



UNITED NATIONS EDUCATIONAL, SCIENTIFIC AND CULTURAL ORGANIZATION
INTERNATIONAL ATOMIC ENERGY AGENCY
INTERNATIONAL CENTRE FOR THEORETICAL PHYSICS
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H4.SMR/916 - 29

SEVENTH COLLEGE ON BIOPHYSICS:
*Structure and Function of Biopolymers: Experimental and Theoretical
Techniques.*
4 - 29 March 1996

Protein Folding - 1

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Towards a Realistic Folding Funnel for Proteins

UCSD { JNO
N. Socci
M. Betancourt
P. Leopold

⊕ UI { P. Wolynes
Z. Luthey-Schultze

Initial Funnel Idea
Numerical Simulation

Collaboration
Analytical ⊕ Numerical
Relation to Real
systems

SUPPORT:

NSF

Beckman Foundation

OUR APPROACH —

HOW CAN WE UNDERSTAND THE FOLDING MECHANISM

- What are the key features of proteins that allow them to perform that?
- How do the proteins select the appropriate structure (out of 10^{60} for 60 amino acids in a relatively short time (Levinthal Paradox)?

BIOLOGICAL VIEWPOINT

- Is it something specific (unique) to proteins?

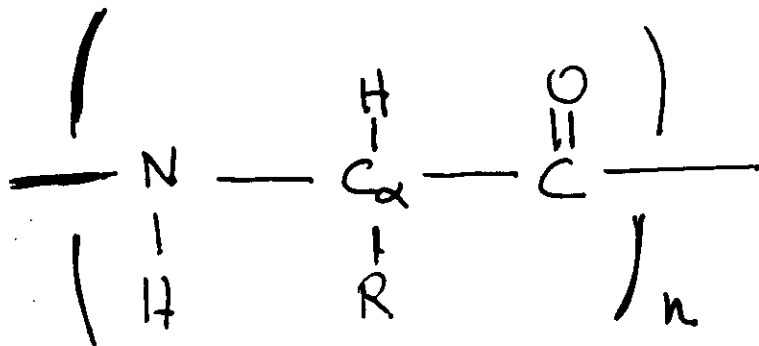
OR

PHYSICAL VIEWPOINT

- Are there generic, universal features shared by a class of systems which have these properties?

What are PROTEINS ?

Proteins are heteropolymers of Amino-Acids.



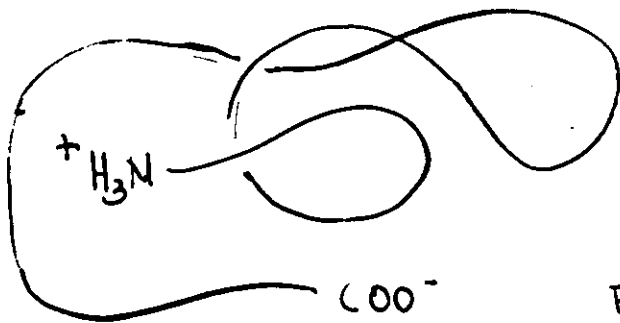
back bone

n - 40-400
small
globular
proteins



proper
condition

— physiological
conditions



UNIQUE
STATE

FOLDED
STATE

STABILITY
 $\sim 10 \frac{\text{kcal}}{\text{mol}}$ (0.4eV)

⊕ NOT ALL SEQUENCES FOLD!
BIOLOGICAL ONES TO FOLD!

How ??

FOLDING MECHANISM

Anfinsen '60s

Small globular proteins (ribonuclease) fold (refold) in the absence of any catalytic molecule.



THERMODYNAMIC HYPOTHESIS
(Global Stable Minimum)

Levinthal '68

Too many states to probe to find the right one !!! PARADOX

Example: | each A.A. 2 positions 2^N states $\begin{cases} N=100 \\ 10^{30} \end{cases}$

protein probes 10^{12} states / sec $\Rightarrow 10^{18}$ s

To save the T.H.

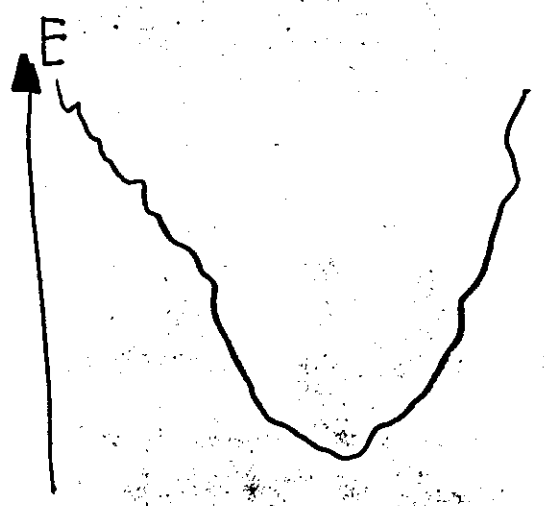
Baldwin 1983 \rightarrow Folding Pathway

REVISITING LEVINTHAL

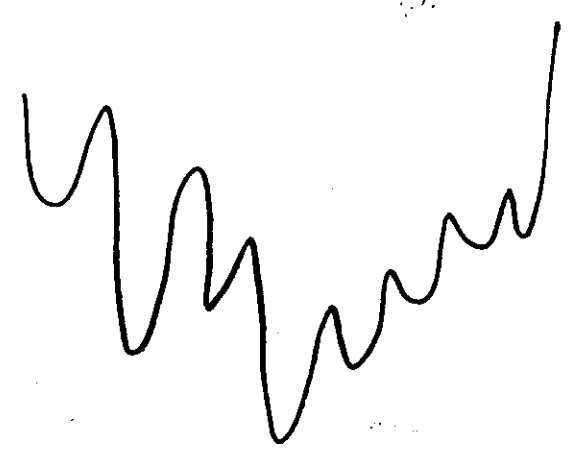
BUT CRYSTALIZATION WORKS !!!

HOW ABOUT PROTEIN FOLDING ???

SMOOTH LANDSCAPE



ROUGH LANDSCAPE



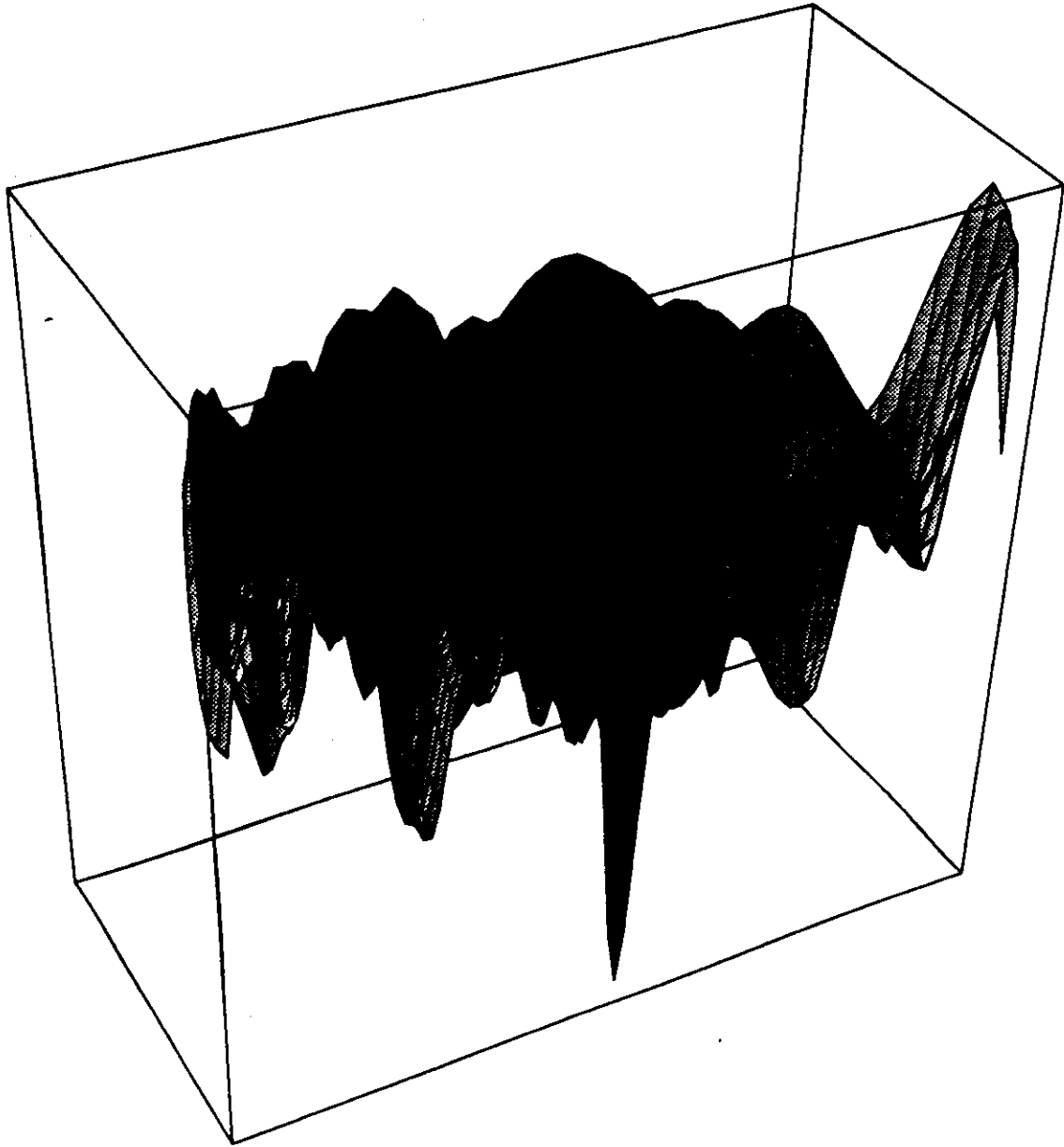
configurational
 entropy loss
 is balanced
 energy gain from
 the slope

