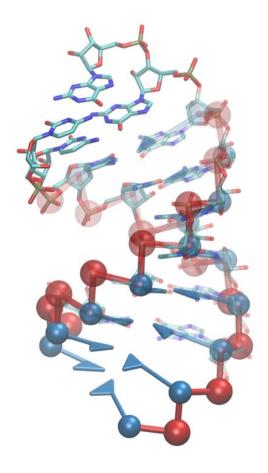
ICTP, Trieste, March 2017



Introducing atomistic details into a base-centered representation of RNA

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Molecular and Statistical Biophysics - SISSA, Trieste, Italy

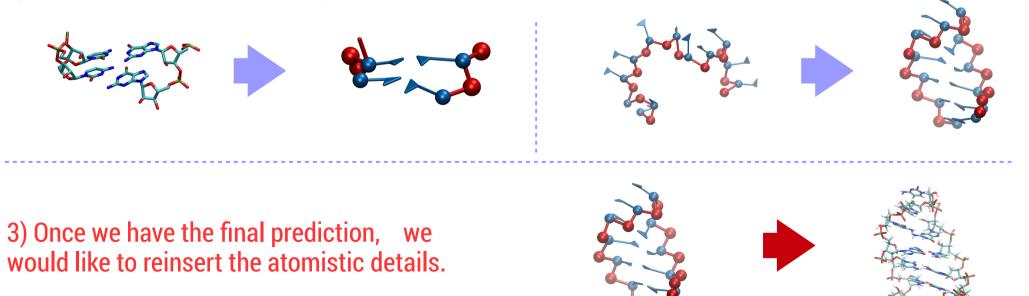


My talk in a nutshell

1) We want to predict 3D structure of RNA fragments from their sequence.



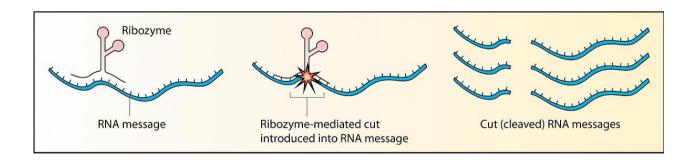
2)We use a simplified, coarse-grained model for folding the structure.

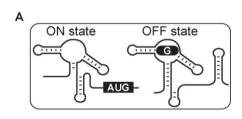


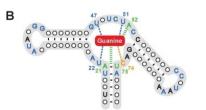
Introduction

RNA has a considerable biological importance :

- Transcription
- Translation
- Catalytic role (ribozymes)
- Regulatory function (miRNAs, riboswitches, ...)



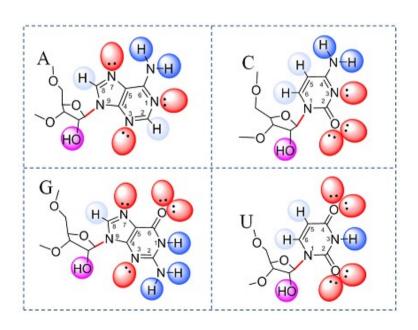


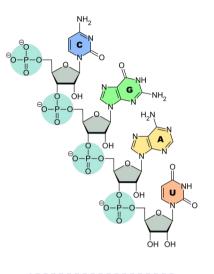


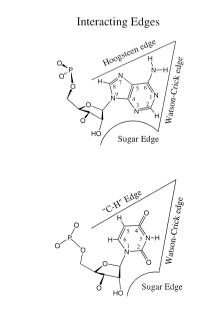
Structure determines function!

Something about RNA...

- Structure: Sugar, Base and Phosphate group.
- Differences with DNA:
 - Ribose sugar instead of deoxyribose : more reactive.
 - Uracil instead of Thymine.
- Rich set of base-base interactions





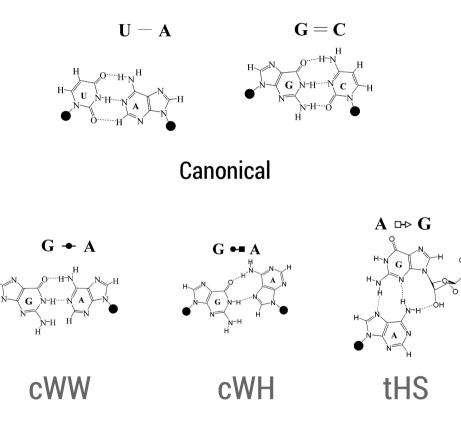


Leontis, Stombaugh and Westhof, Nucleic Acids Res. (2002)

Something about RNA...

