

Elettra Sincrotrone Trieste



X-ray imaging and spectromicroscopy using XAS and XRF

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Outline

- X-ray microscopes: Full-field, STXM, projection
- Standard imaging technique and new advanced ones (CDI, Ptychography)
- Special attention to the TwinMic beamline (Elettra, Trieste)
- Multidisciplinary applications: life sciences, food science, environmental science, materials science, cultural heritage...

X-ray microscopy types



X-ray microscopy: <u>bridge</u> between <u>visible light</u> microscopy and <u>electron</u> microscopies

Spatial resolution:

visible light microscopy < X-ray microscopy < **<u>electron</u>** microscopy

Air Vacuum (or air) Vacuum

X-ray Microscopy vs Electron Microscopy:

- Easier sample preparation (no metalisation)
- Higher penetration depth of X-rays compared to electrons
 thicker samples can be analysed



Scheme of a projection microscope with the source to specimen plane distance R_1 and specimen to detector distance R_2



Kaulich B., Thibault P., Gianoncelli A., Kiskinova M. "Transmission and emission x-ray microscopy: operation modes, contrast mechanisms and applications" *Journal of Physics: Condensed Matter*, Vol. 23 - 8, pp. 083002 (2011)

Projection microscopy





3D phase contrast dataset view, reconstructed cross section of part of a ceramisphere (projection X-ray microscope at the CSIRO, Australia).

The total collection time of the microtomogram was 10 h. The diameter of the sphere is 110 μ m

A state-of-the art instrument for projection microscopy, including phase-sensitive imaging and microtomography based on a converted scanning electron microscope, is reported for example by Mayo et al. (Mayo et al 2002; Mayo et al 2003).

Mayo S C, Miller P R, Wilkins S W, Davis T J, Gao D, Gureyev T E, Paganin D, Parry D J, Pogany A and Stevenson A W 2003 *Journal De Physique IV* 104 543-546





Full field Imaging mode



- Similar to conventional visible light microscope
- Analysis of morphology in transmission
- Fast imaging, dynamics, microtomography



Full-field X-ray imaging or "one shot" X-ray image acquisition can be considered as the optical analogon to a visible light transmission microscope

BUT

Refractive index n is very close to unity and smaller than unity!!!

 $n = 1 - \delta(\lambda) - i\beta(\lambda) < 1$

Background info: X-ray microscopy types





- + versatile detectors can run simultaneously;
- + easier optics set-up;
- long exposure time;
- complex electronics.

Ideal for spectromicroscopy

- + short exposure time;
- + higher resolution
- static system;
- complex optical alignment.

Ideal for dynamic studies and tomography



Background info: Diffraction by a grating





The complex refractive index

$$n = 1 - \frac{n_a r_e \lambda^2}{\pi} (f_1 + i f_2) \equiv 1 - \delta + i\beta \le 1$$

"Conventional refractive index" describing phase change:

$$\varphi(z) = \frac{2\pi}{\lambda} \, \delta z$$

Exploitation of phase contrasts possible using X-rays ? Lower radiation damage ? Describing photoelectric absorption with coefficient:

$$\mu = \frac{4\pi}{\lambda}\beta$$

Consequence: Emission of Auger, photo-electrons and fluorescence photons, but also causes radiation damage (energetic secondary electrons!)



Delta versus beta



Delta is orders of magnitude larger !!!



Absorption mode

X-ray photons are selectively absorbed by the material according to its density and thickness (ex. radiography)



Beer - Lambert's law: I = $I_0 e^{-mx}$

Phase contrast mode

Absorption can produce little contrast for light (transparent) materials or for materials with similar atomic number (similar attenuation factors).

