### Structure, mechanism, and inhibition of the membrane motor of the ATP synthase inferred from quantitative integrative modeling

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# ATP synthases uses the electrochemical gradient to catalyze the production of ATP

#### During cell metabolism proton or sodium ions are exported across membranes, establishing an electrochemical potential gradient



From von Ballmoos *et al.,* Annu Rev Biochem. '09



How the ions translocation power the ATP production?

#### Architecture of the ATP synthase



No atomistic detail of complex or isolated subunit-a CryoEM maps and low resolution X-ray structure show a horizontal arrangement of subunit-a helices, but different topologies are proposed





Zhou et al. Elife 2015



Goal: provide a quantitate interpretation of the low resolution (~7 Å) cryoEM structural data

Morales-Rios et al. PNAS 2015





and Cys Xlinks

variant residue pairs



### Homology model c-ring of Polytomella based on yeast ring



		Mitochondrial largeting sequence
Polytomella sp. Pr.	-53	
C. reinhardtii	-85	MASSQKAVQMSLGAVRSLSTGMTRLQAASAQGMLASSQGAAGQM <mark>E</mark> KPVMVATQVPILGASPSAIASGIRASAKASPMSLAPQRSM-1
Bos taurus	-68	
S. cerevisiae		M 1
l. tartaricus	1	
S. oleracea	1	
S. platensis	1	
B. pseudofirmus		М 1
		Outer Helix
• • •		
Polytomella p. Pr.	1	<mark>S V L A A S K M V G A G C A T</mark> I A L A G V G A G L G V M F G S L I N G A A R N P N I A K Q L V G Y A L L G F A L T E S I A L F S L L V V F L I L F A 74
C. reinhardtii	1	S V L A A S KM V G A <mark>G C A T</mark> I A L A <mark>G V G</mark> A <mark>G L G V M F G S</mark> L I <mark>N G</mark> A A <b>R N P N</b> I A <b>K Q</b> L V G Y A L L G F A L T <mark>E S</mark> I A L F <mark>S</mark> L L V V F L I L F A 74
Bos taurus	1	DIDTAAKFIGAGAATVGVAGSGAGIGTVFGSLIIGYARNPSLKQQLFSYAILGFALSEAMGLFCLMVAFLILFAM 75
S. cerevisiae	2	QLVLAAK <mark>Y</mark> IGAGI <mark>ST</mark> IGLLGAGIGIAIVFAALI <mark>NGVSRNPS</mark> IK <mark>DT</mark> VFPMAILGFAL <mark>SE</mark> ATGLFCLMV <mark>S</mark> FLLLFGV76
I. tartaricus	8	TVVLAASAVGAGTAMIAGIGPGVGQGYAAGKAVESVARQPEAKGDIISTMVLGQAVAESTGIYSLVIALILLYANPFVGLLG. 89
S. oleracea	8	S V I A A G L A V G L A <mark>S I G P G V G Q G T</mark> A A G Q A V <mark>E</mark> G I A R Q P E A <mark>E</mark> G K I R G T L L L S L A F M E A L <b>T</b> I Y G L V V A L A L L F A N P 78
S. platensis	10	SVIAAALAVG IG <mark>S</mark> IGPGLGQGQAAGQAV <mark>E</mark> GIARQPEA <mark>E</mark> GKIRGTLLLSLAFMEAL <mark>T</mark> IYGLVVALVLLFA <mark>NPF</mark> V 82
B pseudofirmus	2	AFLGAAIAAGLAAVAGAIAVAIIVKATIEGTTROPELRGTLOTLMFIGVALAEAVPIIAIVISLLILF69

