

# Principles of Microscopy II: Super-resolution

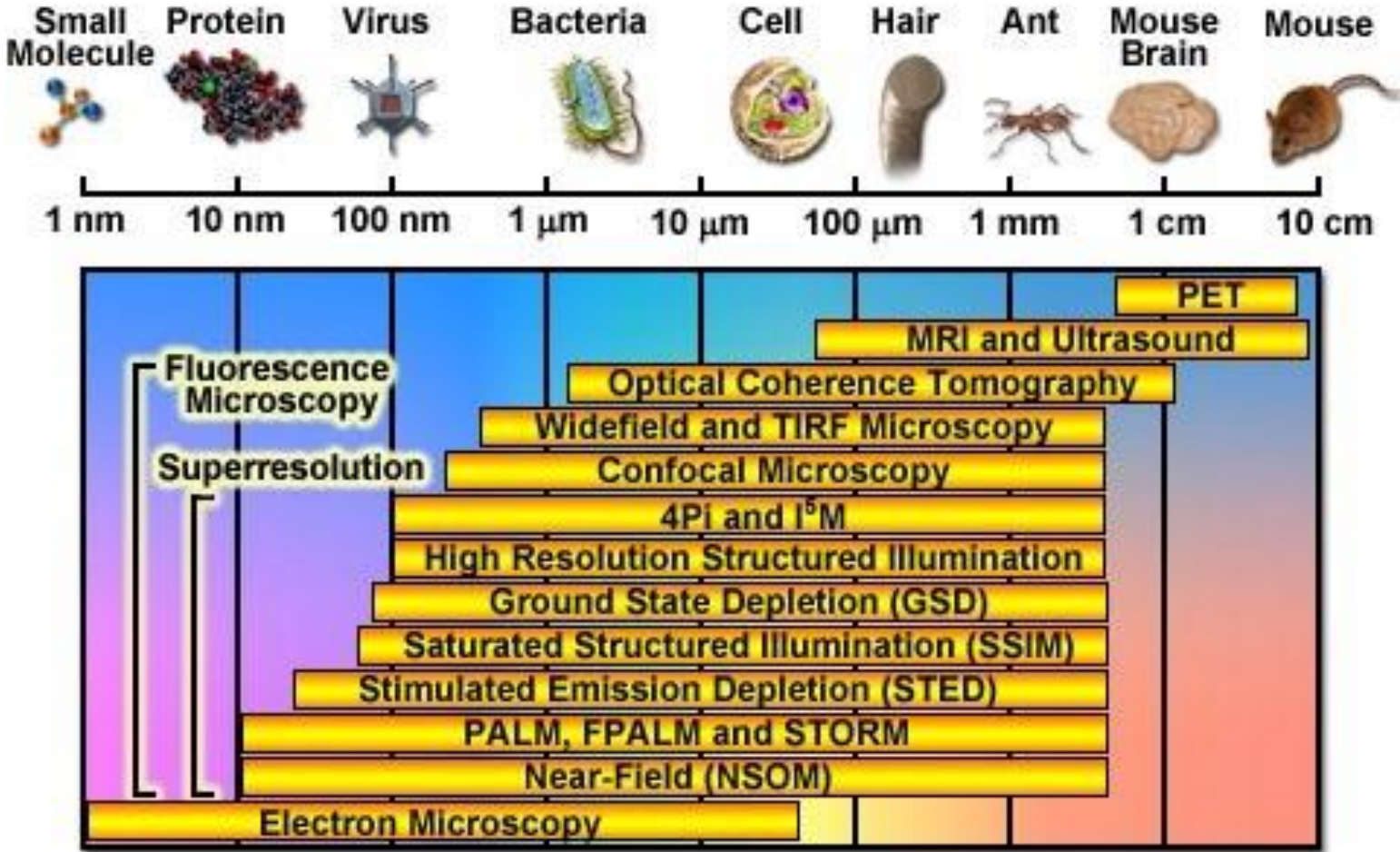
**Humberto Cabrera**

Venezuelan Institute for Scientific Research  
International Centre for Theoretical Physics

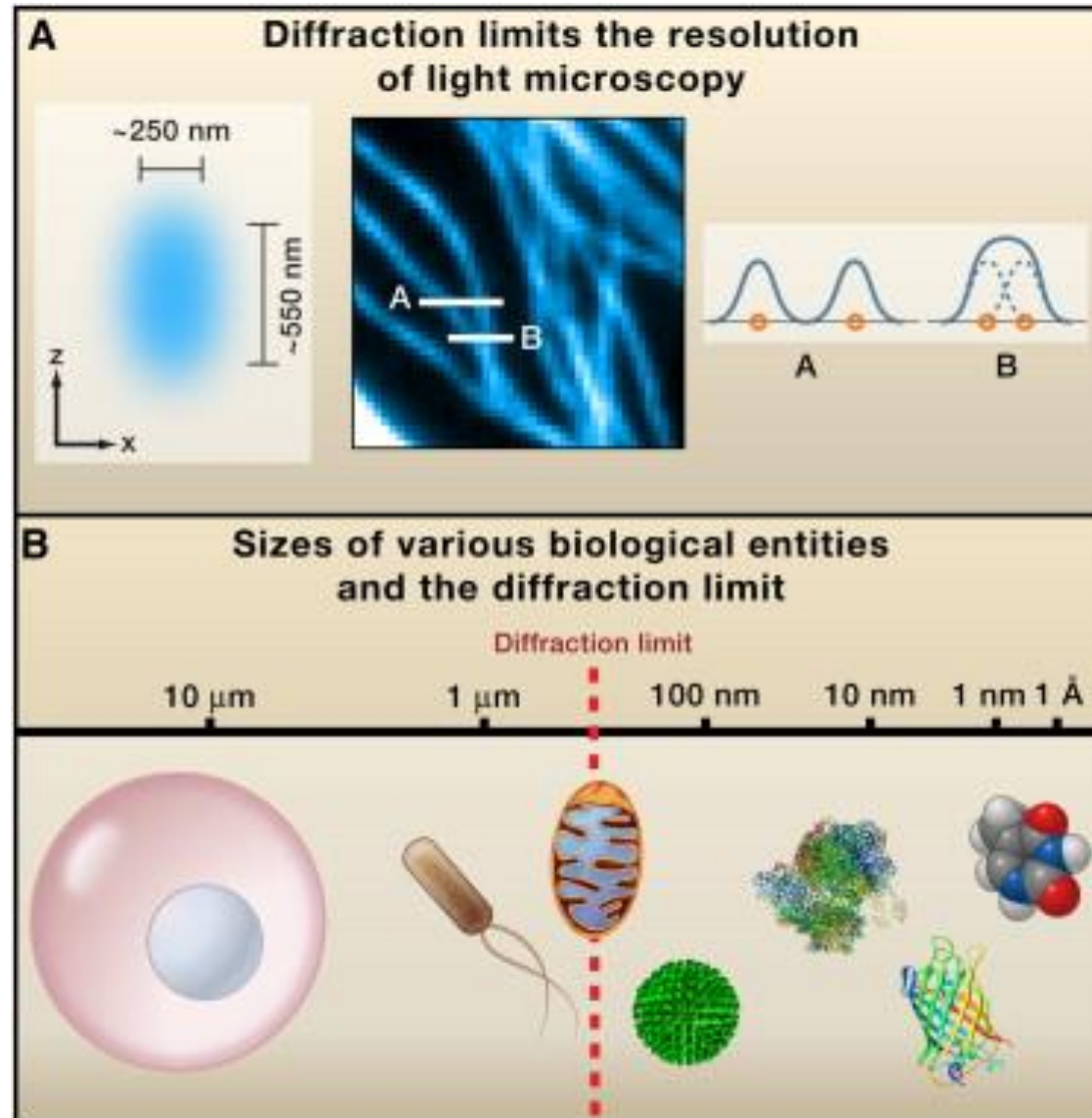
Preparatory School to the Winter College on Optics: Advanced Optical Techniques for Bio-  
imaging