Rich set of base-base interactions

No.	Glycosidic bond orientation	Interacting edges		Abbreviation	Symbol	Triangle
		NT1	NT2	Abbreviation	Symbol	abstraction
1	Cis	Watson-crick	Watson-crick	cWW	-	
2	Trans	Watson-crick	Watson-crick	tVVVV	-0-	
3	Cis	Watson-crick	Hoogsteen	cWH	$\bullet\blacksquare$	
		Hoogsteen	Watson-crick	cHW	H	
4	Trans	Watson-crick	Hoogsteen	tWH	Ю	
		Hoogsteen	Watson-crick	tHW	Б	
5	Cis	Watson-crick	Sugar edge	cWS		
		Sugar edge	Watson-crick	cSW	-	
6	Trans	Watson-crick	Sugar edge	tWS	O-⊅	
		Sugar edge	Watson-crick	tSW	ф	
7	Cis	Hoogsteen	Hoogsteen	сНН	-#-	
8	Trans	Hoogsteen	Hoogsteen	tHH		
9	Cis	Hoogsteen	Sugar edge	cHS		
		Sugar edge	Hoogsteen	cSH	╇	
10	Trans	Hoogsteen	Sugar edge	tHS		
		Sugar edge	Hoogsteen	tSH	\triangleleft	
11	Cis	Sugar edge (priority)	Sugar edge	cSs	+	
		Sugar edge	Sugar edge (priority)	csS	-	
12	Trans	Sugar edge (priority)	Sugar edge	tSs		
		Sugar edge	Sugar edge (priority)	tsS	$ \rightarrow$	



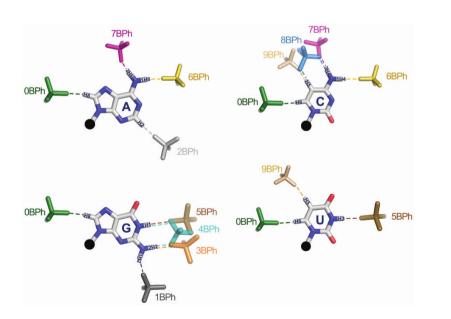
Non-canonical

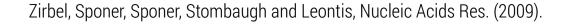
Leontis, Stombaugh and Westhof, Nucleic Acids Res. (2002)

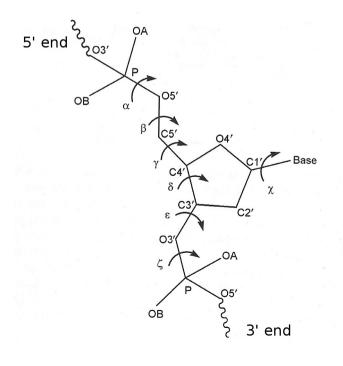
Something about RNA...

Also, interactions between bases and phosphate groups

Backbone conformation defined by 6 (7) dihedral angles

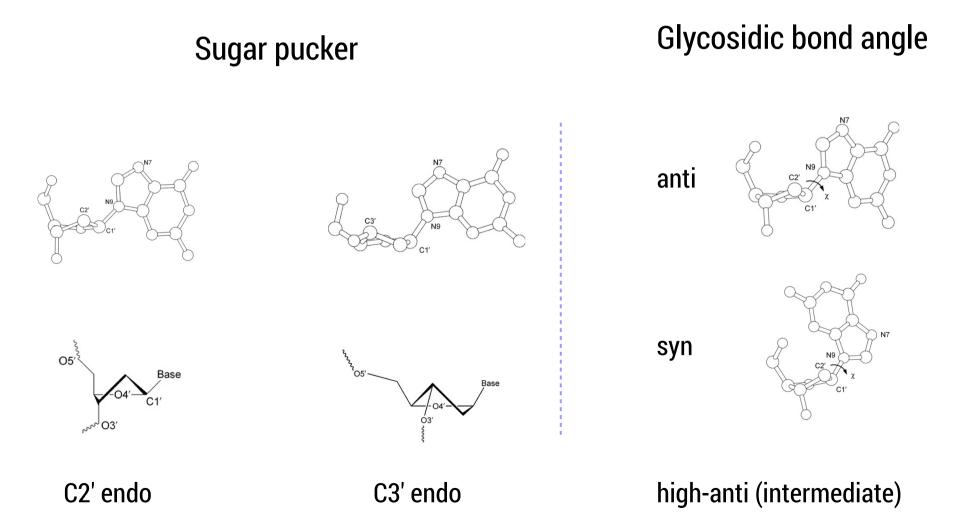






Richardson et al., RNA (2008).