#### Extract spatial restrains and transfer them to Polytomella c-ring



**2.0Å resolution** Xray of **yeast** c-ring; 47% % % Widentity with *Polytomella* c-subunit









Ca-Ca distances;

hydrogen bonds;

main chain & side

chain dihedrals





2000 models

### Homology model c-ring of Polytomella based on yeast ring



		Mitochondrial largeting sequence
Polytomella sp. Pr.	-53	м <mark>5</mark> V Q R L <mark>S L G</mark> A A R C L S A G V A R V Q A S Q A L V A Q K A V A V A P T R A Q A A P A <b>Р</b> V A Q V R S M -1
C. reinhardtii	-85	MASSQKAVQMSLGAVRSLSTGMTRLQAASAQGMLASSQGAAGQM <mark>E</mark> KP VMVATQVPILGASPSAIASGIRASAKASPMSLAPQRSM-1
Bos taurus	-68	
S. cerevisiae		M 1
l. tartaricus	1	
S. oleracea	1	NPLIAAA 7
S. platensis	1	M <mark>e</mark> snl <mark>tt</mark> aa 9
B. pseudofirmus		M 1
		Outer Helix
	1	
Polytomena p. Pr.	1	
C. reinhardtii	1	SVLAAS KMVGAGCATTALAGVGAGLGVMFGSLTNGAAKNPNTAKQLVGYALLGFALTESTALFSLLVVFLTLFA /4
Bos taurus	1	DIDITAAKFIGAGAATVGVAGSGAGIGTVFGSLIIGYARNPSLKQQLFSYAILGFALSEAMGLFCLMVAFLILFAM 75
S. cerevisiae	2	QLVLAAKYIGAGI <mark>ST</mark> IGLLGAGIGIAIVFAALI <mark>NGVSRNPS</mark> IK <mark>DT</mark> VFPMAILGFAL <mark>SE</mark> ATGLFCLMV <mark>S</mark> FLLLFGV 76
I. tartaricus	8	TVVLAASAVGAGTAMIAGIGPGVGQGYAAGKAVESVARQPEAKGDIISTMVLGQAVAESTGIYSLVIALILLYANPFVGLLG 89
S. oleracea	8	<mark>S</mark> VIAAGLAVG LA <mark>S</mark> IGPGVGQG TAAGQAVEGIARQPEAEGKIRG TLLLS LAFMEAL TIYGLVVALALLFANP 78
S. platensis	10	SVIAAALAVG IGSIG PG LG OG O AAG O AVEG IA ROPE AEGKIRG TLLLS LAFME AL TIYG LVVALVLLFAN PF V 82

#### Extract spatial restrains and transfer them to Polytomella c-ring



Ca-Ca distances; hydrogen bonds; main chain & side chain dihedrals

**2.0Å resolution** Xray of **yeast** c-ring; **47% %identity** with *Polytomella* c-subunit



Models of Polytomella c-ring that satisfy all the restraints as well as possible

Select one model based on DOPE and GA341 score

BUT clashes and some wrong angle lengths



TM regions < 1 Å

Cα-RMSD from native structure



Forrest et al. BJ 2006



#### Refinement of the c-ring homology model into the cryoEM map





and Cys Xlinks

variant residue pairs



#### Model of TM4 in/TM5 out and TM4 out/TM5 in positions



#### Model of TM4 in/TM5 out and TM4 out/TM5 in positions



TM4-TM5 covariant residues cannot distinguish the two C-ter positions

Data between subunit-a/c-ring is needed

# Covariant residues between subunit-a and c-ring select TM4 out/TM5 in assignment



#### Cys crosslinked residues on *E.coli* Fo select TM4 out/TM5 in assignment







How to distinguish between TM4 out/TM5 in best traces?

Conserved Arg on TM4 can be translated to a conserved Gln on TM5 (252 in *E.coli*) retaining the enzymatic function

Ishmukhametov et al. BBA Bioener 2008 Bae & Vik et al. BBA Bioener 2009

**Conserved Arg and GIn** of subunit-a must be **proximal to conserved Glu** of c-ring

#### Threading-1 of is selected based on functional data



Maximal distance (Å)



![](_page_19_Figure_1.jpeg)

Model of TM2-TM3 hairpin in a clockwise topology respect to TM4-TM5 as indicated by subunit-a residue covariance

![](_page_20_Figure_1.jpeg)

![](_page_21_Figure_1.jpeg)

![](_page_22_Figure_1.jpeg)

#### Mapping Cd<sup>2+</sup> accessible residues on c-ring/subunit-a structure

![](_page_23_Figure_1.jpeg)

# Structure of the c-ring/subunit-a complex supports the two-half-channel hypothesis

![](_page_24_Figure_1.jpeg)