Moreover as the energy increases the contrast diminishes (absorption coefficient $\propto 1/E^3$)

Phase contrast is more sensitive to edges and borders in the sample

Contrast techniques using the real, phase-shifting part of the complex refractive index are in many cases superior to absorption contrast because:

- (i) the x-ray dose can be reduced dramatically
- (ii) the throughput is higher (the phase shift

dominates the absorption in the x-ray regime)



Full field Imaging mode



- Similar to conventional visible light microscope
- Analysis of morphology in transmission
- Fast imaging, dynamics, microtomography





Resolution tests in full-field imaging

ZP parameters: 110 μm diameter 50 nm outer zones f=3.2 mm @ 720 eV fabricated by TASC/ INFM



2<u>μ</u>m

Test pattern with 30 nm features (fabricated by TASC/ INFM)

2 µm

Experiment performed by M. Prasciolu and D. Cojoc, TASC/ INFM)

Natural amplitude contrast between water and organic matter

X-ray energy (eV) 500 í000 1500 10.0 X-ravs 1/μ (water) Penetration distance (µm) edge Oxygen 1.0 Electrons 1/µ (protein) edge (water) (protein Sarbon America (protein) 0.1 100 200 300 400 0 Electron energy (keV)

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The "Water Window":

Due to dramatic difference in the f2 values of two materials, especially water and organic matter between the C and O K-absorption edges.

Note the penetration distance compared to electrons !!!

H. Wolter: Spiegelsysteme streifenden Einfalls als abbildende Optiken fuer Roentgenstrahlen, Ann. Phys. 10, 94-114, 286 (1952)

Environmental science: Imaging in liquids



Bacteria and clay dispersion: Destruction of associations of clay particles by soil microbes



X-ray images acquired with the full-field imaging microscope at BESSY I @ 520 eV

Samples analysed in the natural hydrated state: \rightarrow no alteration of the environment of the sample

J. Thieme et al., IRP, Uni Goettingen / G. Machulla, Uni Halle, D



Across edge imaging



Discontinuities due to absorption

The absorption occures when the incoming X-rays are matching the electron binding energies

Absorption edges are fingerprints \Rightarrow they can be used to identify the chemical elements

By taking two images, one above and one below a specific absorption edge, the correspondent chemical element will give a high contrast difference in the two images

Brightfield imaging at higher photon energies



Characterization of morphology and defects in modern semiconductors with a full-field imaging microscope (@ 1.8 keV, XM1/ ALS)

Sample preparation: Back side thinning of Si wafer





G. Schneider et al., BESSY II



Environmental science: Analysis of air particulate matter



P. Barbieri et al., Dept. of Chem., Univ. Triest





Darkfield or darkground imaging



Darkfield illumination requires blocking out of the central light which ordinarily passes through and around (surrounding) the specimen, allowing only oblique rays from every azimuth to "strike" the specimen.



Visible light micrographs of silica skeletons from a small marine protozoan (radiolarian)

Darkfield imaging in scanning X-ray microscopy





Brightfield image of a cell with Au labelling spheres overlayed with a darkfield image

Images acquired with STXM at the NSLS

Technique is especially suited for small, strongly scattering particles as for example a few 10nm diameter labelling spheres



S. Vogt, M.A. thesis, SUNY Stony Brook (1997).



Detector based contrast technologies in scanning X-ray microscopy:





Computational extraction of contrasts by masking:



Raw data acquisition of first diffraction order image for each pixel of the raster scan

Applying different masks



Bright field







Darkfield

A. Gianoncelli et al., Appl. Phys. Lett.

Principle: Differential phase contrast





- The detector can be split into several elements
- The sum signal gives the incoherent bright-field signal
- Anti-symmetric signal combinations relate to the *phase gradient* of the object transmittance.