BUT REAL
 SYSTEMS
 HAVE SOME
 ROUGHNESS
SOME FRUSTRATION

PRINCIPLE OF MINIMAL
 FRUSTRATION

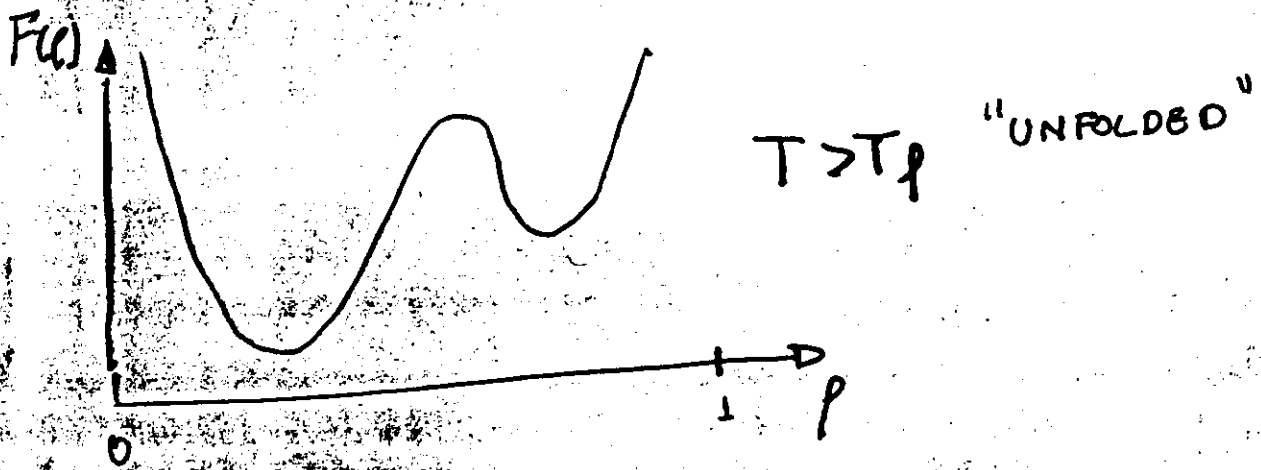
Folding time considerable small ⁶⁵ wolyws
 compared to Levinthal

⇒ TO ADDRESS THIS QUESTION → SIMPLE MODEL

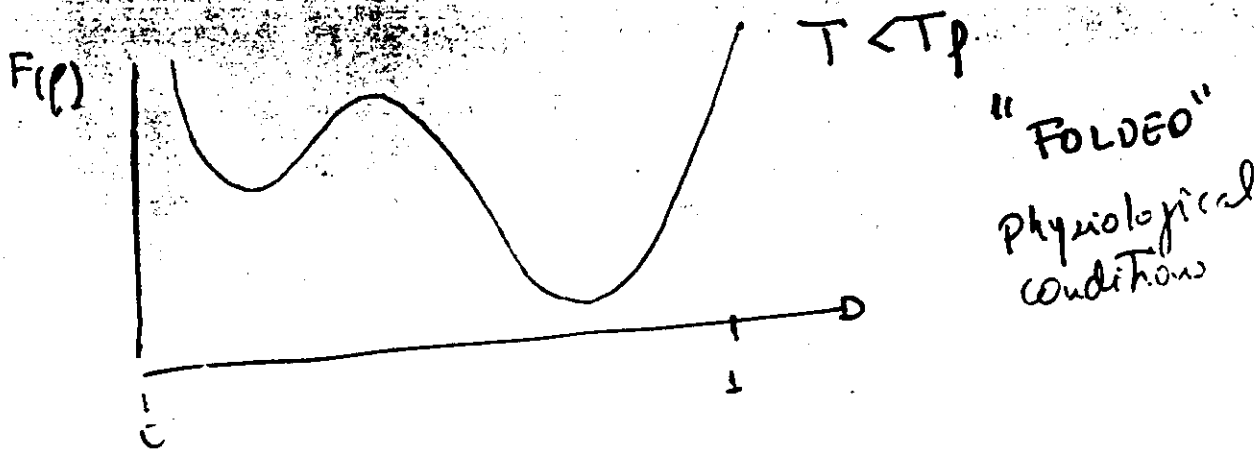


FOLDING THERMODYNAMICS

FOLDING IS AN "ALL OR NONE" TRANSITION
(FIRST ORDER LIKE)



$p = \% \text{ of folding}$



Double well Free Energy Surface.

SPIN GLASS IDEAS - Simple view

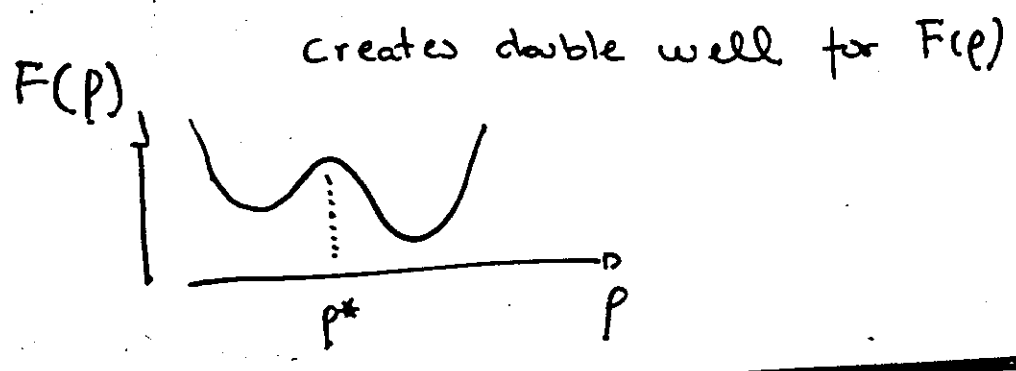
Bryngelson and Wolynes

Thirumalai
Wolynes
Bryngelson
Shakhnovich
Dill
Orland
○
○
○

ORDER PARAMETER $\xrightarrow{\text{CALL}}$ $P \equiv \% \text{ of folding}$

(A)

P increases $\rightarrow S(P)$ gets smaller
 $\rightarrow \bar{E}$ gets smaller



(B) Include roughness in the "potential" energy landscape
The "non-folded" part of the "protein" is treated as a Random Energy Model (REM)

* for every P there is a glass transition temperature $T_g(P)$

Bryngelson/Wolynes Model

one order parameter P !
REM ⊕

ENTROPY CRISIS TRANSITION

(Good folding sequences $\rightarrow T_f > T_g(P^*)$)

| Best Folding occurs $T_f > T > T_g$ |

Random Energy Model

17

Spin Glass

$$H_p(\text{tot}) = - \sum_{(i_1, i_2, \dots, i_p)} A_{i_1 \dots i_p} \sigma_{i_1} \dots \sigma_{i_p}$$

As p increases Energy configurations become random variables

$$\{ \sigma_{i_1}^{(1)} \} \quad \{ \sigma_{i_2}^{(2)} \}$$

$$P(E_1, E_2) = P(E_1) P(E_2)$$

therefore

2^N energy levels

NJ^2 - variance of energy levels

$$P(E) = \frac{1}{\sqrt{N+J^2}} e^{-E^2/NJ^2}$$

if $n(E) \gg 1$

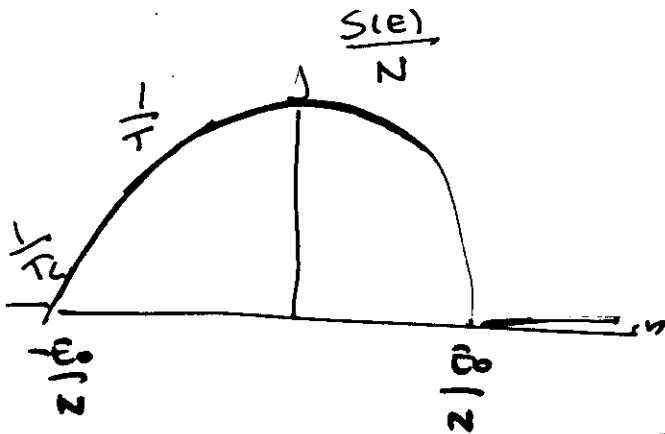
$$\langle n(E) \rangle = 2^N P(E) \sim \exp \left\{ N \left[\ln 2 - \left(\frac{E}{NJ^2} \right)^2 \right] \right\}$$