# Diffraction limit of imaging techniques

Spatial Resolution of Biological Imaging Techniques



# The diffraction limit



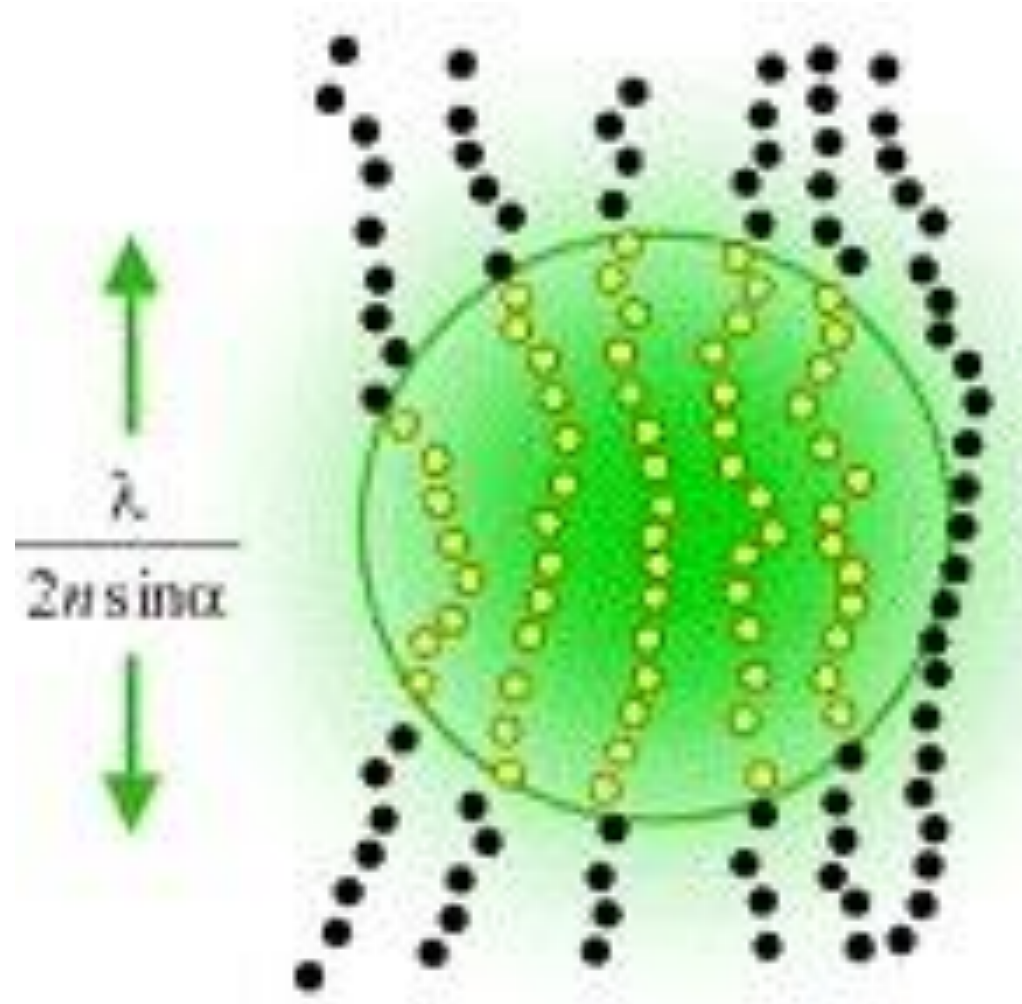
# The diffraction limit of light



Ernst Abbe  
(1840-1905)