Marine biology: Imaging of giant diatoms





Planktonic diatom "Casciodiscus sp." (provided by LBM, Trieste, I)



Bright field imageDPC mode – X-momentImages acquired in STXM mode with FRCCD camera; E=1320 eV, 200x190 px, 50ms dwell/px

B. Kaulich et al., JOSA A **19** (4), 797-806 (2002)

Brightfield and differential phase contrast images acquired simultaneously with configured detector



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ffee bean cell membranes







B. Bonnlaender, F. Sicilia, Illy AromaLab, et al.



Ptychography (CDI)

- Ptychography is rapidly developing into an important imaging tool in X-ray microscopy.
- It doesn't require X-ray Optics
- The technique involves successively illuminating overlapping regions of a specimen with a coherent probe and recording the resulting diffraction patterns.
- It is important that the illuminated areas overlap significantly since those common regions provide duplicate information that allows computer algorithms to reconstruct reliably both the sample transmission function and the illuminating probe from the measured diffraction patterns.



Ptychography (CDI)



- K. Giewekemeyer, M. Beckers, T. Gorniak, M. Grunze, T. Salditt, and A. Rosenhahn Optics Express Vol. 19, Issue 2, pp. 1037-1050 (2011)
- D. A. Shapiro, Y.-S. Yu, T. Tyliszczak, J. Cabana, R. Celestre, W. Chao, K. Kaznatcheev, A. L. D. Kilcoyne, F. Maia, S. Marchesini, Y. Shirley Meng, T. Warwick, L. Lisheng Yang, H. A. Padmore Nature Photonics 8, 765–769 (2014)



200

100

Ptychography Algorithms

Start Guess Object



Processed Data

Probe from test object

0.6 0.4



Soft X-ray spectromicroscopy using ptychography with randomly phased illumination



A. M. Maiden, G. R. Morrison, B. Kaulich, A. Gianoncelli, J. M. Rodenburg, *"Soft X-ray spectromicroscopy using ptychography with randomly phased illumination"*, Nature Communications 4, 1669, (2013)



Soft X-ray spectromicroscopy using ptychography with randomly phased illumination



Balb/3T3 mouse fibroblast cells that had been exposed to cobalt ferrite ($CoFe_2O_4$) nanoparticles

Ptychography reconstruction using the ePIE algorithm (Uni of Sheffiled)



STXM absorption image

A. M. Maiden, G. R. Morrison, B. Kaulich, A. Gianoncelli, J. M. Rodenburg, *"Soft X-ray spectromicroscopy using ptychography with randomly phased illumination"*, Nature Communications 4, 1669, (2013)



Soft X-ray spectromicroscopy using ptychography



First direct measurements from cobalt ferrite nanoparticles of the phase variations across the iron L edge, with the phase variations the observed variations in modulus contrast across FeL₃ edge are consistent with estimates based on total showing (stronger) and sclear enaferatures than the modulus data provided

A. M. Maiden, G. R. Morrison, B. Kaulich, A. Gianoncelli, J. M. Rodenburg, *"Soft X-ray spectromicroscopy using ptychography with randomly phased illumination"*, Nature Communications 4, 1669, (2013)



SXRI Beamline





Active position stabilitisation: ~17 nm RMS stability - no thermal drift



fCDI





ZP: Diameter 160µm, Central stop 30µm, Outermost zone 30nm


Preliminary tests

Diffraction pattern (through unfocused ZP)



Reconstruction



G. Kourousias, B. Bozzini, A. Gianoncelli, M. W. M. Jones, M. Junker, G. van Riessen, M. Kiskinova "Shedding light on electrodeposition dynamics tracked in situ via soft X-ray coherent diffraction imaging" **Nano Research**, 9 – 7, 2046-2056 (2016)

Scanning and oversampling







G. Kourousias, B. Bozzini, A. Gianoncelli, M. W. M. Jones, M. Junker, G. van Riessen, M. Kiskinova "Shedding light on electrodeposition dynamics tracked in situ via soft X-ray coherent diffraction imaging" **Nano Research**, 9 – 7, 2046-2056 (2016)



Electrochemical processes at local nanoscales by in situ soft FCDI imaging TwinMic @ Elettra – SXRI @ Australian Synchrotron