$$S(E) \sim \ln n(E)$$

$$= N \left[\ln 2 - \left(\frac{E}{NJ} \right)^2 \right]$$

$$\text{if } S(E) > 0 \quad \Rightarrow \quad |E| < E_0$$

$$\frac{E_0}{N} = J \sqrt{\ln 2}$$



$$\frac{dS}{dE} = \frac{1}{T}$$

$$T_c = \frac{J}{2\sqrt{\ln 2}}$$

$$S=0 \Rightarrow \Omega = 2^N = e^{N S_c^* / k_B}$$

$$k_B \ln \Omega = N S_c^* = \frac{N J^2}{(k_B T)^2}$$

S_c^*
configurational
entropy

$$\frac{E}{N} = -T \ln 2 - \frac{J^2}{4T} \quad T > T_c$$

$$\frac{E}{N} = -\frac{E_0}{N} = -J \sqrt{\ln 2} \quad T < T_c$$

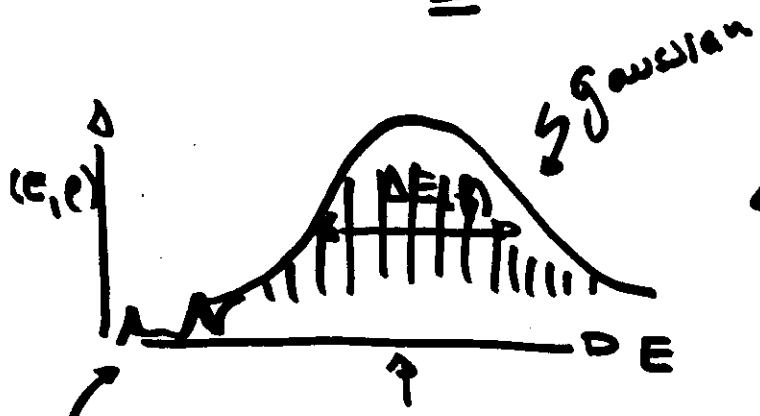
S.O.P.T

In Principle

For every T
there is a ρ
 $T_g(\rho) = T$

A view on $T_g(\rho)$

For every ρ



Total $\Omega(\rho)$ states

deep states depend on sequence

mean depends on similarity to native state

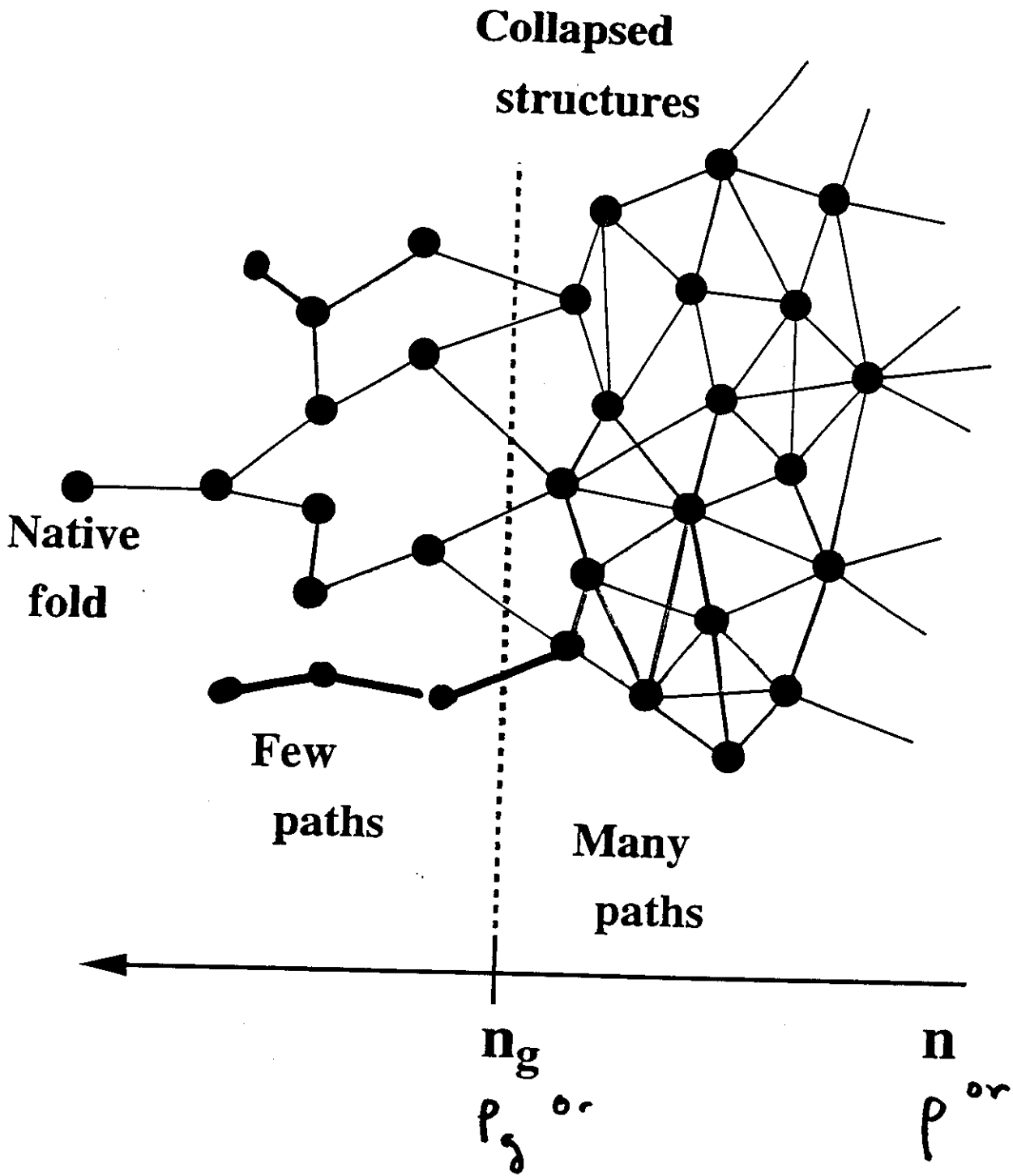
$$-E_{\text{not}} \rho$$

$$\langle E \rangle = -E_{\text{not}} \rho - \frac{\Delta E^2(\rho)}{2T}$$

$$S = k_B \ln \Omega(\rho) - \frac{\langle E \rangle^2}{2\Delta E^2} \sim 0$$

glass transition

$$T_g(\rho) \sim \frac{\Delta E}{\sqrt{2 \ln \Omega(\rho)}}$$



Simple Models ⊕ Analytical Theories
is a powerful tool for studying
and understanding the
folding mechanism



LAW OF corresponding States

Several other groups
working on simple models:

- Chan and Dill
- Thirumalai and Comacho
- Shakhnovich
- Orland
- Bryngelson
- Watshell and Levitt

⋮

"More Realistic
Models"

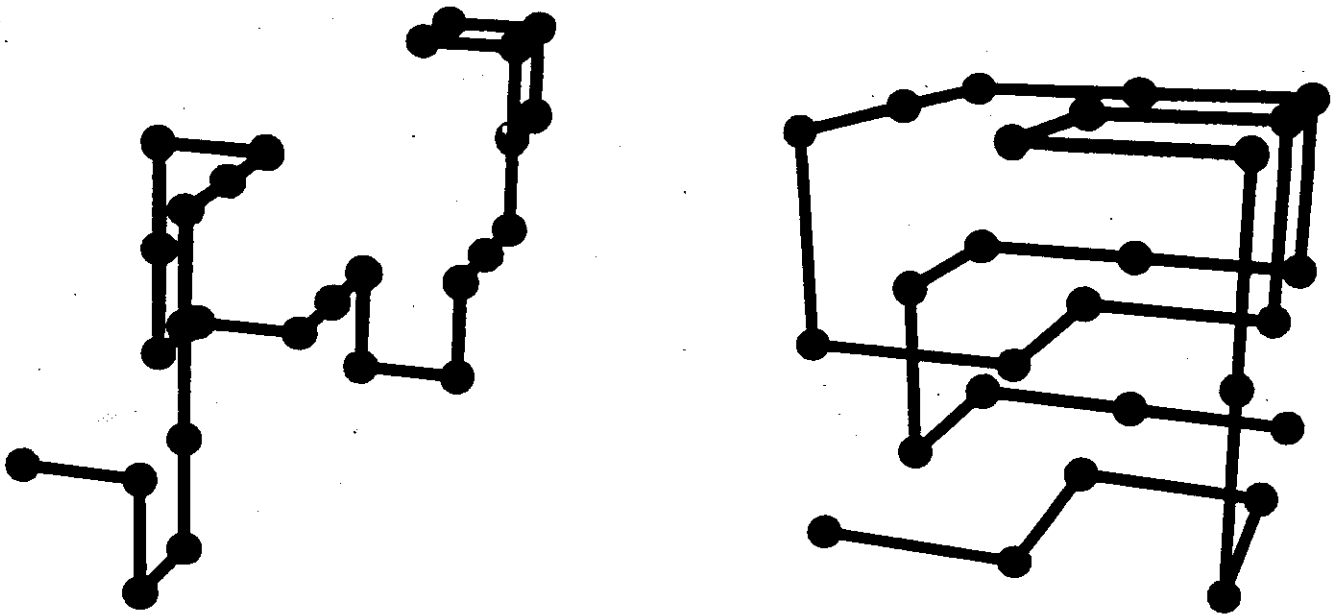
Jeff Skolnick

Charlie Brooks

ATO
MIT
C

- Levitt and
Daggett
- Scheraga
- Honig and
Friesner

Model: 3D Simple Cubic Lattice Copolymer



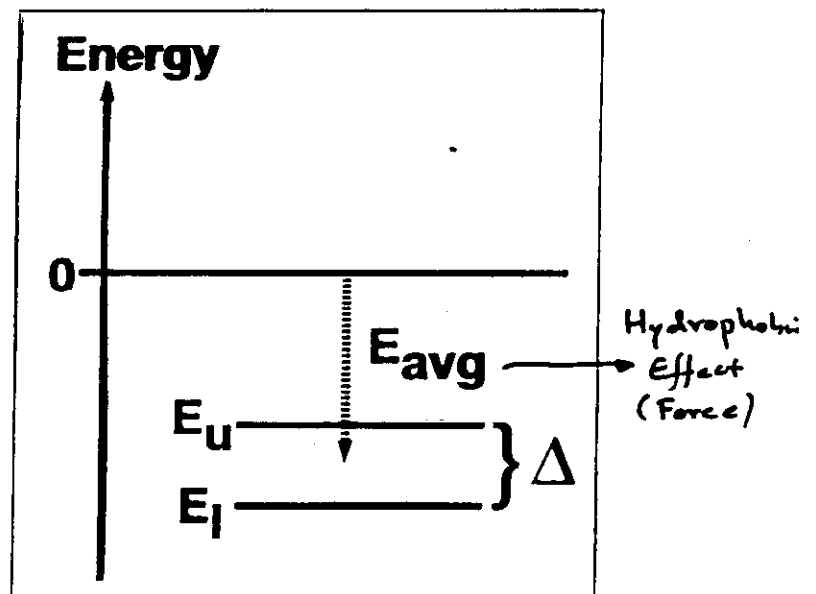
- 27 monomers long. (Can not enumerate all conformations, but can enumerate cubes.) $N_c = \text{MAX} \# \text{ CONTACTS } 28$
- Potential: $Q = \text{correct contacts}$

$$E = N_l E_l + N_u E_u$$

$$E_l < E_u < 0$$

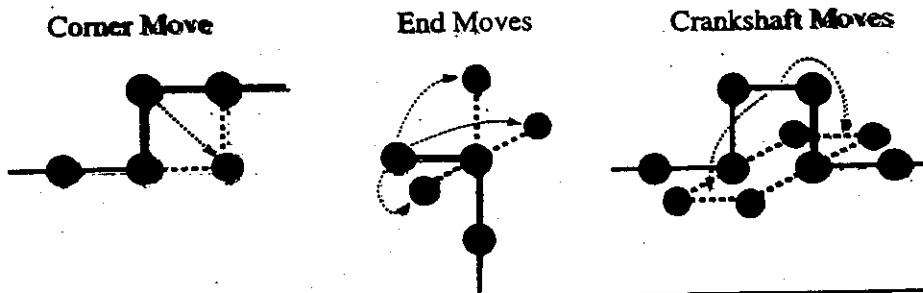
Initially

$$E_l = -3, E_u = -1$$



These values insure rapid collapse and that the ground state (native conformation) will be a cube, which we can find by enumeration

Dynamics—Local move Monte Carlo.^a



Simulations

1. Kinetic

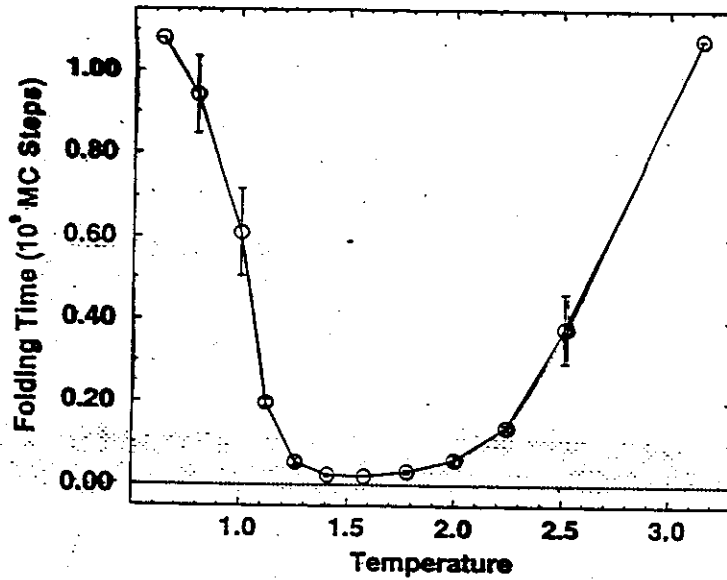
- Starting from unfolded initial condition, run many folding simulations at various temperatures.
- Maximum simulation time: $T_{max} = 1.08 \times 10^9$
- Calculate mean first passage time (MFPT) for collapse and folding.
- Define kinetic glass transition temperature: T_g