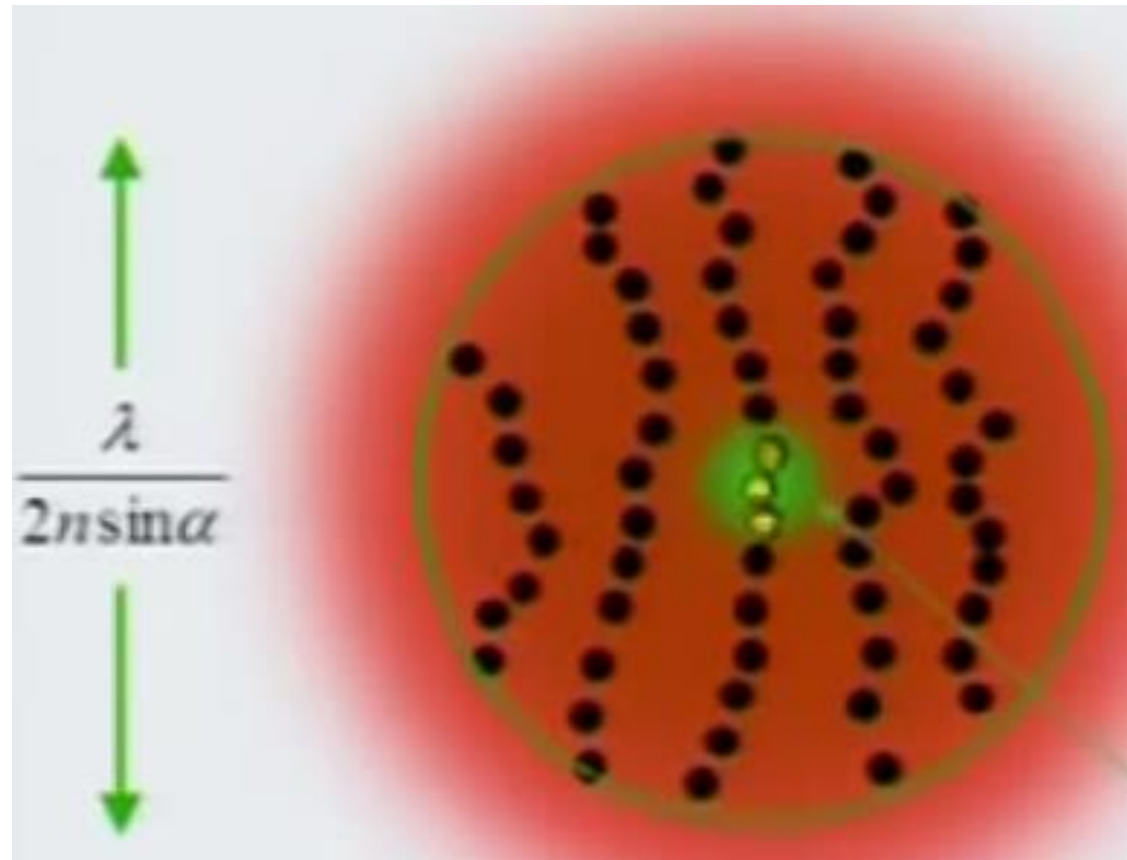


## The diffraction limit

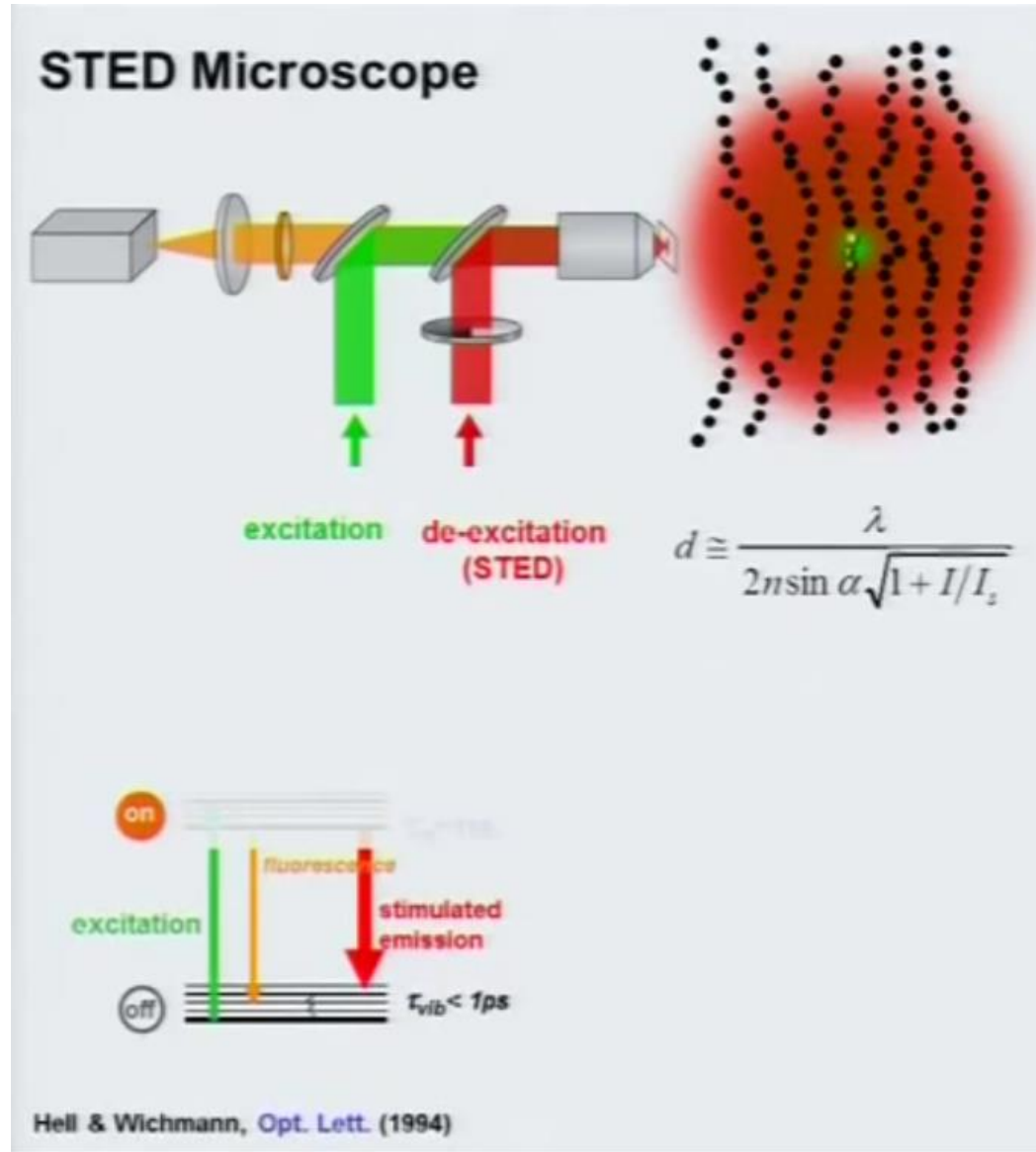


Molecules within the región  $< 200$  nm are not discernible

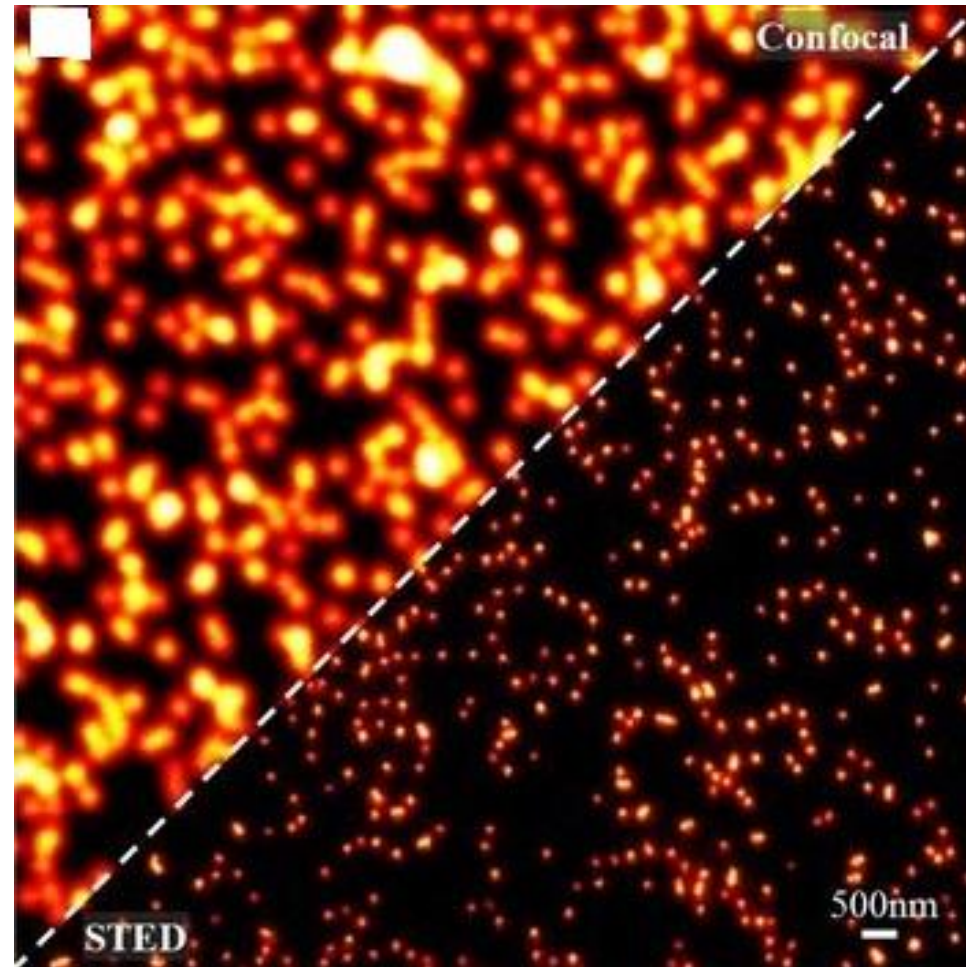
# STED microscopy



# Experimental setup



## Confocal versus STED microscopy





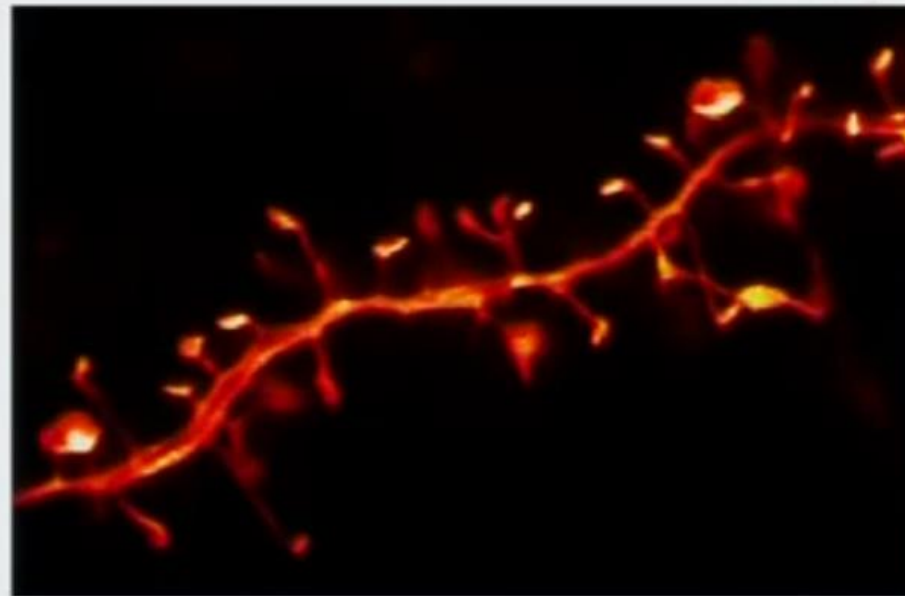