G. Kourousias, B. Bozzini, A. Gianoncelli, M. W. M. Jones, M. Junker, G. van Riessen, M. Kiskinova "Shedding light on electrodeposition dynamics tracked in situ via soft X-ray coherent diffraction imaging" **Nano Research**, 9 – 7, 2046-2056 (2016)



Spectroscopy across Mn edge through Ptycography



G. Kourousias, B. Bozzini, A. Gianoncelli, M. W. M. Jones, M. Junker, G. van Riessen, M. Kiskinova "Shedding light on electrodeposition dynamics tracked in situ via soft X-ray coherent diffraction imaging" **Nano Research**, 9 – 7, 2046-2056 (2016)



TwinMic: Integration of both imaging modes into a single instrument





The European team that initiated the project

- Morphological analysis, XANES and AAEI
- Different contrasts incl. brightfield, differential phase and interference contrast, darkfield, etc
- Versatile specimen environment



TwinMic microscope 400 – 2200 eV

TwinMic – Combination of scanning and full-field imaging in a single instrument





- •Biotechnology
- Nanotechnology
- •Environment
- •Geochemistry
- •Food Science
- Medicine
- Pharmacology
- •Cultural Heritage
- •New Materials



Scanning X-ray microscope (STXM)





STXM mode

Differential phase contrast with a fast read-out CCD camera



Morrison, G. et al., IPAP Conf. Series 7, 377-379 (2006) Gianoncelli A. et al., Appl Phys Lett 89, 251117 (2006)



Simultaneous acquisition of:

- Absorption or transmission
- Differential phase contrast
- Darkfield images



Photoionization







X-ray absorption (through photoelectric effect)

The primary X-ray photon causes the ejection of electrons from the inner shells, creating vacancies

X-ray Fluorescence

The vacancy created by the primary X-ray photon is filled by an electron coming from an outer shell causing the emission of a characteristic X-ray photon whose energy is the difference between the two binding energies of the corresponding shells

Auger effect

The vacancy created by the primary X-ray photon is filled by an electron coming from an outer shell and the energy is transferred directly to one of the outer electrons, causing it to be ejected from the atom.



LEXRF

Low-energy X-ray fluorescence for elemental analysis:



A. Gianoncelli, B. Kaulich, M. Kiskinova, R. Alberti, T. Klatka, A. Longoni, A. de Marco, A. Marcello, Simultaneous Soft X-ray Transmission and Emission Microscopy, Nucl. Instr. and Meth. A 608 (1), 195-198



Detecting trace elements:

X-ray fluorescence: ~1000x better sensitivity than electrons for trace elemental mapping (ion concentrations etc.).

Low fluorescence yields for soft X-rays! !!



LEXRF

Low-energy X-ray fluorescence:



TwinMic LEXRF spectrum with unfocused beam of a test organic matrix on a metal shim

Dynamic range: up to 30 kcounts/s

Average FWHM energy resolution @ C- K edge: 69 eV





A. Gianoncelli, B. Kaulich, M. Kiskinova, R. Alberti, T. Klatka, A. Longoni, A. de Marco, A. Marcello, Simultaneous Soft X-ray Transmission and Emission Microscopy, Nucl. Instr. and Meth. A 608 (1), 195-198





LEXRF





Aluminium toxicity

Soluble Al – "the most important growth-limiting factor for plants in most strongly acid soils and mine spoils" **Foy (1984**)

Acid soils occupy ~ 40 billion hectares (~ 30 %) of the world's ice free land area **von Uexküll and Mutert (1995**)

In Australia alone, acid soils cost \$1.5 billion p.a. in lost productivity





Aluminium toxicity

Soluble Al – "the most important growth-limiting factor for plants in most strongly acid soils and mine spoils" **Foy (1984**)

Although known since 1904 that Al is the primary factor causing a reduction in plant root growth in acid soils, the mechanism by which Al is toxic remains unclear

Recent research (2014) has shown that Al exerts its toxic effects very quickly, reducing root growth in \leq 30 min. Therefore, a crucial step in elucidating how Al exerts its toxic effects is to examine where the Al is accumulating within the roots







Aluminum toxicity

30 minutes, 6 mm, Sample 1

P. M. Kopittke, K. L. Moore, E. Lombi et al, "Identification of the Primary Lesion of Toxic Aluminum in Plant Roots" Plant Physiology, 2015, 167, 140



7 μm-thick transverse cross section of soybean roots

Exposed to 30 µM AI for 0.5 h.