Something about RNA



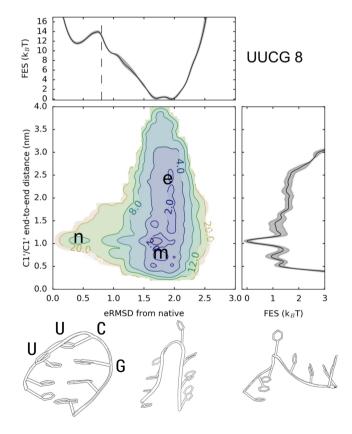
Motivation

- Structure prediction is an important, but tough task. Brute-force Atomistic MD might be extremely inneficient.
- RNA atomistic force fields still require refinement.

UUCG tetraloop :

Misfolded structures are more stable than native

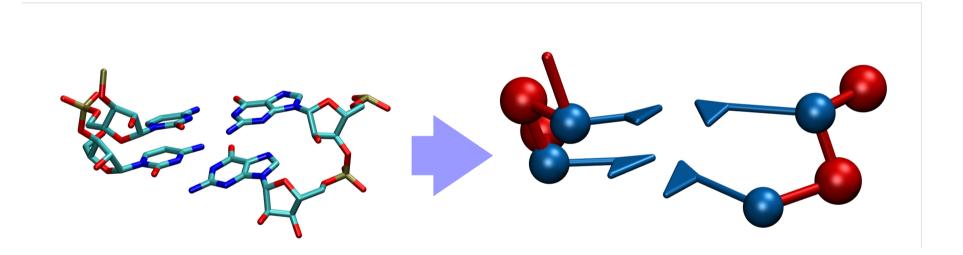
Bottaro, Banas, Sponer and Bussi, J. Phys. Chem. Lett. (2016)



Coarse-grained model

A simpler, faster representation, with effective interactions.

SPQR model : a nucleoside is an anisotropic particle; phosphate group is a point particle.



Thoroughly description of canonical and non-canonical base pairs.

Base-phosphate interactions

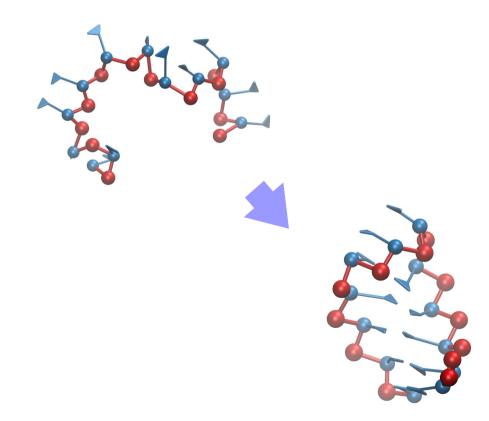
Puckers and glycosidic bond angles specified

Poblete, Bottaro and Bussi, submitted.

Coarse-grained model

A simpler, faster representation, with effective interactions.

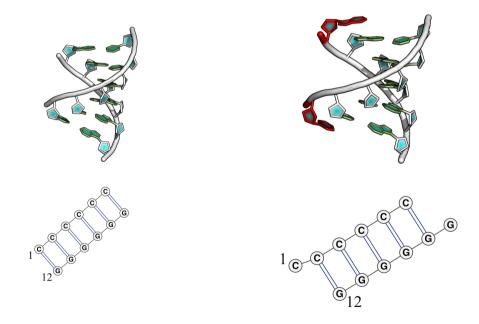
SPQR model : a nucleoside is an anisotropic particle; phosphate group is a point particle.