# Applications of STED microscopy

## Neurophysiology

Dendritic Spines [Living Neuron](#)

Hippocampal organotypic slices

CA1 pyramidal neurons

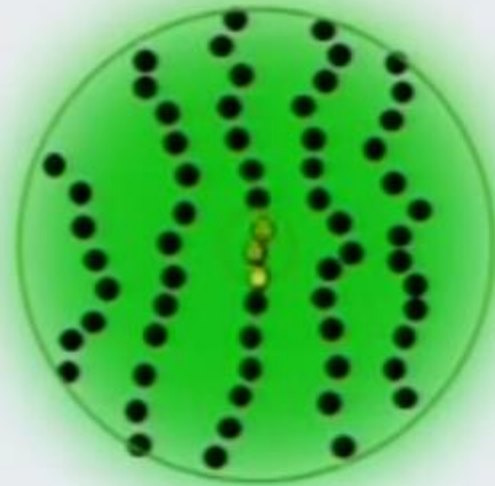


Urban, Willig, Hell, Nägerl, *Biophys J* (2011)

2 $\mu$ m

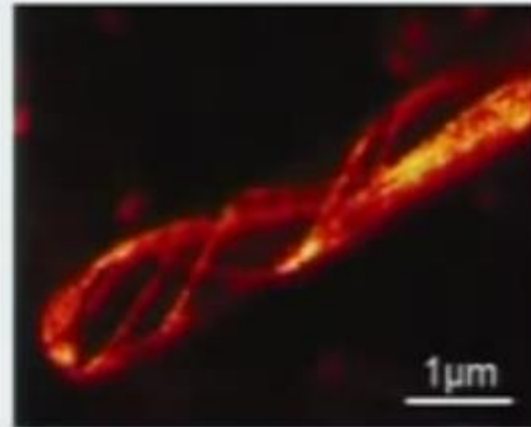
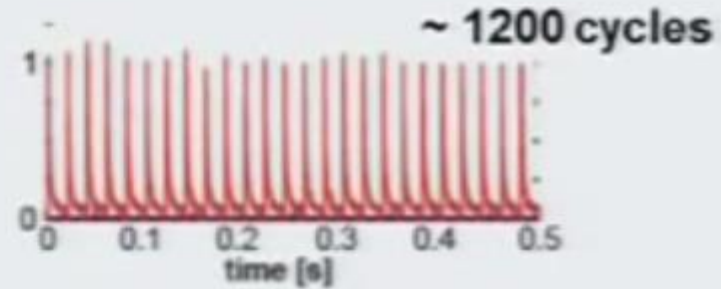
# STED, RESOLFT microscopy