P. M. Kopittke, K. L. Moore , E. Lombi et al, "Identification of the Primary Lesion of Toxic Aluminum in Plant Roots" Plant Physiology, 2015, 167, 140



Soybean roots exposed to 30 μM Al for 24 h

Al



P. M. Kopittke, K. L. Moore , E. Lombi et al, "Identification of the Primary Lesion of Toxic Aluminum in Plant Roots" Plant Physiology, 2015, 167, 140



Soybean roots exposed to 30 μM Al for 0.5 h



P.M. Kopittke et al "Identification of the Primary Lesion of Toxic Aluminum in Plant Roots" Plant Physiology 2015, 167, 140



Soybean roots exposed to 30 μM Al for 0.5 h

Al

Root growth decreased by 25 % after 90 min at 10 μ M Al or only 5 min at 75 μ M Al.

This rapid effect was caused by AI binding strongly to the cell walls, thereby inhibiting loosening as required for root elongation.

These findings show the importance of focusing on traits related to cell wall composition as well as mechanisms involved in wall loosening to overcome the deleterious effects of soluble Al

P.M. Kopittke et al "Identification of the Primary Lesion of Toxic Aluminum in Plant Roots" Plant Physiology 2015, 167, 140



Al in wheat

- In wheat (Triticum aestivum), it is commonly assumed that Al is detoxified by the release of organic anions into the rhizosphere, but it is also possible that detoxification occurs within the apoplast and symplast of the root itself
- Rhizosphere = plant-root interface, not well definable in size or shape





XRF analyses @ TwinMic



FIGURE 5 | The distribution of AI, examined using LEXRF, in 5-µm-thick transverse root sections taken 3 mm from the apex of ES8 and ET8 exposed to 3.5 or 50 µM AI for either 3 h (A,C,E) or 48 h (B,D,F). In all cases, the rhizodermis (and exterior of the root) is on the right-hand side of the image, with only the rhizodermis and 1–2 layers of cortical cells shown. The signal intensity is presented as a color scale, with brighter colors corresponding to higher concentrations. All images were scaled to the same values, and hence intensities can be compared between images. The scale-bar in (D) applies to all images. See Supplementary Figure 1 for corresponding light micrographs.



XANES on Al edge (@ CLS)



FIGURE 6 *In situ* analyses of AI speciation using synchrotron-based X-ray absorption near edge structure (XANES). **(A)** AI K-edge XANES spectra of five standard compounds and for root tissues of the near-isogenic wheat lines where ET8 was exposed to 50 μM AI for 3 or 48 h and ES8 was exposed to 3.5 μM AI for 48 h. **(B)** Enlarged spectra (1,562–1,575 eV) for the three root tissues and two standards of interest, AI-malate and AI-pectin. The dotted lines are provided for reference, being 1,566 eV (corresponding to the strong single maximum of four-fold coordinated AI, such as for AI-phosphate), 1,567.7 eV (corresponding to the six-fold coordinated peak of γ-AI₂O₃).

- ET8 exposed for 3 h being a mixture of Al species, but mainly Al organic species including that of Al-malate and Al-pectin
- After exposure to Al for 48 h, much of the Al was four-fold coordinated (particularly for ES8, with ET8 still having a higher proportion of six-fold coordinated Al), although the exact form of this four-fold coordinated Al is not clear and further work is required.