Simulated annealing to predict structures of minimum energy

GCAA tetraloop (pdb 1ZIH)

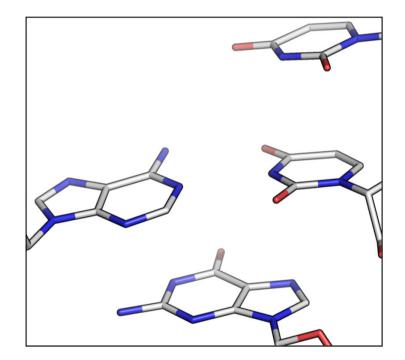
- Once the structure is predicted, we wish to reintroduce atomistic detail.
- Minimize "distance" between atomistic and predicted structure.
- RMSD : Misleading, not suitable to compare nucleic acids structure



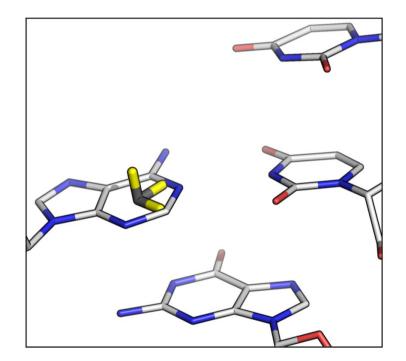
1.9 A (backbone-only 1.4 A, bases-only 1.9)

Bottaro, Di Palma and Bussi, Nucleic Acids Res. (2014).

A simplified geometrical description of base arrangement

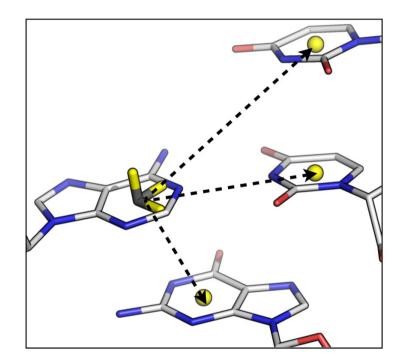


A simplified geometrical description of base arrangement



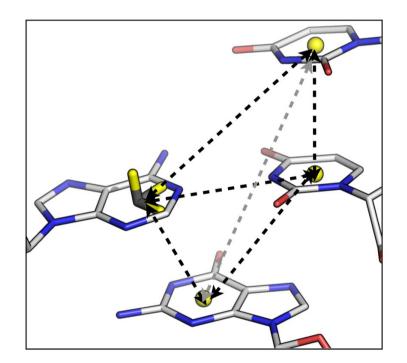
Sit on a base

A simplified geometrical description of base arrangement



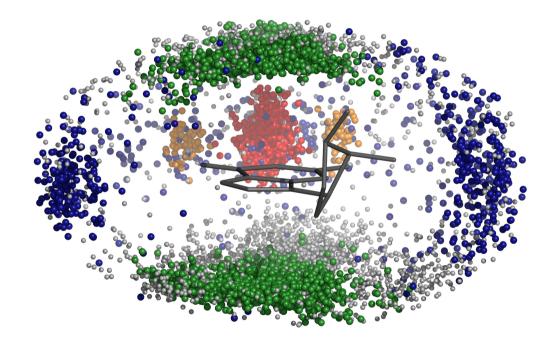
Calculate the distance vectors from here

A simplified geometrical description of base arrangement



Repeat the procedure inside a cutoff distance

A simplified geometrical description of base arrangement



Ellipsoidal distance within a cutoff,

Captures better base pairing and stacking,

And describe better the nucleobase volume

Data from large ribosomal subunit PDB:1S72, Klein, Moore, and Steitz, J. Mol. Biol. (2004).

Bottaro, Di Palma and Bussi, Nucleic Acids Res. (2014). Bottaro, Di Palma and Bussi, RNA and Disease (2015).

With this collection of vectors, we introduce the $\mathcal{E}\text{RMSD}$, a more specific distance between RNA structures.

$$\mathbf{G}(\tilde{\mathbf{r}}) = \begin{pmatrix} \sin(\gamma \tilde{r}) \frac{\tilde{r}_x}{\tilde{r}} \\ \sin(\gamma \tilde{r}) \frac{\tilde{r}_y}{\tilde{r}} \\ \sin(\gamma \tilde{r}) \frac{\tilde{r}_z}{\tilde{r}} \\ 1 + \cos(\gamma \tilde{r}) \end{pmatrix} \times \frac{\Theta(\tilde{r}_{\text{cutoff}} - \tilde{r})}{\gamma}$$

$$\gamma = \pi / \tilde{r}_{\rm cutoff}$$

$$\mathcal{E}\text{RMSD} = \sqrt{\frac{1}{N} \sum_{j,k} |\mathbf{G}(\tilde{\mathbf{r}}_{jk}^{\alpha}) - \mathbf{G}(\tilde{\mathbf{r}}_{jk}^{\beta})|^2}$$