Hell, Jakobs, Kastrup, *Appl Phys A* (2003)  
 Hell, *Nat Biotech* (2003)  
 Hofmann et al, *PNAS* (2005)



~ 10<sup>5</sup> times lower  
 intensity than in STED

$$d \cong \frac{\lambda}{2n \sin \alpha \sqrt{1 + I/I_2}}$$



Grotjohann, Testa et al, *Nature* (2011)  
 Brakemann et al, *Nat Biotech* (2011)

$$I_2 = 1/(\sigma\tau) \sim \text{W/cm}^2$$

# STED, RESOLFT microscopy

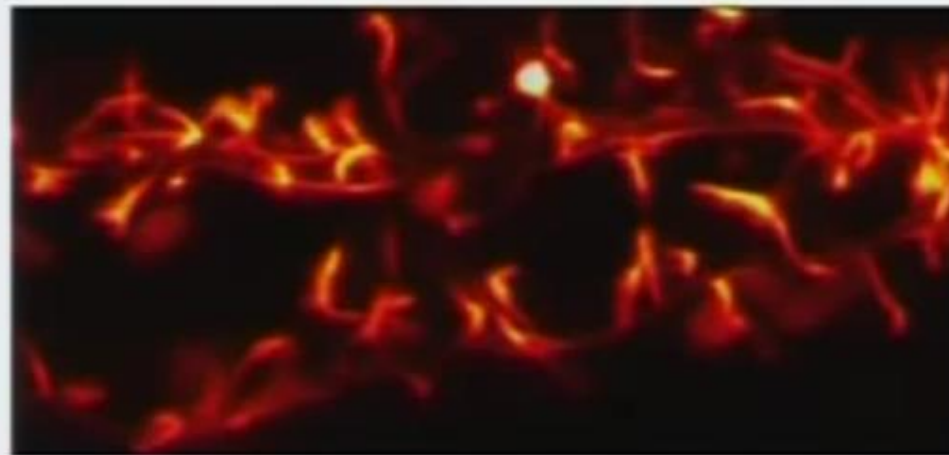
Hell, Jakobs, Kastrup, *Appl Phys A* (2003)

Hell, *Nat Biotech* (2003)

Hofmann et al, *PNAS* (2005)

*2 hours continual scanning, no visible photodamage.*

Living neuron, hippocampal organotyp slice, F-Actin

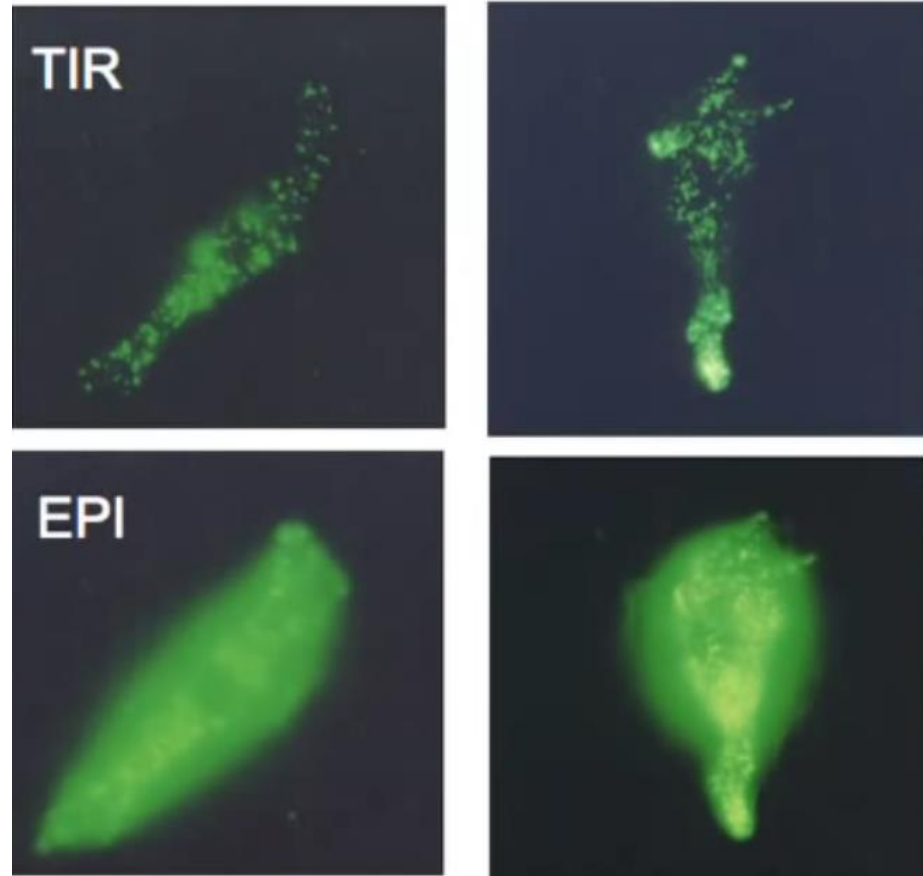


1  $\mu\text{m}$

Testa, Urban et al, *Neuron* (2012)