Conclusions

- When ES8 and ET8 were grown at 3.5 mM Al, roots of ES8 accumulated more Al than did those of ET8 (as is evident when comparing Figs 5A,B with Figs 5C,D).
- However, when grown at Al concentrations that resulted in the same reduction in RER (root elongation rate), root tissues of ET8 tended to accumulate more Al than did those of ES8
- The concentration of Al was highest in the rhizodermis in all six treatments, decreasing in the outer cortical tissues and even more so in the inner cortex.
- Furthermore, Al accumulated primarily in the cell wall in apical root tissues of both ES8 and ET8, with comparatively small amounts of Al found within the symplast. This indicates that Al complexed by malate within the root tissues of ET8 accumulated primarily within the apoplast of the rhizodermis and outer cortex.
- Roots, particularly for ES8, ruptured when exposed to high concentrations of Al this having been observed previously in the roots of a wide range of plant species. These ruptures form initially in the elongation where Al was found to accumulate to high concentrations. This reaffirms the importance of Al accumulation in the cell walls, thereby decreasing root elongation by rapidly inhibiting the ability of the walls to loosen.
- Thus, it is apparent that the complexation of Al by malate in ET8, both in the rhizosphere and apoplast, reduces the strong binding of Al to the cell wall, and thereby reduces the damaging interactions of Al with the root cells.



The twin X-ray microscopy station @ Elettra

Food Science: Inside the wheat



Ivan Kreft, University Ljubljana

Functionality and toxicity of Zn in wheat and buckwhe analyzed on subcellular level





Structure of a wheat grain



M. Regvar, D. Eichert, B. Kaulich, A. Gianoncelli, P. Pongrac, K. Vogel-Mikus, I. Kreft, New insights into globoids of protein storage vacuoles in wheat aleurone using synchrotron soft X-ray microscopy, Journal of Experimental Botany, Vol. 62, No. 11, 3929–3939, 2011.





Ivan Kreft, Fac. of Biotechnology, University Ljubljana

Functionality and toxicity of Zn in wheat and buckwheat analyzed on subcellular level

Healthy control wheat

E=1686 eV 80 x 80 mm² 80 x 80 px 8 s dwell/ px 1 mm resolution 4 detectors



M. Regvar, D. Eichert, B. Kaulich, A. Gianoncelli, P. Pongrac, K. Vogel-Mikus, I. Kreft, New insights into globoids of protein storage vacuoles in wheat aleurone using synchrotron soft X-ray microscopy, Journal of Experimental Botany, Vol. 62, No. 11, 3929–3939, 2011.



Biogenetics and Food Science: Inside the wheat



Ivan Kreft, Fac. of Biotechnology University Ljubljana

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Nanotoxicology

Cellular distribution and degradation of CoFe₂O₄ NPs in Balb/3T3 Fibroblast cells

Localization of engineered nanoparticles (ENPs) inside a cell and on the possible effects on the cell metabolic behaviour

DPC BF Co/C Fe/

 $CoFe_2O_4$ in mouse 3T3 fibroblast cells, E=2019 eV, 60um x 60 um

P. Marmorato, G. Ceccone, A. Gianoncelli, L. Pascolo, J. Ponti, F. Rossi, M. Salomé, B. Kaulich, and M. Kiskinova, *Cellular distribution and degradation of Cobalt Ferrite Nanoparticles in Balb/3T3 Fibroblasts*, Toxicology Letters, 2011, 207 - 2, 128-136.

G. Ceccone, P. Marmorato et al., EC Joint Research Center, Ispra, I



Balb/3T3 exposed to 1000mM



Similar behaviour (but less evident) in the nuclear region for 500µM concentration



Energy [keV]

P. Marmorato, G. Ceccone, A. Gianoncelli, L. Pascolo, J. Ponti, F. Rossi, M. Salomé, B. Kaulich, and M. Kiskinova, *Cellular distribution and degradation of Cobalt Ferrite Nanoparticles in Balb/3T3 Fibroblasts*, Toxicology Letters, 2011, 207 - 2, 128-136.