2. Thermodynamic—MC Histogram Method

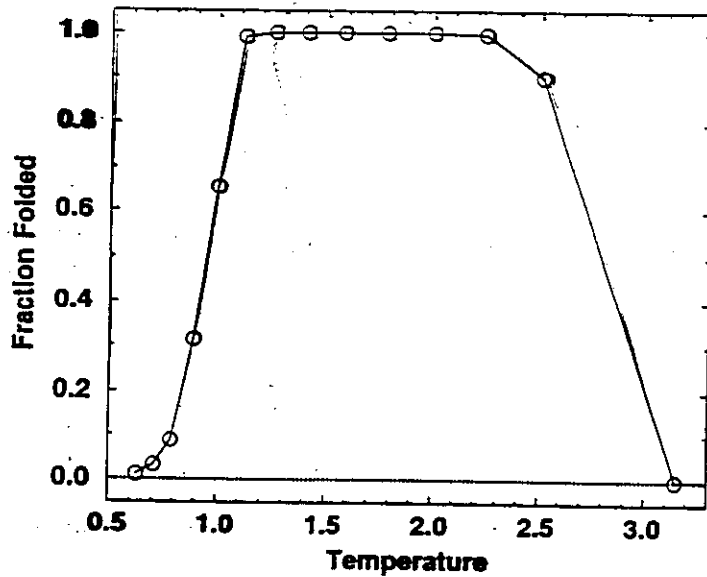
- Determine folding temperature, T_f . Compare with T_g for several sequences.
Protein-like: $T_f > T_g$

^aSOCCI & ONUCHIC, *J. Chem. Phys.*, 101 1519 (1994)
J. Chem. Phys., in press

Kinetic Glass Transition



* Also in
 2D-Chen, Dill, Halle
 Adu-Savari, Wang
 Wolynes



Temperature at which the system slows down (also loses self-averaging property). Folding time (τ_f) diverges and few runs find native state.

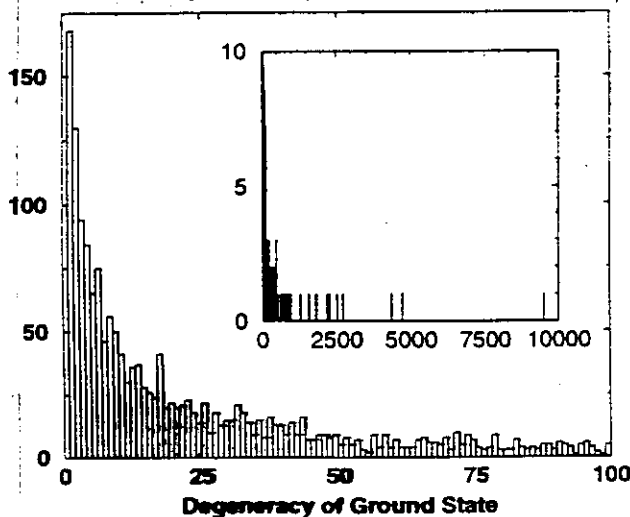
$$\tau_f(T_g) = \frac{1}{2}(\tau_{max} - \tau_{min}) \approx \frac{1}{2}\tau_{max}$$

$$T_g \approx 1$$

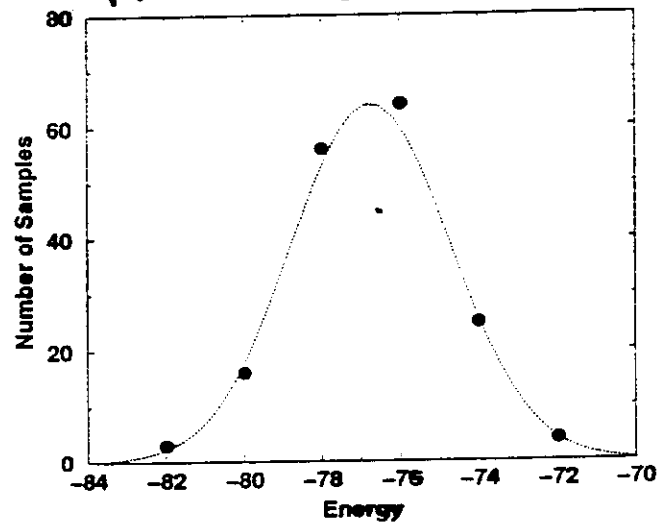
Run	Sequence	E_{\min}
002	ABABBBBABBABABAAABBAAAAAB	-84 ^a
004	AABAABAABBABAAAABBABABABBB	-84
005	AABAABAABBABBAABABBABABABBB	-82
006	AABABBABAABBABAAAABABAABBBB	-80
007	ABBABBABABABAABABABABBBEABAA	-80
013	ABBBABBABAABBBAAABBABAABABA	-76

- All composed of 2 monomer types: same A:B ratio (14:13).
- All have unique ground states.
- Varying values for E_{\min} . Different amounts of "frustration," in ground state.
- 2 (-84) were designed, 4 chosen at random. (In 2000 samples did not get a -84.)

Ground State Degeneracy
19,500-2000 Samples

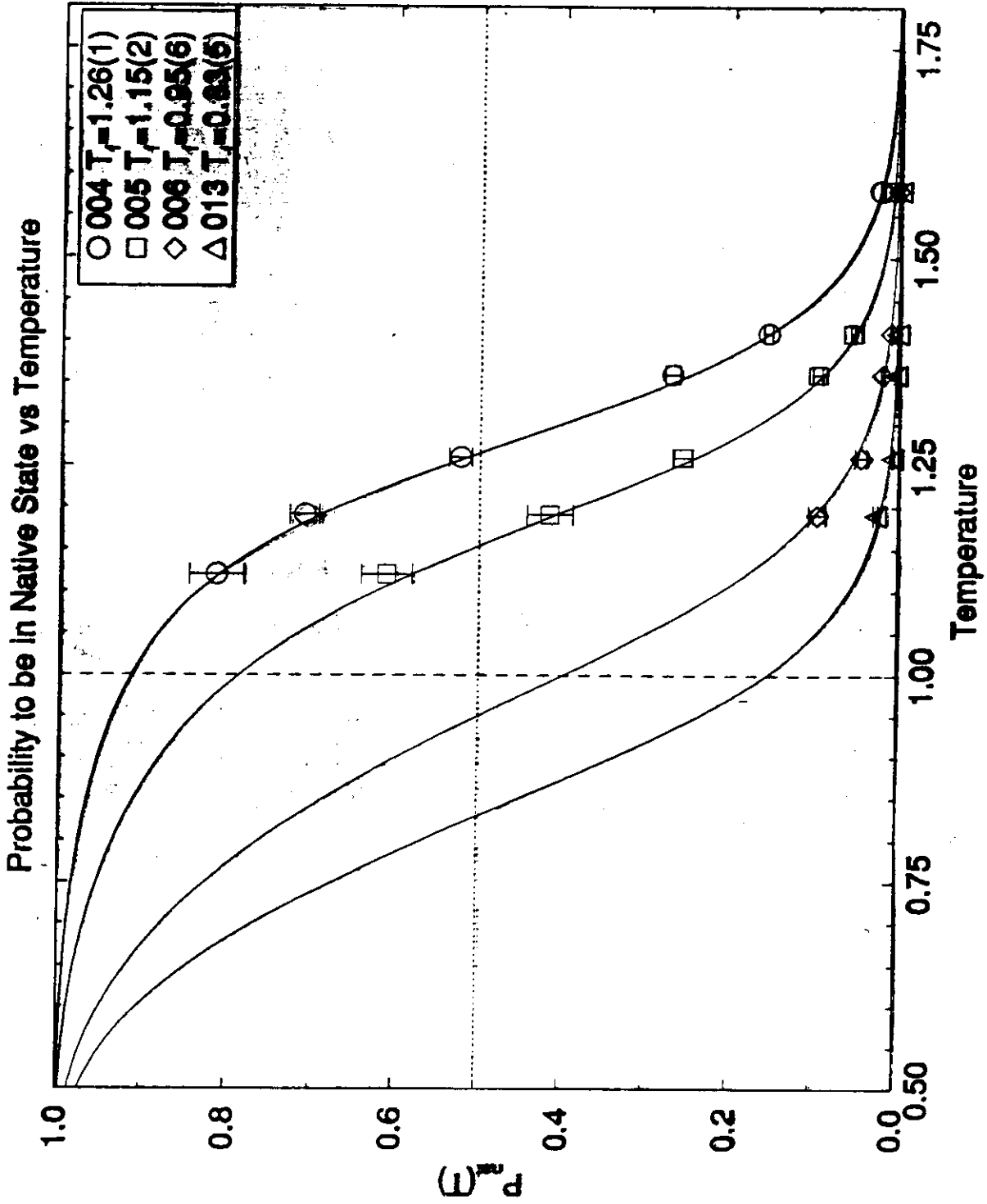


Distribution of Ground State Energies
1462-100 Non-degenerate Cubes



^a Shakhnovich, PNAS, 90 7195 (1993)

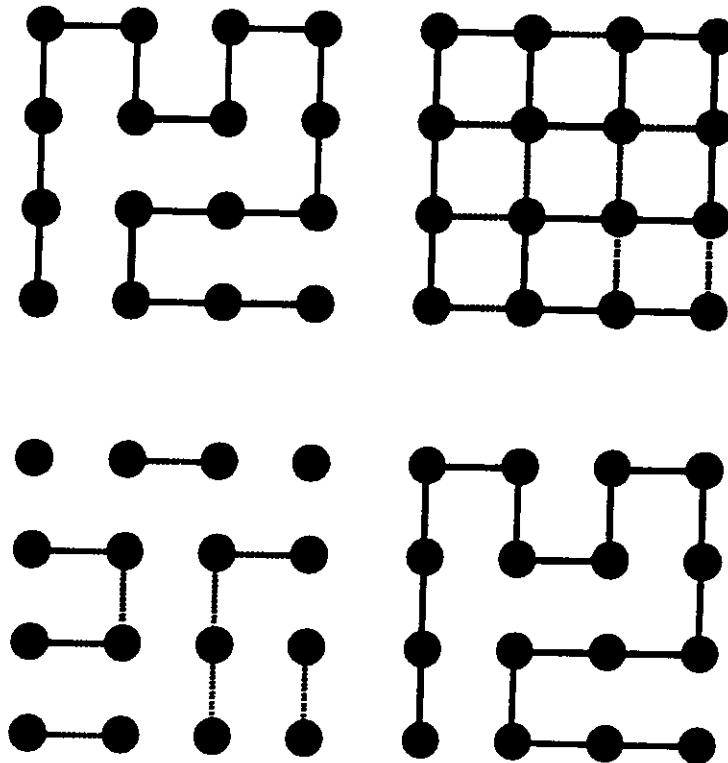
Folding Temperature



- 2 monomer type sequences ($2 LC$) are qualitatively protein-like ($T_f > T_g$), but not too good quantitatively.