# Total internal reflection fluorescence (TIRF) microscopy

Objective-based TIRF with NA=1.65  
GFP-marked chromaffin granules



# Principles of TIRF microscopy

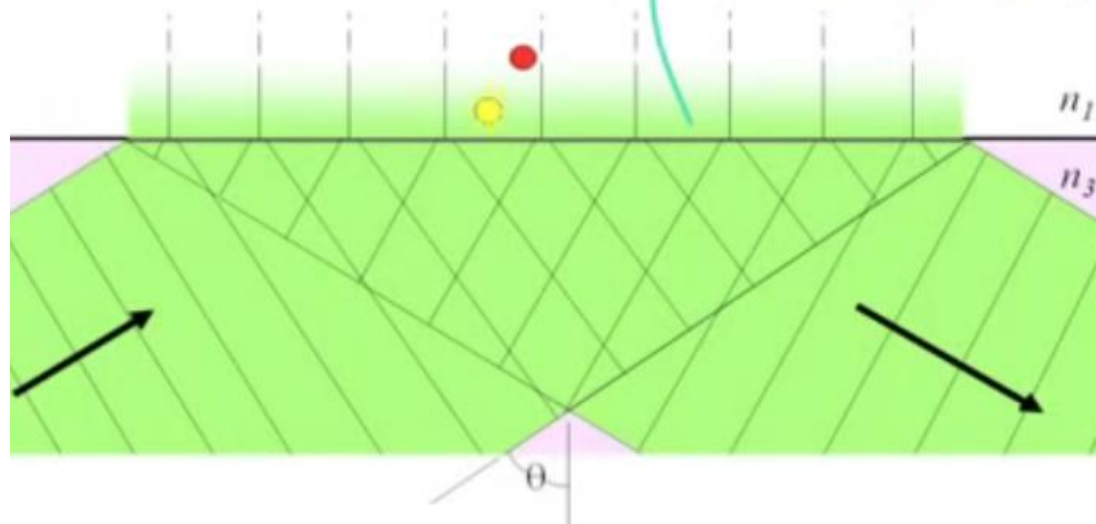
Evanescent field produced by TIR

$$I(z) = I(0) e^{-z/d}$$

$$d = \frac{\lambda_o}{4\pi} \left( n_3^2 \sin^2 \theta - n_1^2 \right)^{1/2}$$