Control

40 µM

 $250 \ \mu M$

500 µM

High-resolution scanning transmission soft X-ray microscopy for rapid probing of nanoparticle distribution and sufferance features in exposed cells Kourousias G, Pascolo L, Marmorato P, Ponti J, Ceccone G, Kiskinova M, Gianoncelli A X-Ray Spectrometry (2015)



Fibroblast cells exposed to CoFe₂O₄ NPs 60um x 40um, 480x320 pixels, 20ms dt, 900eV

32um x 60um, 256x480 pixels, 20ms dt, 900eV

Spot size: 135nm

High-resolution scanning transmission soft X-ray microscopy for rapid probing of nanoparticle distribution and sufferance features in exposed cells Kourousias G, Pascolo L, Marmorato P, Ponti J, Ceccone G, Kiskinova M, Gianoncelli A *X-Ray Spectrometry (2015)*



60um x 40um, 480x320 pixels, 20ms dt, 900eV

32um x 60um, 256x480 pixels, 20ms dt, 900eV



Х

Spot size: 135nm

High-resolution scanning transmission soft X-ray microscopy for rapid probing of nanoparticle distribution and sufferance features in exposed cells Kourousias G, Pascolo L, Marmorato P, Ponti J, Ceccone G, Kiskinova M, Gianoncelli A X-Ray Spectrometry (2015)



Red oil specifically stains lipids



Fig. 6. Optical images of Balb/3T3 control cells (a and b) and incubated for 24 h with 500 μM CoFe₂O₄ NPs suspension (c and d). Red spots represent lipids stained by Red Oil O solution (b and d) whilst nuclei are stained in blue by Hoechst. Bar = 10 μm. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of the article.)

P. Marmorato, G. Ceccone, A. Gianoncelli, L. Pascolo, J. Ponti, F. Rossi, M. Salomé, B. Kaulich, and M. Kiskinova, *Cellular distribution and degradation of Cobalt Ferrite Nanoparticles in Balb/3T3 Fibroblasts*, Toxicology Letters, 2011, 207 - 2, 128-136.



Nanotoxicology: CoFe₂O₄ ENPs

Control



Exposed to 500µM



Ca

24 21

18

15

12

Exposed to 40uM



P. Marmorato, G. Ceccone, A. Gianoncelli, L. Pascolo, J. Ponti, F. Rossi, M. Salomé, B. Kaulich, and M. Kiskinova, Cellular distribution and degradation of Cobalt Ferrite Nanoparticles in Balb/3T3 Fibroblasts, in press in Toxicology Letters



Exposure to Asbestos



L. Pascolo, M. Melato, Burlo Hospital, Trieste, Italy

Mesothelioma and differentiation of lung tissue due to asbestos; the role of Mg



E=2019 eV, 50mm x 50 mm, 100 x 100 pixels, 15s/pixel LEXRF, 4 SDDs

L. Pascolo, A. Gianoncelli, et al. Particle and Fibre Toxicology 2011, 8:7. L. Pascolo, A. Gianoncelli, et al. Scientific Reports 2013, 3.



Tissue with a phagocytated asbestos fibre.



L. Pascolo, A. Gianoncelli, et al. Particle and Fibre Toxicology 2011, 8:7. L. Pascolo, A. Gianoncelli, et al. Scientific Reports 2013, 3.



Fe K-edge XANES measured in selected ~ 1 mm² spots of an asbestos body

- Most of the Fe detected around asbestos fibres (coating and ferruginous bodies) is compatible with the presence of <u>ferritin</u> and the Fe3+ oxidation state of iron.
- The most novel and intriguing result was the detection of significant percentages of <u>haematite</u> in the asbestos bodies that we suppose is the results of ferritin transformation occurring during the long residence time in the asbestos bodies in the lung tissues.