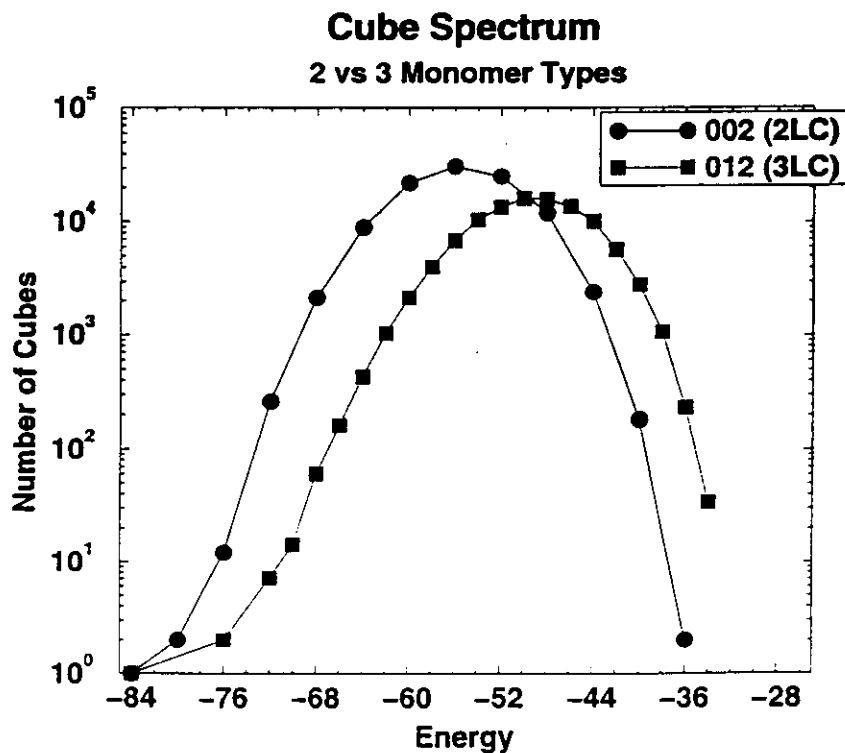
$T_f/T_g \approx 1.3$ is a little too small.

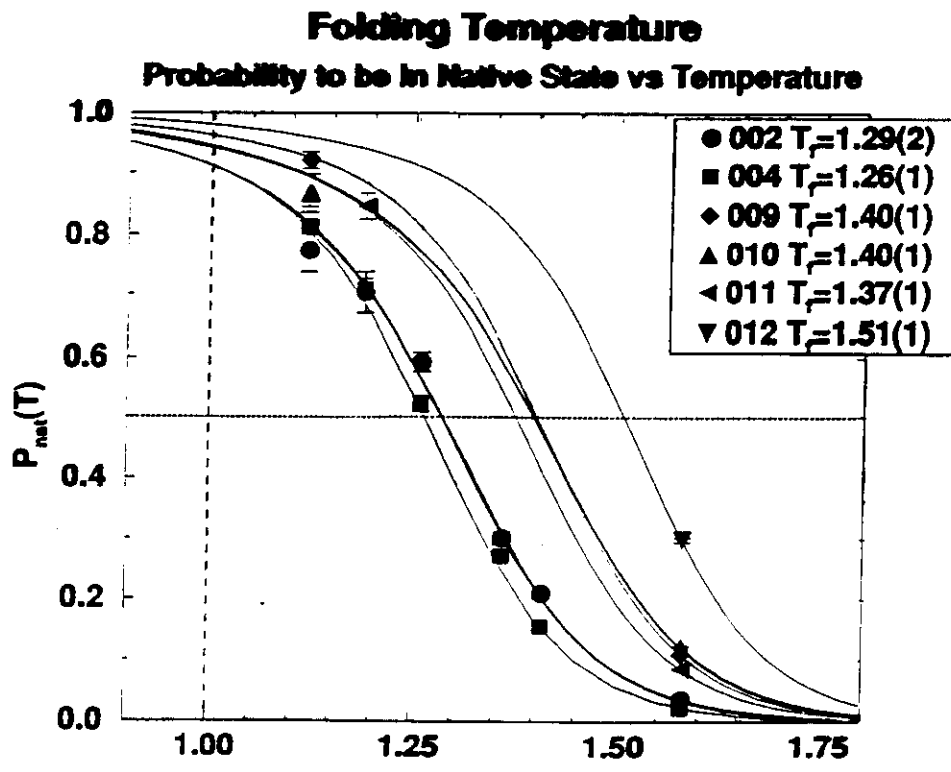
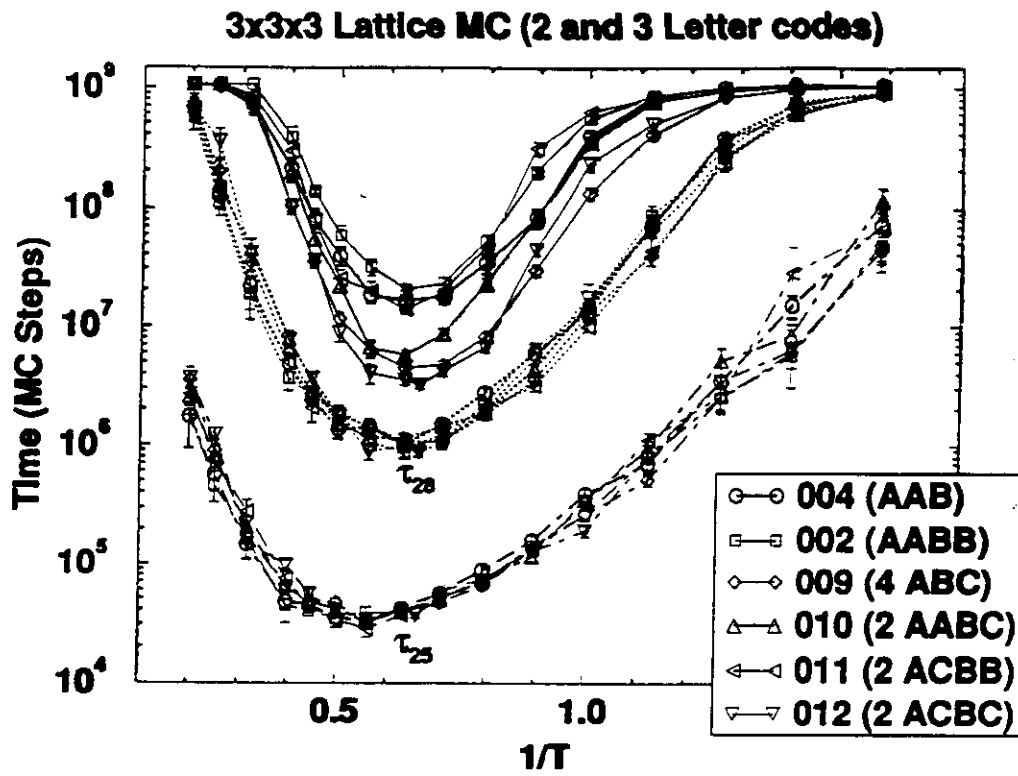
- Can we tune the model to give a higher T_f/T_g ratio? Ie, can we make it more protein like?
- Change number of monomer types ($2 \rightarrow 3 LC$).
Idea: reduce “energetic frustration” in the collapsed states by reducing the number of strong by incorrect (non-native contacts).
- Design sequences by domain decomposition.



Run	Sequence	τ_{min}	T_g	T_f
002	ABABBBBBABBABABAAAABBAAAAAAB	2.0×10^7	1.02	1.285(15)
010	ababbbCbabababaaaCbaaaaaab	6.2×10^6	0.94	1.40(1)
011	ababbbababababCaabaCaaCab	1.4×10^7	1.03	1.37(1)
012	ababbbCbabababCaCbCaaCab	2.9×10^6	0.91	1.509(1)
004	AABAABAABBABAAAABBBABABBBB	1.6×10^7	0.99	1.26(1)
009	aaCaabaabbabaaaCabCabababbb	4.3×10^6	0.86	1.40(1)

- Energy of native (ground state) remains unchanged.
- Energies of many compact non-native states are increased.
- Smooths out the energy landscape, removing traps.





- Adding a third amino acid type, can decrease folding time by almost a factor of 5 (collapse and folding converge).
- Can lower T_g slightly and increase T_f dramatically.

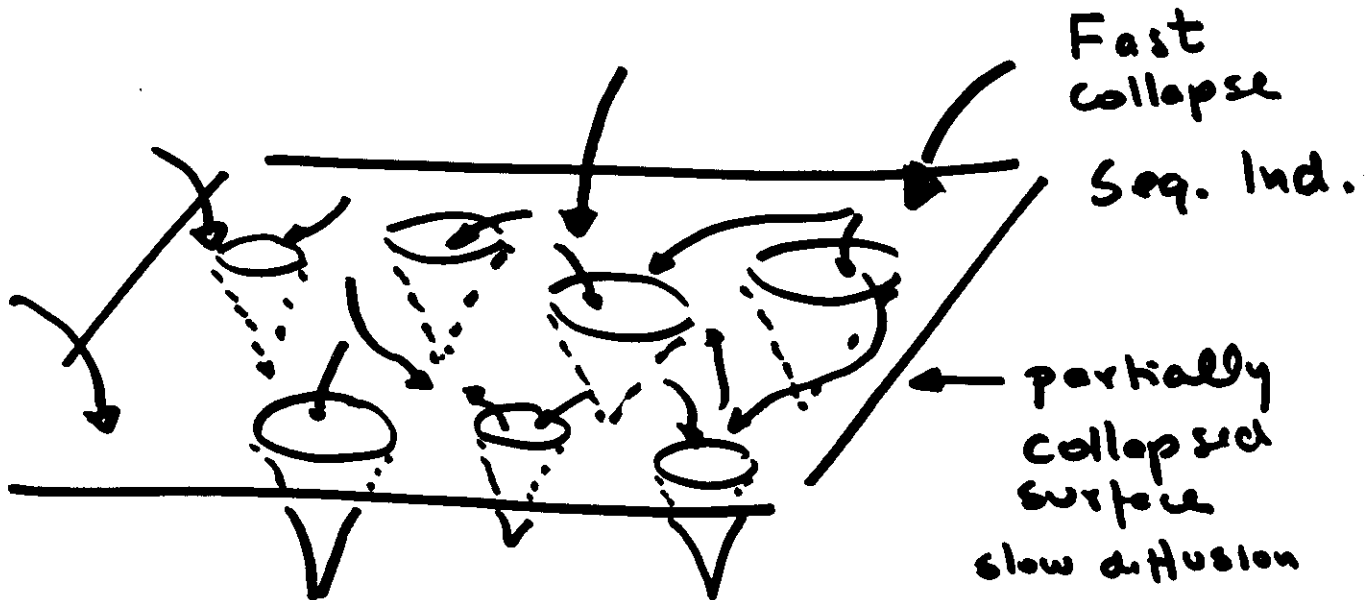
$$T_f/T_g \rightarrow 1.6 \quad (\approx 20\%)$$