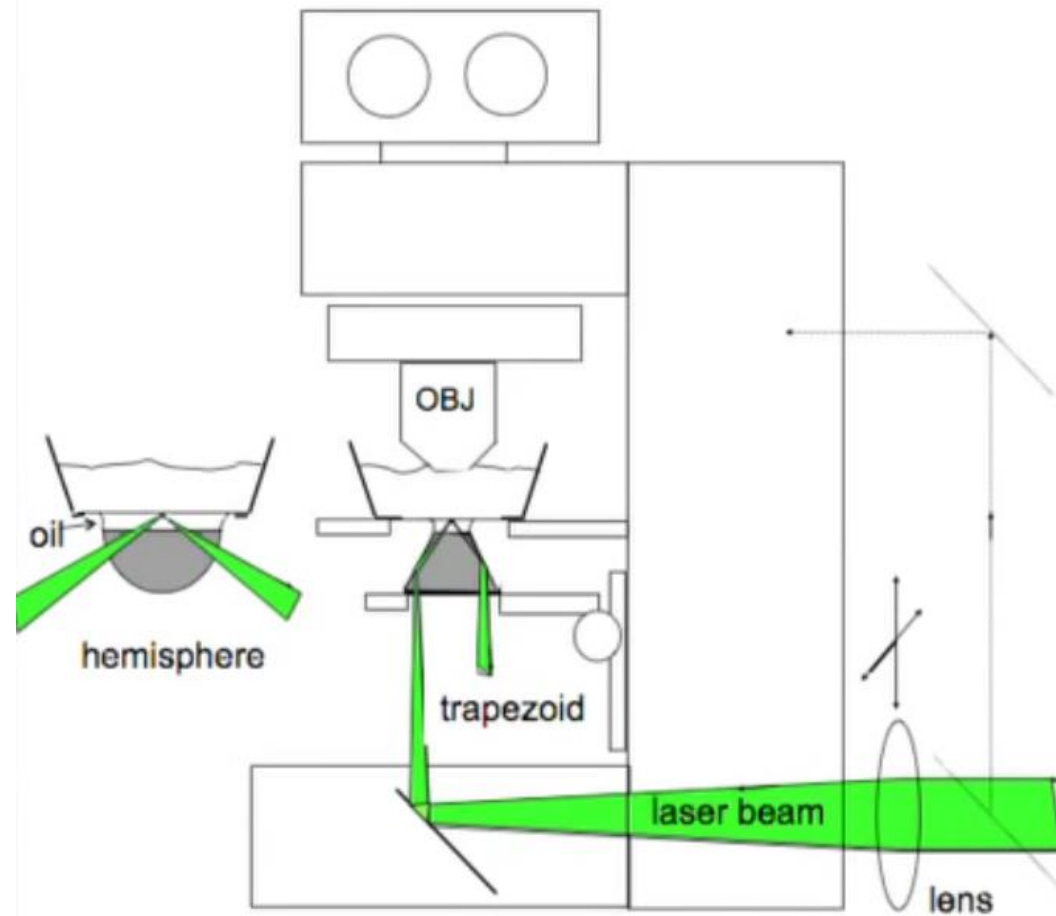
$$d \leq \sim 100 \text{ nm}$$

$I(0)$  similar to  
incident intensity



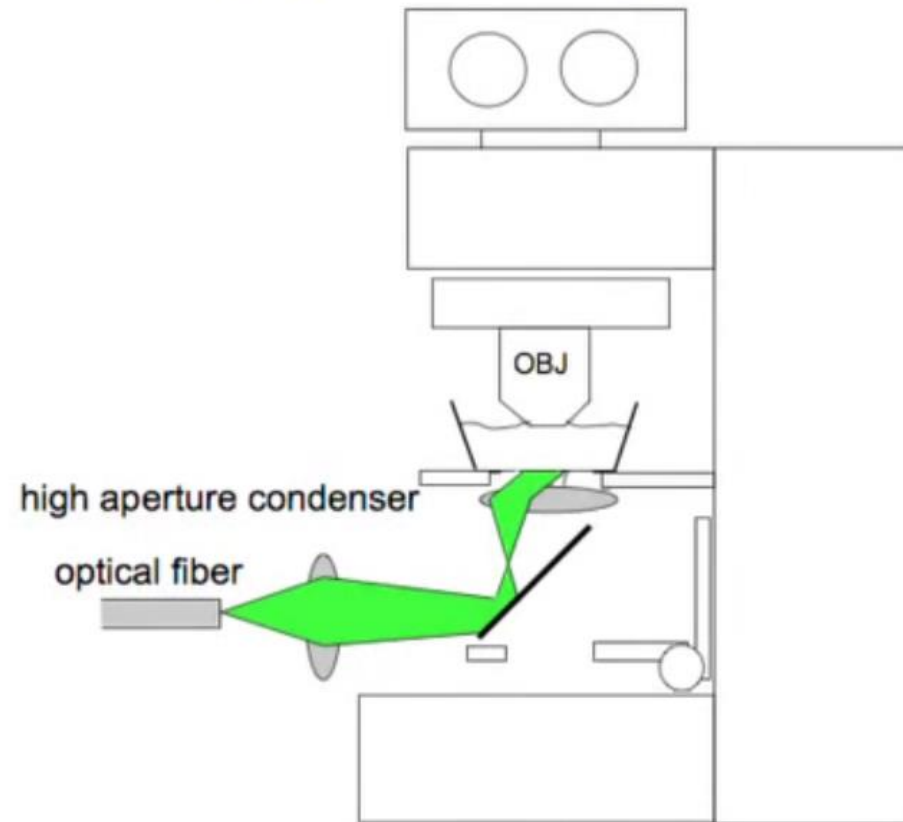
# Experimental setup

## Prism-based TIRF for upright microscope



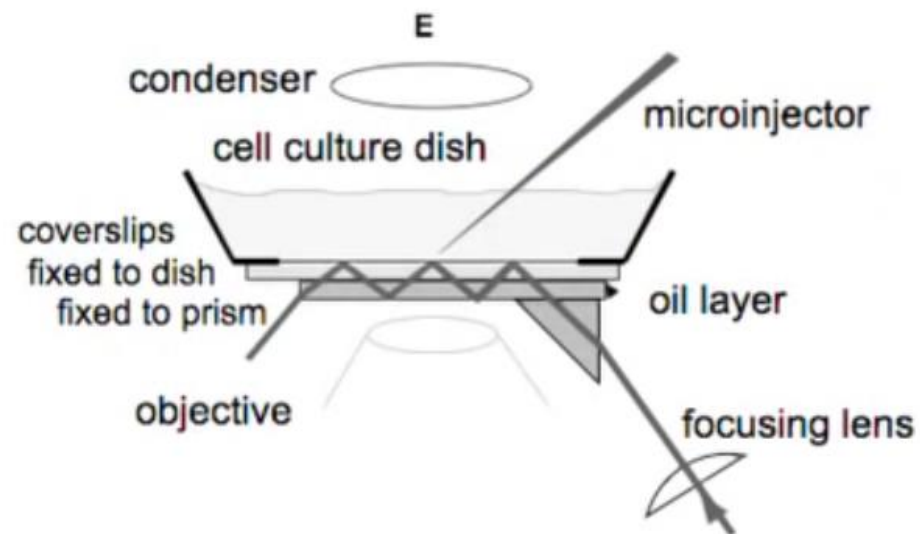
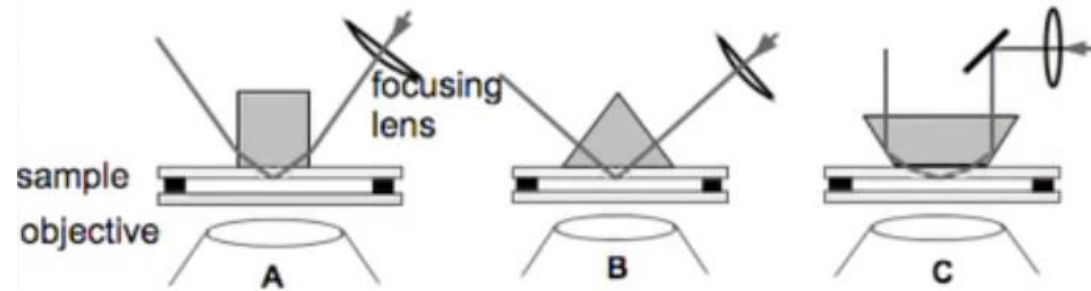
# Experimental setup