Backmapping procedure:

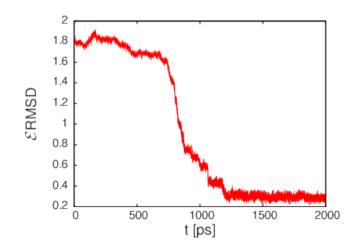
Steered MD, minimizing the ERMSD with respect to a reference structure (typically, predicted with our coarse-grained method)

Short simulation of 3 ns

Explicit solvent, GROMACS 4.6.7 (*), harmonic constraint implemented in PLUMED(**)

$$V(ec{s},t)=rac{1}{2}\kappa(t)(ec{s}-ec{s}_0(t))^2$$
 .



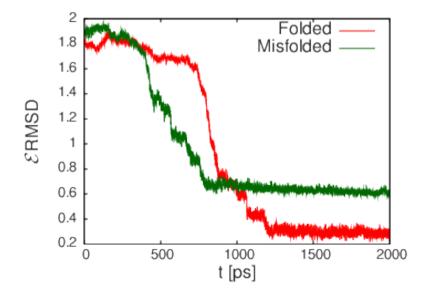


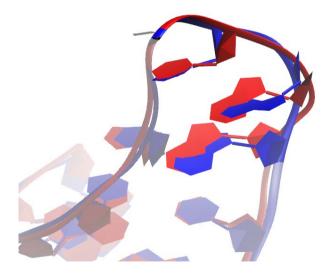
* www.gromacs.org, Amber99 (parmbsc0+OL3), TIP3P water, Na+ ions (Joung and Cheatham, J. Phys. Chem. B (2008)).
** www.plumed.org

Backmapping procedure:

Steered MD, minimizing the \mathcal{E} RMSD with respect to a reference structure (typically, predicted with our coarse-grained method).

Misfolded cases of bases not flipped are detected





What happens with the backbone conformations?

We performed a test on :

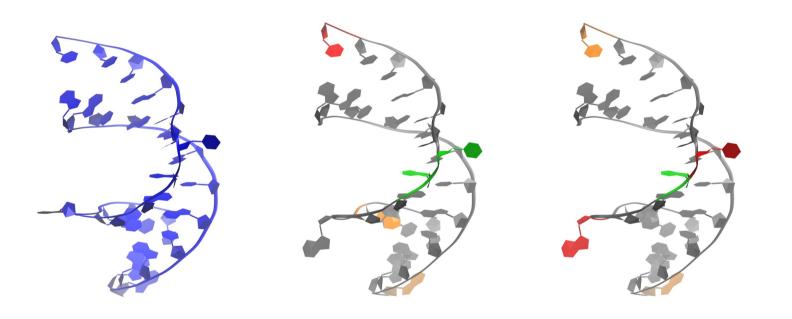
Five crystal structures to their native state.

Three predicted structures from **SPQR** CG annealing.

Ran 6 independent simulations. We chose the 50 lowest \mathcal{E} RMSD configurations, and analyze their pucker and glycosidic bond angle using **dangle**.

http://kinemage.biochem.duke.edu/software/dangle.help.html

Backmapping on the crystal structures



In the lowest 50 & RMSD configurations, right conformation found in:

More than 80%

Between 50% and 80%

Less than 50%

Well predicted, uncommon cases

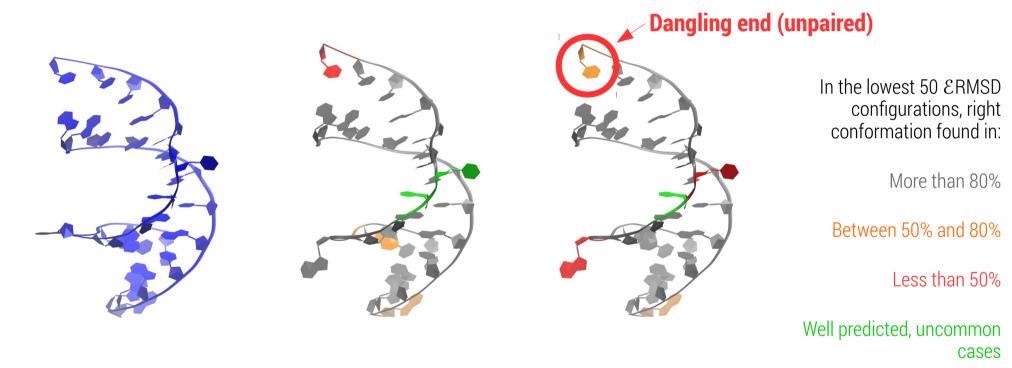
Native

Sugar puckers

Glycosidic bond angle conformation

PDB:1csl

Backmapping on the crystal structures



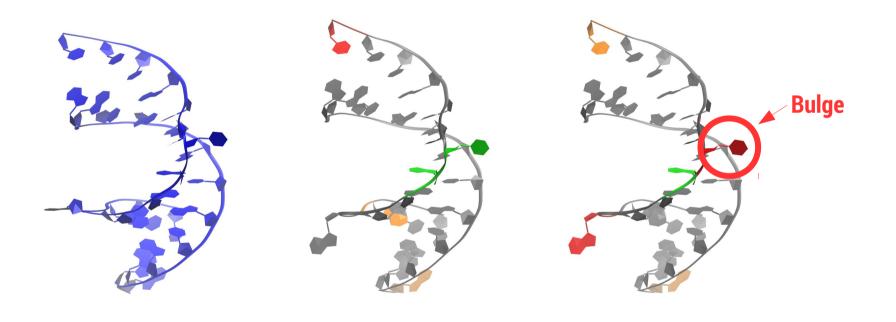
Native

Sugar puckers

Glycosidic bond angle conformation

PDB:1csl

Backmapping on the crystal structures



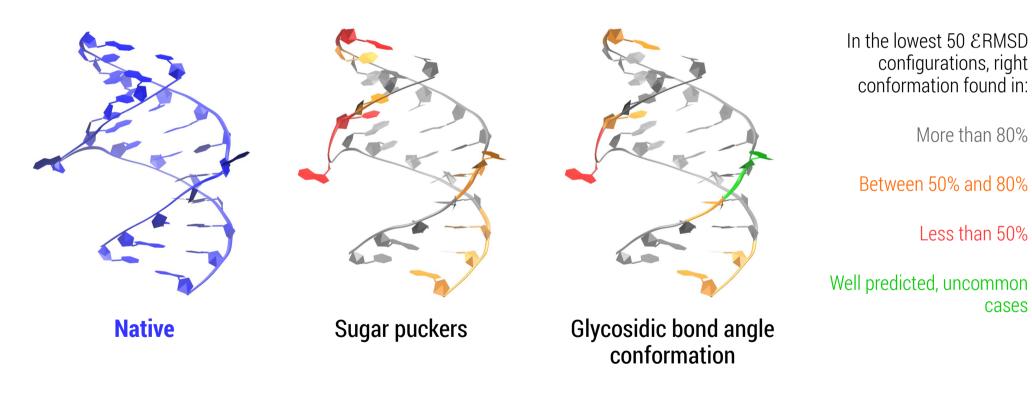
Native

Sugar puckers

Glycosidic bond angle conformation

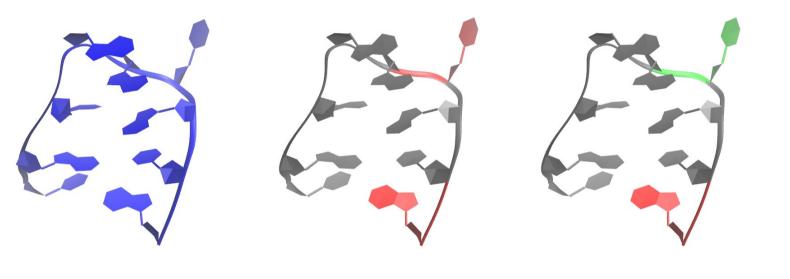
PDB:1csl

Backmapping on the crystal structures



PDB:1i9x

Backmapping on the crystal structures



In the lowest 50 & RMSD configurations, right conformation found in:

More than 80%

Between 50% and 80%

Less than 50%

Well predicted, uncommon cases

Native

Sugar puckers

Glycosidic bond angle conformation

PDB:1xjr

Backmapping on the crystal structures



In the lowest 50 & RMSD configurations, right conformation found in:

More than 80%

Between 50% and 80%

Less than 50%

Well predicted, uncommon cases

Native

Sugar puckers

Glycosidic bond angle conformation

PDB:2gdi

Backmapping on the crystal structures



In the lowest 50 & RMSD configurations, right conformation found in:

More than 80%

Between 50% and 80%

Less than 50%

Well predicted, uncommon cases

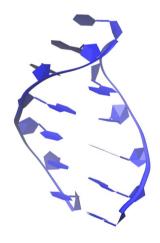
Native

Sugar puckers

Glycosidic bond angle conformation

PDB:354d

Backmapping on the predicted structures from coarse-grained model.







In the lowest 50 & RMSD configurations, right conformation found in:

More than 80%

Between 50% and 80%

Less than 50%

Well predicted, uncommon cases

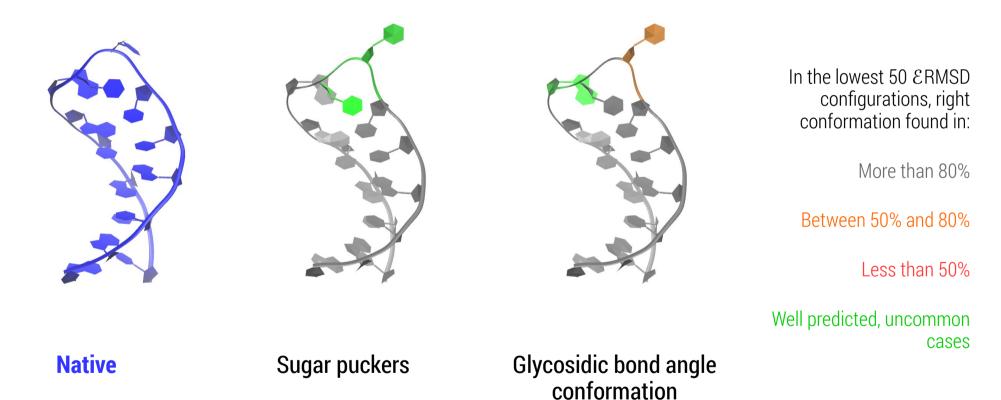
Native

Sugar puckers

Glycosidic bond angle conformation

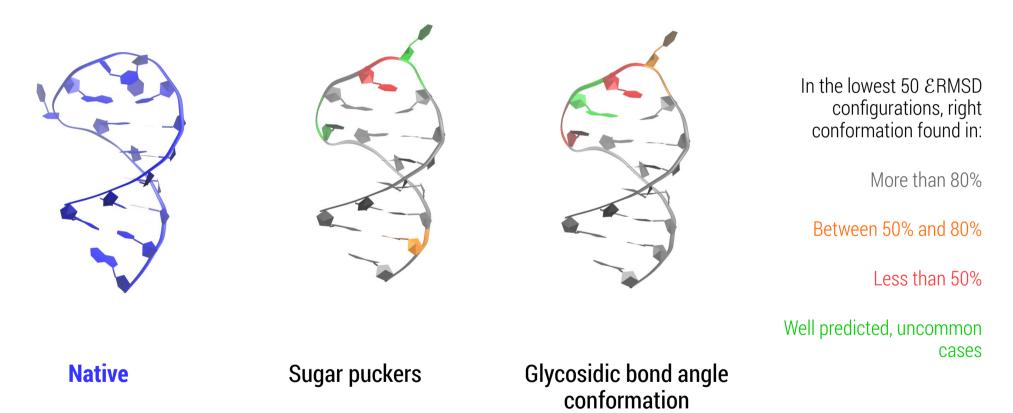
PDB:1zih

Backmapping on the predicted structures from coarse-grained model.



PDB:2koc (UUCG tetraloop) "Flipped" G and C, with not typical (A-form) glycosidic and pucker conformations, respectively.

Backmapping on the predicted structures from coarse-grained model.



Conclusions

- Introduced a backmapping procedure for nucleic acids, based on \mathcal{E} RMSD, more suitable metric for RNA structures.
- Good results, but particularly difficult for bulges, dangling ends and end nucleotides in general.
- Effective in cases where bases are "flipped".

Acknowledgements

European Research Council.