- Concept of *Minimal Frustration*. As energy decreases conformations how greater overlap with the native state. Truer for the 3LC sequences than the 2LC.
- Using a 3LC sequences combined with analytical models for chain collapse, we are beginning to make connections with experimental data for real proteins.^a

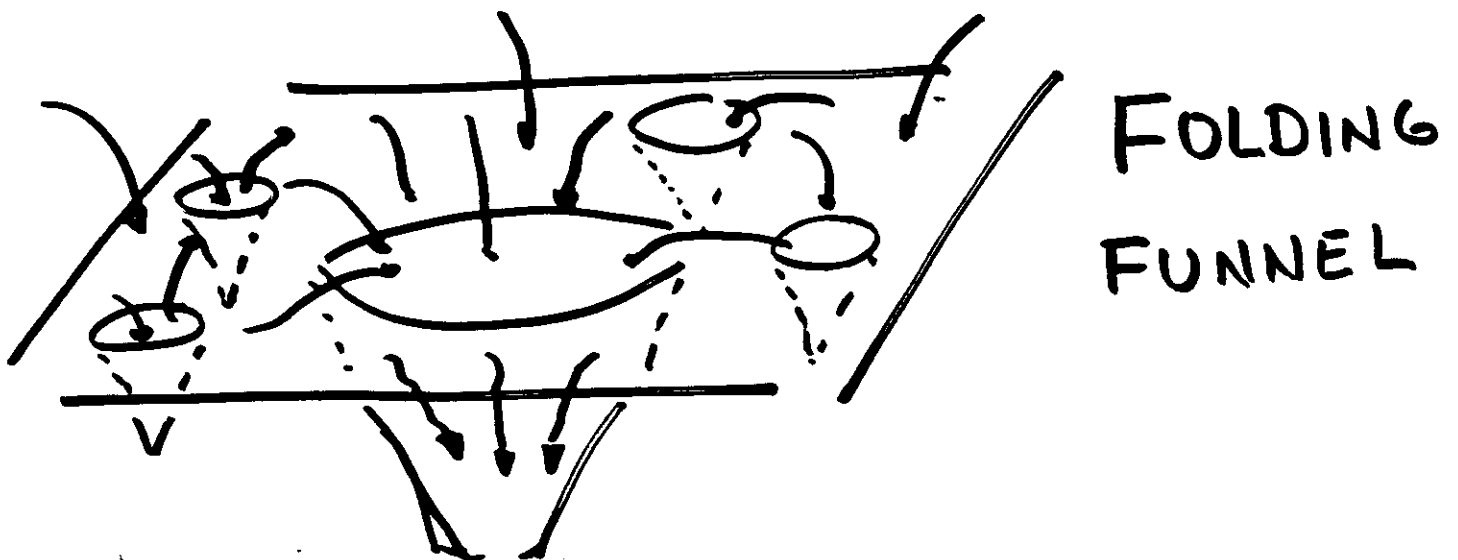
^aONUICHIC, WOLYNES, LUTHEY-SCHULTEN & SOCCI, *PNAS*, in press.

What do we learn from simple models

- Ruggedness large compared to stability
Random Heteropolymer



- Ruggedness small compared to stability
Protein Sequences



NEW FOLDING PICTURE

Onuchic
Wolynes
Luthey-Schw
Socci

● Proteins Have "2" Energy Scales

- H-Bonds
- Interactions Non-Local Sequence !!
but Local Space

LAW OF CORRESPONDING STATES

⑩ EXAMPLE - Small α -Helical Protein

60 AA protein

27-mer lattice model

Collapsed
Molten Globule
State

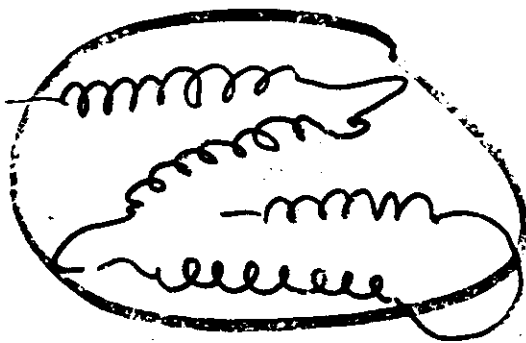


Similar
Levinthal
entropy

(High Degree of Helicity ← Not necessarily native helices)

"Lattice Model"

Ramirez
Schulten
Wolynes



Exp. Support
Callender + Los Alamos
Kallenbach - Dyer et al
Apomyoglobin
Engelman -
Staphylococcal
nuclease

Kinetic View

Reconfiguring among these states

3 IMPORTANT PARAMETERS

1. Size of Landscape \equiv Levinthal Entropy
 S_L
2. Ruggedness of Landscape \cdot $\text{MAX } \frac{T_f}{T_g}$
 ΔE^2
3. Slope of Funnel \rightarrow stability gap \cdot T_g
 δE_s



⊙ Models with Similar Parameters
 \rightarrow Similar Mechanism

LAW OF CORRESPONDING STATES:

relating real proteins with simple model

⊙ Need of experiments to study "early folding" events \rightarrow Dill Eaton !!! 000

$\frac{T_p}{T_g}$ Real Proteins

Comparison of motions for free chains and collapsed chains

$$\tau_{\text{reconf}} = \tau_{\text{free}} e^{\frac{\Delta E^2}{2T^2}}$$

$\tau_{\text{free}} \sim \underline{\underline{ns}}$ { simulations
Rouse-Zimm theory

$\tau_{\text{reconf}} \sim \underline{\underline{ms-s}}$ { NMR e.g. lactalbumin Pamm
apo-glycogene Wanch
b562

$$\frac{\Delta E^2}{2T_p^2} = 11-18$$

Folding Transition

$$\frac{\Delta E_s}{T_p} = S_L + \frac{\Delta E^2}{2T_p^2}$$

energy loss down the funnel compensates entropy loss

$$T_g^2 \sim \frac{\Delta E}{\sqrt{2S_L}}$$

$S_L \sim 30-40$

$\Rightarrow \Delta$

$$\frac{T_p}{T_g} \sim \underline{\underline{1.6}}$$

Order Parameters

$$\underline{N_c} = \# \text{ contacts} \quad 0-28 \quad \text{collapse}$$

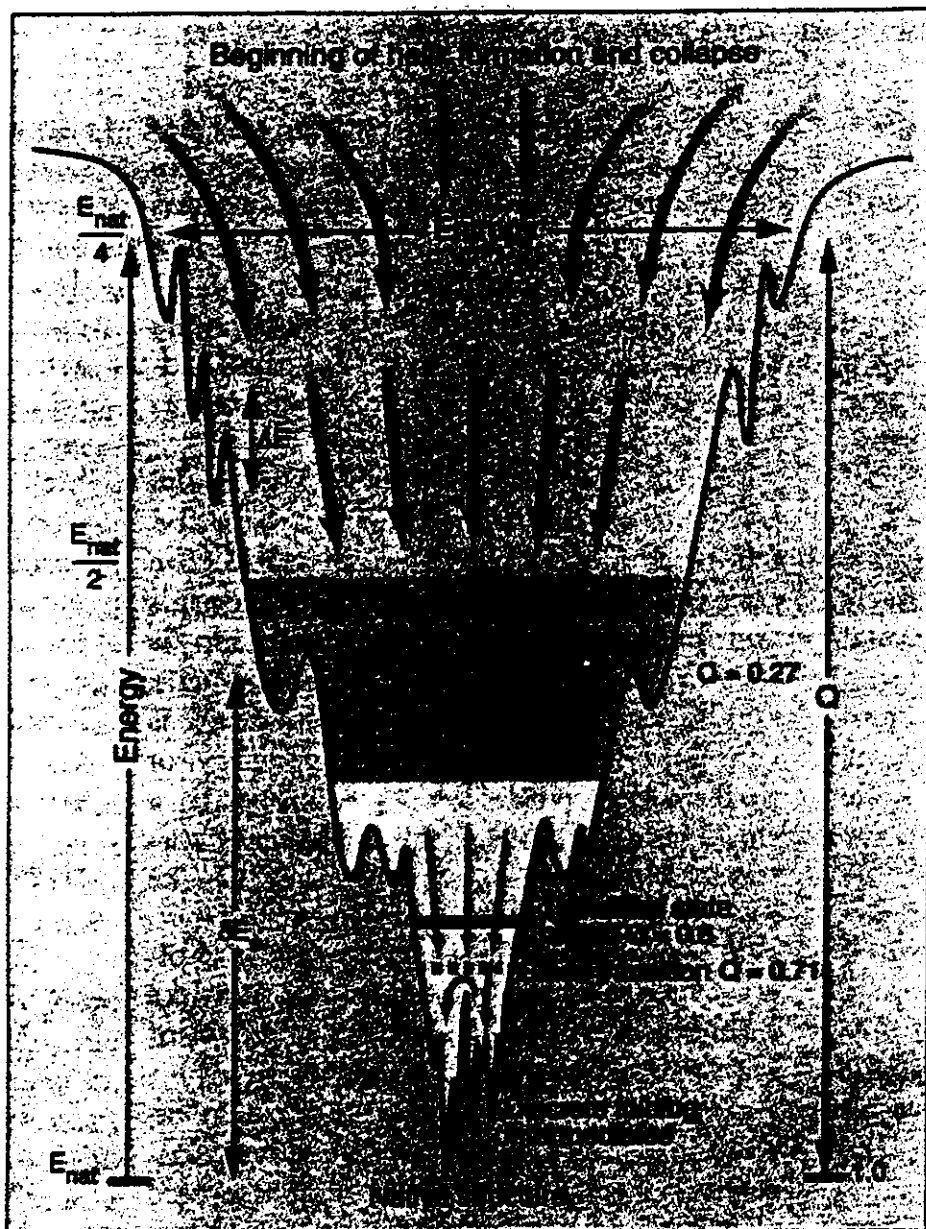
$$\underline{Q} = \# \text{ native contacts} \quad 0-28$$