Ionizable residues lines on the proposed P and N-channel while a/c-interface on P-side is sealed

Conserved **Arg** between the two channels **shortcuts the proton leakage** across them

H154 and E194 are highly covariant; proton buffer

Leone and Faraldo J. Gen Phys. 2016

![](_page_24_Picture_6.jpeg)

Solvent may be stabilized by interaction with polar residues at the N-channel

N-channel

E215 D219

E131 affects kinetics but not H<sup>+</sup>-binding

#### ATP synthase H<sup>+</sup> transport mechanism derived from our model

![](_page_25_Figure_1.jpeg)

Leone and Faraldo J. Gen Phys. 2016

# Model of c-ring/subunit-a complex is in agreement with previously published biochemical data

![](_page_26_Figure_1.jpeg)

Leone and Faraldo J. Gen Phys. 2016

ATP synthase as potential pharmacological target against dormant or resistant bacterial strains

Recent FDA-approved antituberculosis **drug targets** ATP synthase **membrane domain** 

ATP synthase as potential pharmacological target against dormant or resistant bacterial strains

Recent FDA-approved antituberculosis **drug targets** ATP synthase **membrane domain** 

Oligomycin **antibiotic binds** to the **cring ion binding site** (Xray structure) and **resistant mutations** are located in both **subunit-c and –a** 

Probably antibiotics **binds to the a/cinterface** blocking the rotation

ATP synthase as potential pharmacological target against dormant or resistant bacterial strains

Recent FDA-approved antituberculosis **drug targets** ATP synthase **membrane domain** 

Oligomycin **antibiotic binds** to the **cring ion binding site** (Xray structure) and **resistant mutations** are located in both **subunit-c and –a** 

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![](_page_29_Figure_6.jpeg)

ATP synthase as potential pharmacological target against dormant or resistant bacterial strains

Recent FDA-approved antituberculosis **drug targets** ATP synthase **membrane domain** 

Oligomycin **antibiotic binds** to the **cring ion binding site** (Xray structure) and **resistant mutations** are located in both **subunit-c and –a** 

Probably antibiotics **binds to the a/cinterface** blocking the rotation

![](_page_30_Figure_6.jpeg)

![](_page_31_Figure_1.jpeg)

Recent FDA-approved antituberculosis **drug targets** ATP synthase **membrane domain** 

Oligomycin **antibiotic binds** to the **cring ion binding site** (Xray structure) and **resistant mutations** are located in both **subunit-c and –a** 

Probably antibiotics **binds to the a/cinterface** blocking the rotation

![](_page_31_Figure_6.jpeg)

![](_page_32_Figure_1.jpeg)

Recent FDA-approved antituberculosis **drug targets** ATP synthase **membrane domain** 

Oligomycin **antibiotic binds** to the **cring ion binding site** (Xray structure) and **resistant mutations** are located in both **subunit-c and –a** 

Probably antibiotics **binds to the a/cinterface** blocking the rotation

![](_page_32_Figure_6.jpeg)

ATP synthase as potential pharmacological target against dormant or resistant bacterial strains **Recent FDA-approved anti-**ATP synthesis tuberculosis **drug targets** ATP synthase membrane domain Oligomycin **antibiotic binds** to the **c**ring ion binding site (Xray structure) and resistant mutations are located in both subunit-c and -a oligomycin Probably antibiotics binds to the a/cinterface blocking the rotation R145 Q20 resistant

a-subunit

mutants

Oligomycin **inhibit** both **ATP synthesis and hydrolisis** 

![](_page_33_Figure_3.jpeg)

c-ring

**ATP hydrolysis** 

Leone and Faraldo J. Gen Phys. 2016

#### Summary

![](_page_34_Figure_1.jpeg)

We have integrated different types of information to interpret the cryoEM structure

of an ATP synthase membrane rotor

Our c-ring/subunit-a complex model supports a **two-half-channel model** 

Previous **biochemical data is in agreement** with the cryoEM structure

Our model provides insights on how some antibiotics can inhibit both ATP synthesis and hydrolysis

The strategy devised in this work can be applied to interpret low resolution cryoEM structures in other systems

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