## Condenser-based TIRF for upright microscope



# Experimental setup

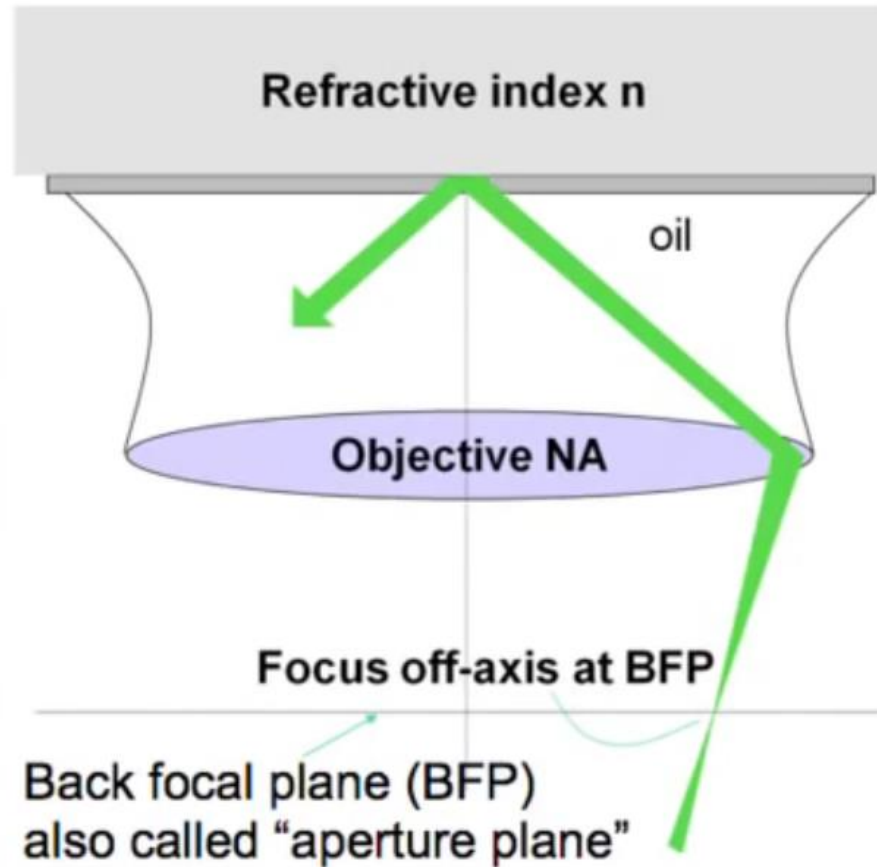
## Prism-based TIRF configurations





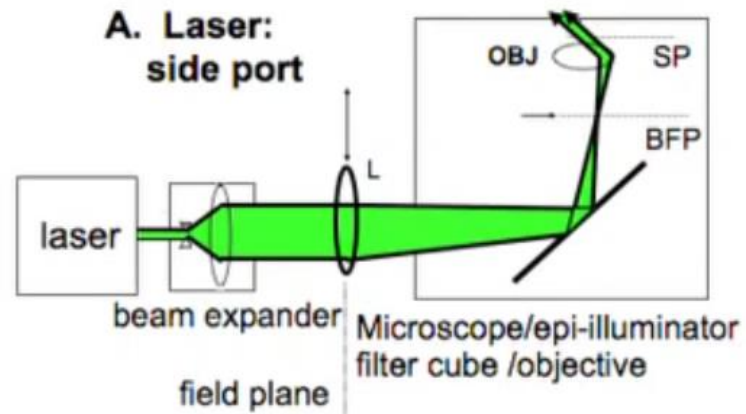
# Experimental setup

Objective-based (prismless)  
TIRFM - **requires**  $NA > n$

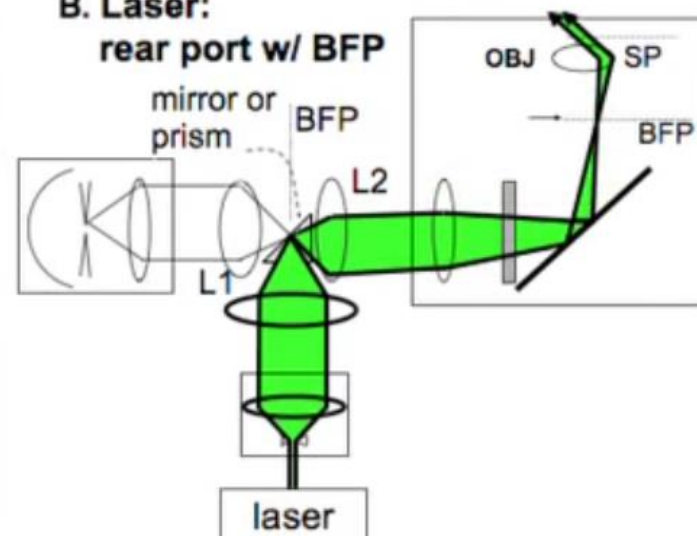


# Experimental setup

## Objective-based TIRFM

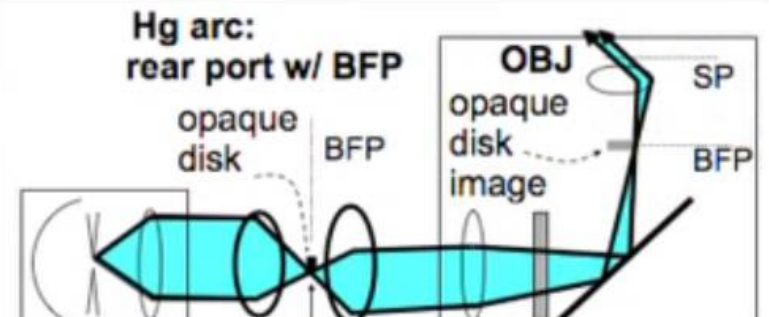
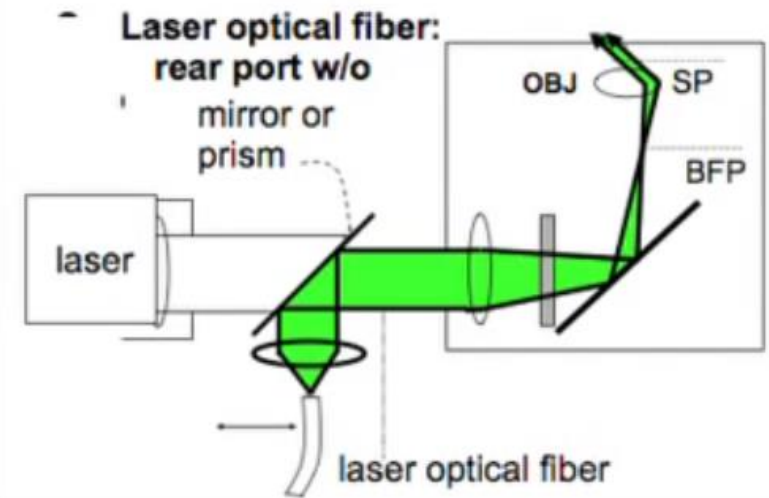


**B. Laser: rear port w/ BFP**



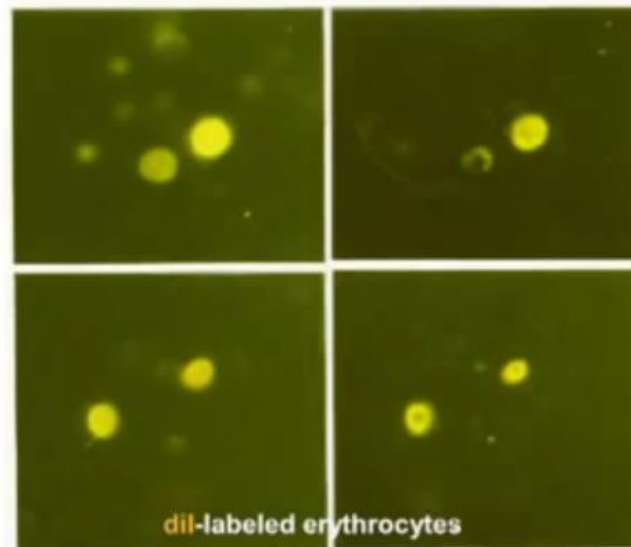
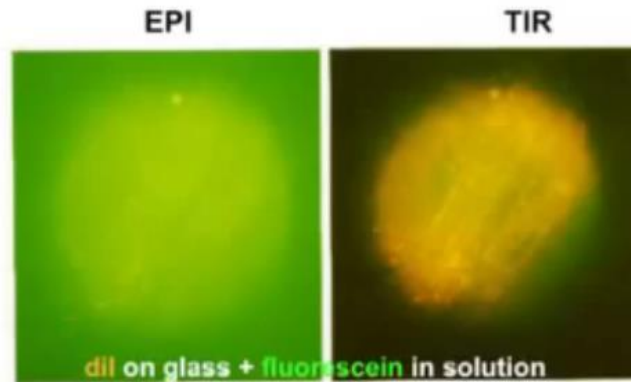
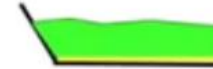
# Experimental setup

## Objective-based TIRFM



# Applications

TIRF with an ARC LAMP source



## Summary

TIRFM is useful to study the **cellular dynamics** of:

1. Small random motions of organelles toward or away from the membrane.
2. Submembrane events: exocytosis, cytoskeletal dynamics
3. Submicroscopic membrane folding and indentations
4. Kinetic rates of association/dissociation at the membrane, even in continuous presence of fluorophore in bath.

TIRFM is also useful in **surface biochemistry**:

1. Single molecule fluorescence (low background)
2. Chemical kinetics and diffusion at surfaces

**Thanks**