$$\underline{A} = \# \text{ native angles} \quad 0-25$$

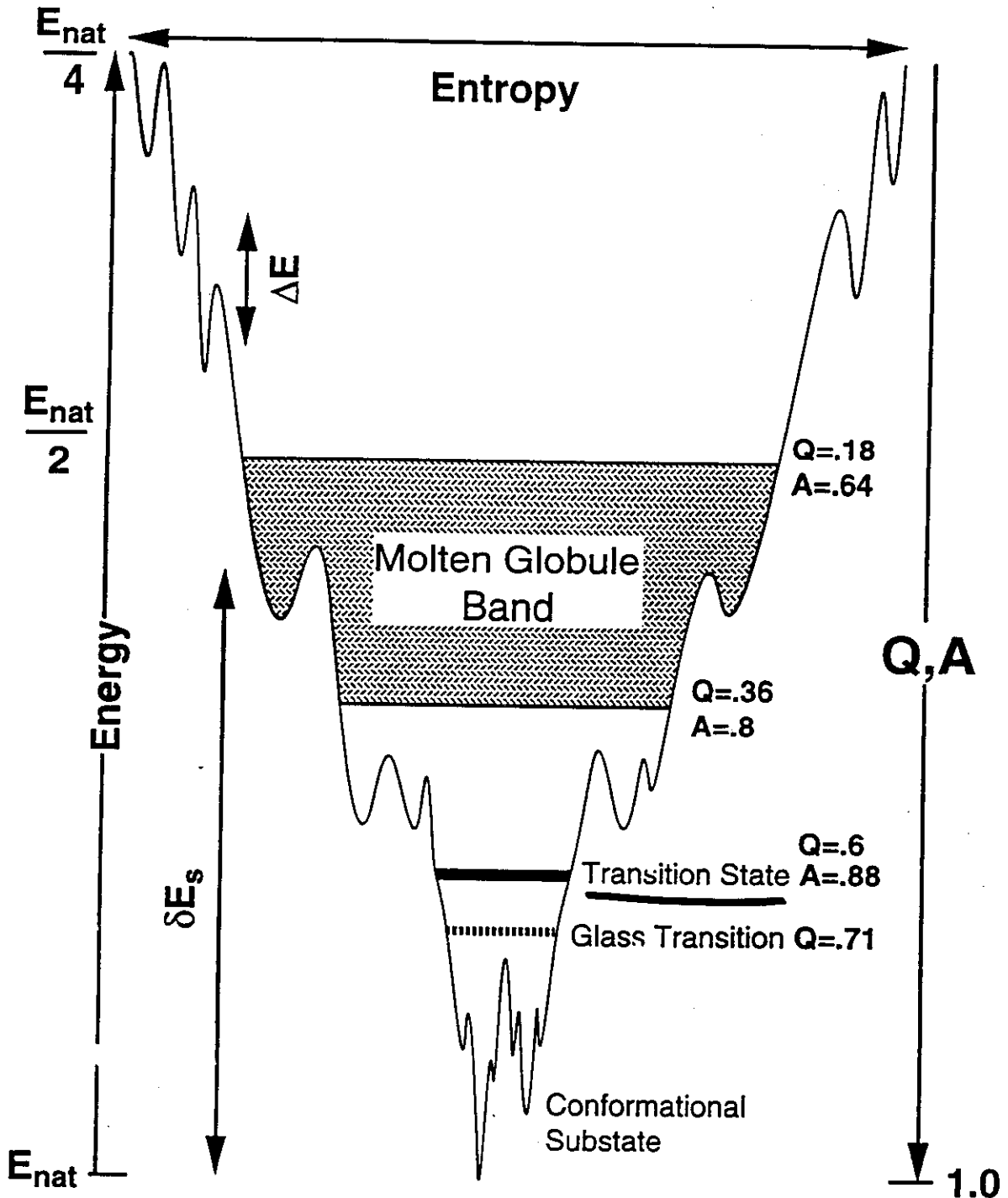
Bryngelson-Wolynes

$$\bar{E}_{\text{NATIVE}} = +28 \times (-3) = -84$$

Beginning of new formation and collapse



$Q = 0.6$ in TS ohay Fusch (CI2)



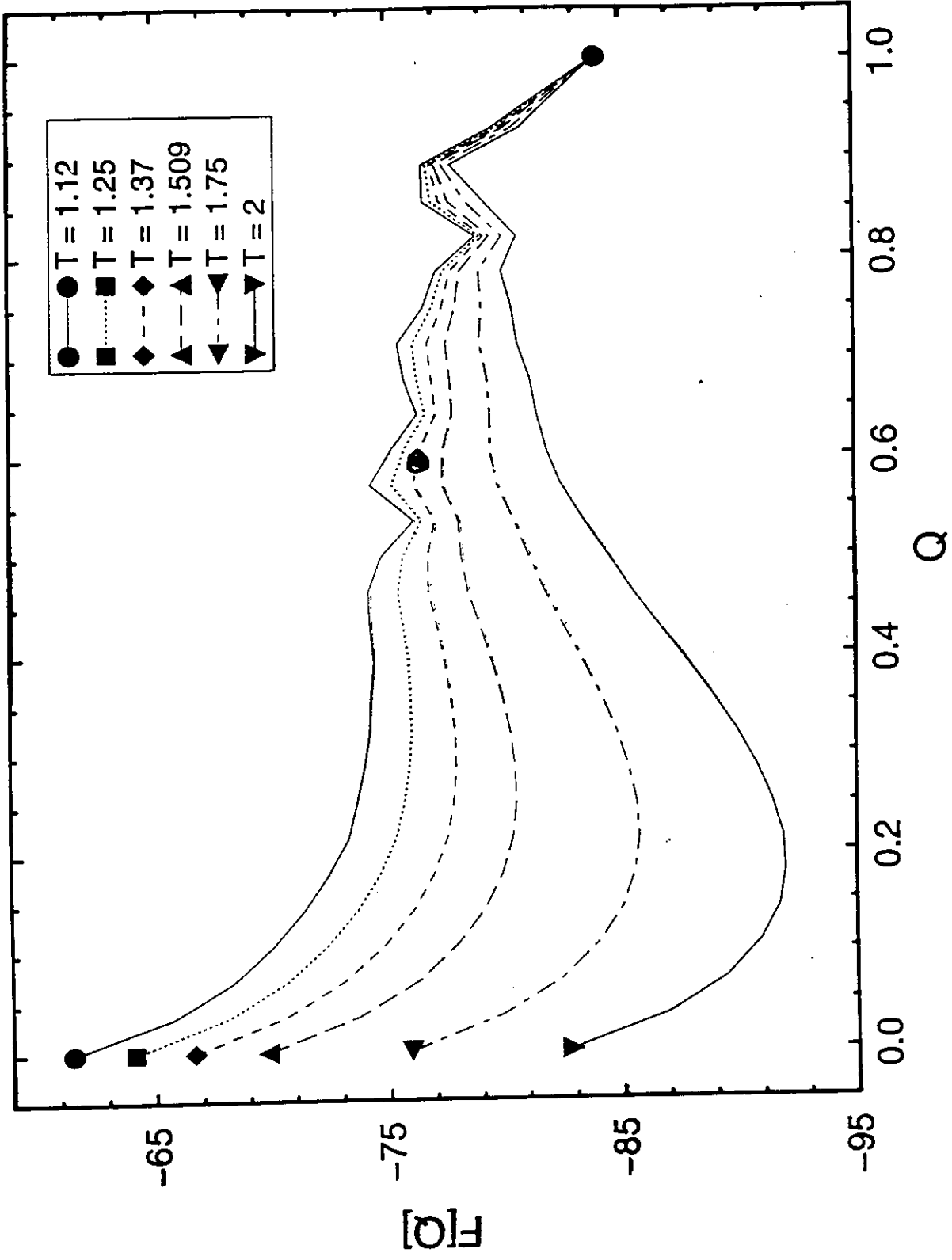
$\frac{\delta E_s}{\Delta E} \sim 14$

$\frac{T_f}{T_g} \sim 1.6_1$

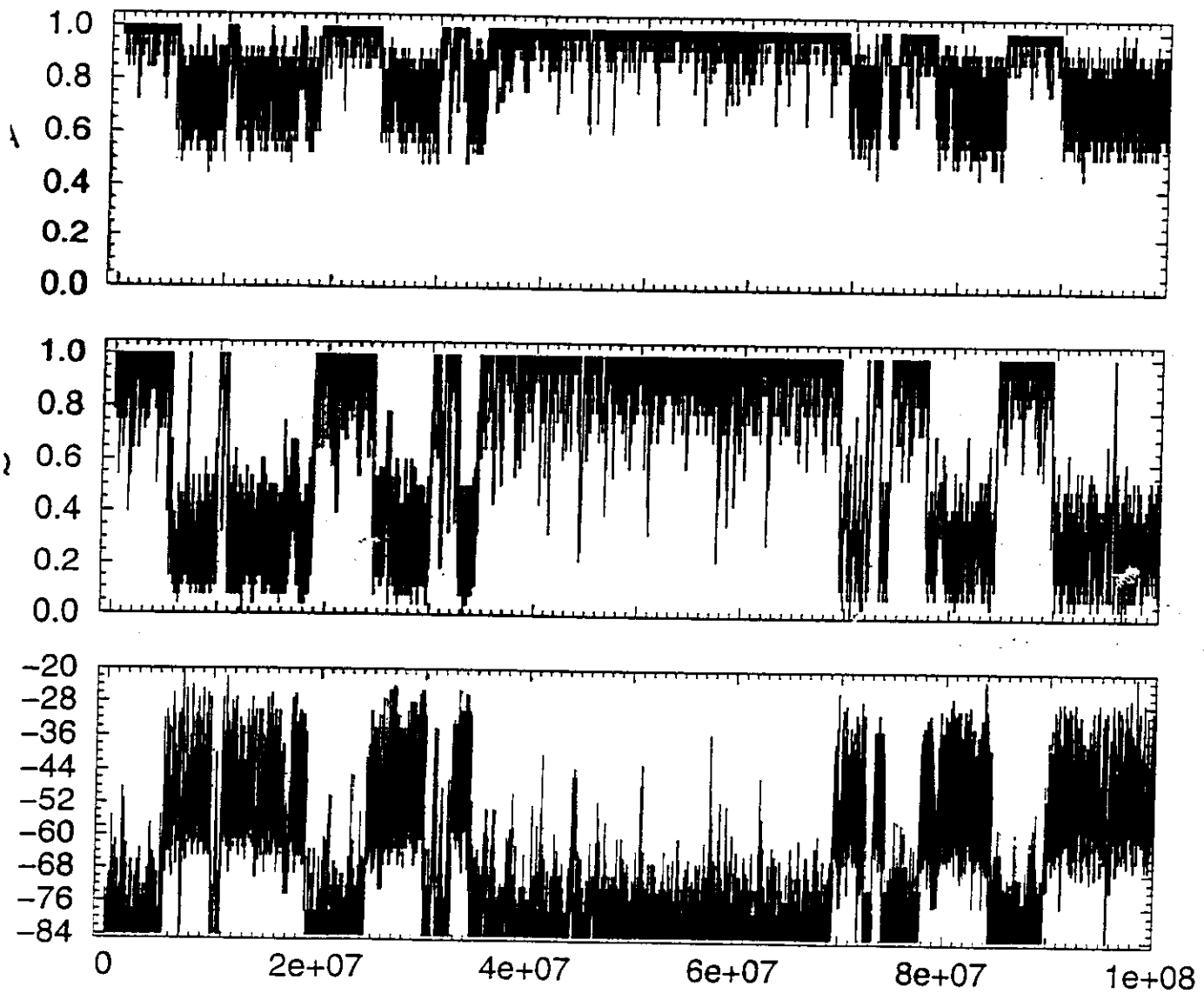
Based on correspondence
states to 3-letter
code