L. Pascolo, A. Gianoncelli, et al. Particle and Fibre Toxicology 2011, 8:7. L. Pascolo, A. Gianoncelli, et al. Scientific Reports 2013, 3.






OPEN

First real Clinical case at TwinMic

SUBJECT AREAS: PATHOGENESIS BIOPHYSICS CHEMICAL MODIFICATION

> Received 19 May 2014 Accepted 9 September 2014 Published 7 October 2014

Calcium micro-depositions in jugular truncular venous malformations revealed by Synchrotron-based XRF imaging

Lorella Pascolo¹, Alessandra Gianoncelli², Clara Rizzardi³, Veronica Tisato⁴, Murielle Salomé⁵, Carla Calligaro⁶, Fabrizio Salvi⁷, David Paterson⁸ & Paolo Zamboni⁹

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Started at TwinMic Then extended to 2 other synchrotron facilities to get complementary information:

- ESRF (ID21)
- Australian Synchrotron (XFM)

Figure 1 | Optical microscopy images of anomalies in MS jugular vein tissues. Images a and b show suggested micro-calcifications (arrows). A scratch in the tissue is evident in a, while b, c and d show the singular appearance of same microvessel. The arrow in c indicates the presence of basophilic-calcified material inside a capillary. The same is revealed in panel d. All images are at 40 × magnification.

Rapid XRF imaging at XFM beamline (Australian Synchrotron)



Rapid XRF analyses on large tissue areas at 12.74 keV showed an increased Ca presence in the pathological samples, mainly localized in tunica adventitia microvessels.



Figure 2 | XRF elemental maps at 12.74 keV in MS2 tissue jugular sections. Three consecutive tissue sections of MS2 sample are used: two unstained for XRF analyses and one HH stained for tissue structure recognition. a) and b): light microscopy images, the boxes indicate the selected regions for XRF analyses in the unstained sections. The corresponding elemental maps of Ca, Fe and Zn of regions 1, 2 and 3 acquired at the XFM beamline at 12.74 keV with 2 μ m spatial resolution on the corresponding unstained tissue slices are shown in rows 1, 2 and 3 respectively. Red arrow in Ca map (1) indicates a calcification further analyzed at 4.12 keV. Arrows in Zn map indicate potential contaminants and tissue debris. The concentrations reported on the scale bars are in ppm. Region 1: 250 μ m × 170 μ m; Region 2: 600 μ m × 400 μ m; Region 3: 700 μ m × 600 μ m.

Investigations at lower energy demonstrated that the high Ca level corresponded to micro-calcifications, also containing P & Mg.



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Figure 3 | XRF analyses of microvessels in a MS2 jugular tissue section. Row 1: elemental maps of Ca, P, S and Fe acquired on regions 1 (100 μ m × 54.5 μ m) at 7.2 keV (ID21 beamline) with 0.5 μ m spatial resolution and 300 ms/pixel acquisition time; Row 2: elemental maps of Ca, P, S and Fe acquired on region 2 (100 μ m × 76 μ m) at 7.2 keV (ID21 beamline) with 0.5 μ m spatial resolution and 300 ms/pixel acquisition time; Row 3: Absorption (Abs) and phase contrast images (PhC) with the corresponding elemental maps of C, O, Mg and Na collected on region 3 (80 μ m × 80 μ m) at 1.5 keV (Twinmic beamline) with 0.5 μ m spatial resolution and 10 s/pixel acquisition time. The analysed regions are indicated in the corresponding visible light image (VL) of the MS2 tissue section. The red arrow indicates a region analysed at 4.12 keV too.





<u>In control tissues</u>, the major contributions in the XANES spectra seem to come from <u>organic Ca salts</u>.

On the contrary, in the <u>diseased subject</u> tissues the XANES results are in line with a substantial presence of <u>hydroxyapatite</u> (and other <u>inorganic calcium salts</u>) in clear connection with the vasa venorum.



Thank you!





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