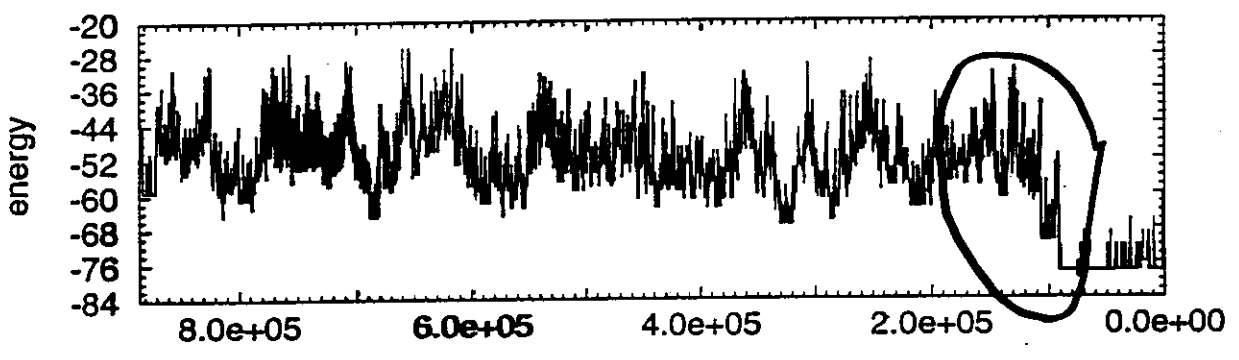
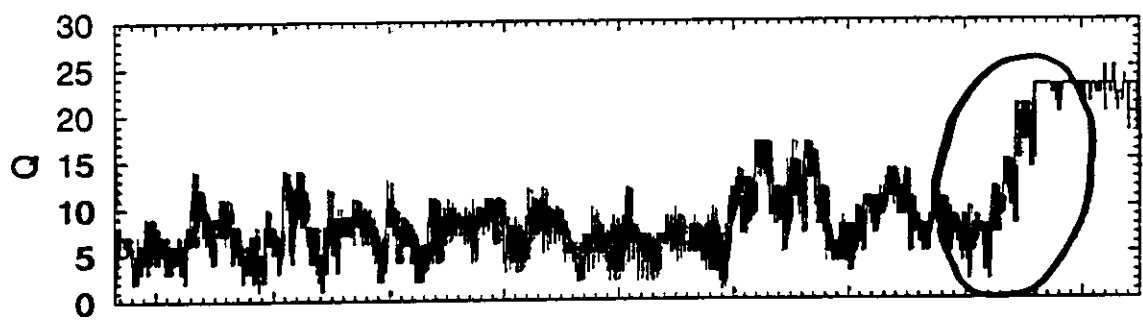
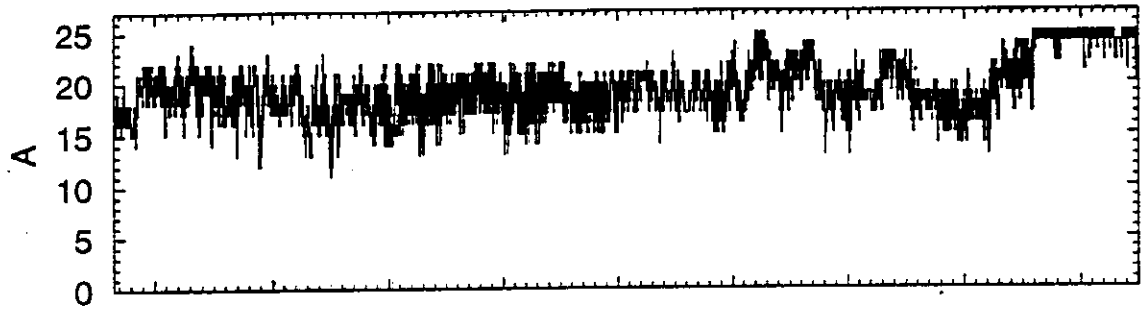
Free Energy vs Q

Seq012 (3LC) $T_s = 1.58$



$$\tau_f \sim 3 \times 10^6$$

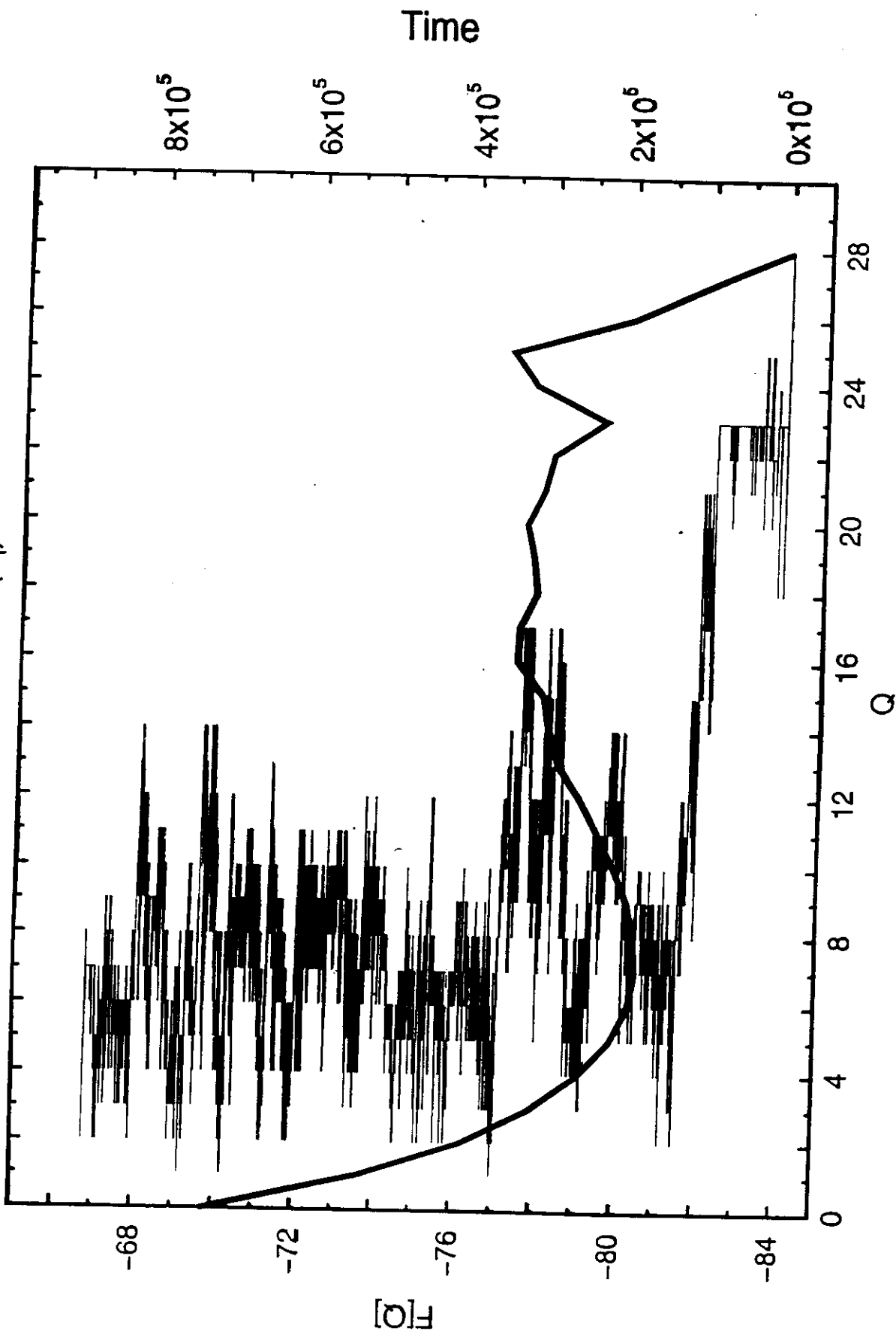


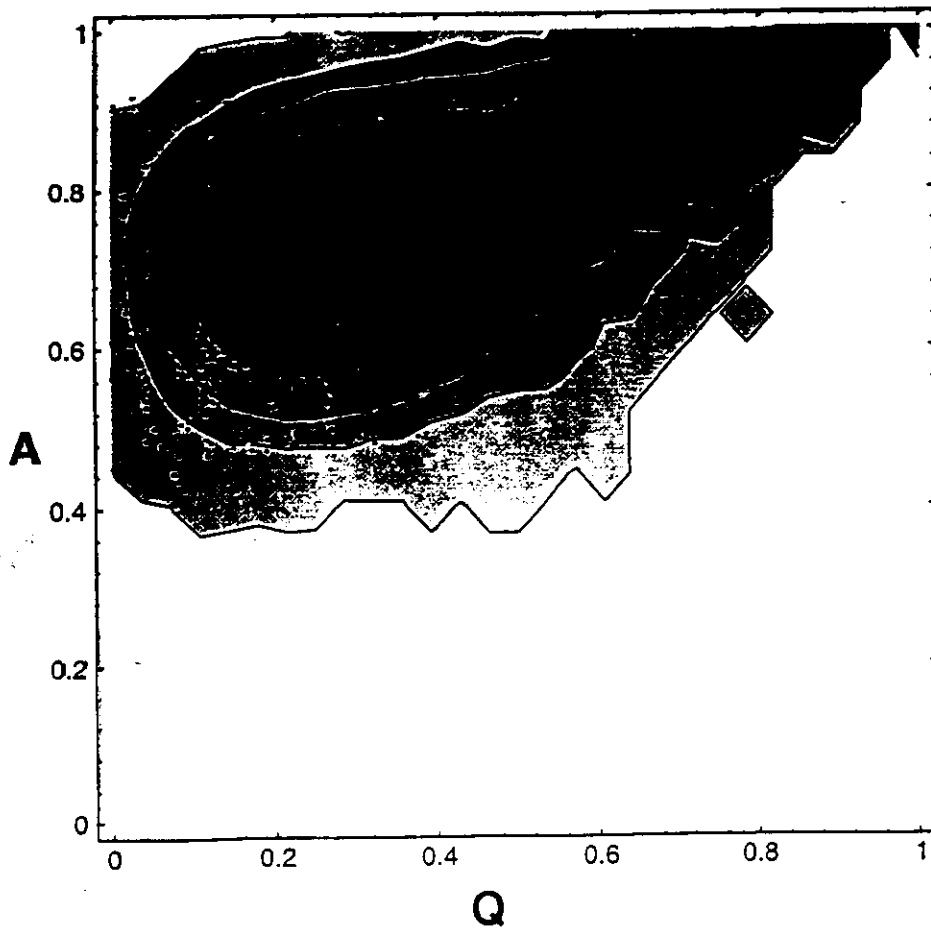
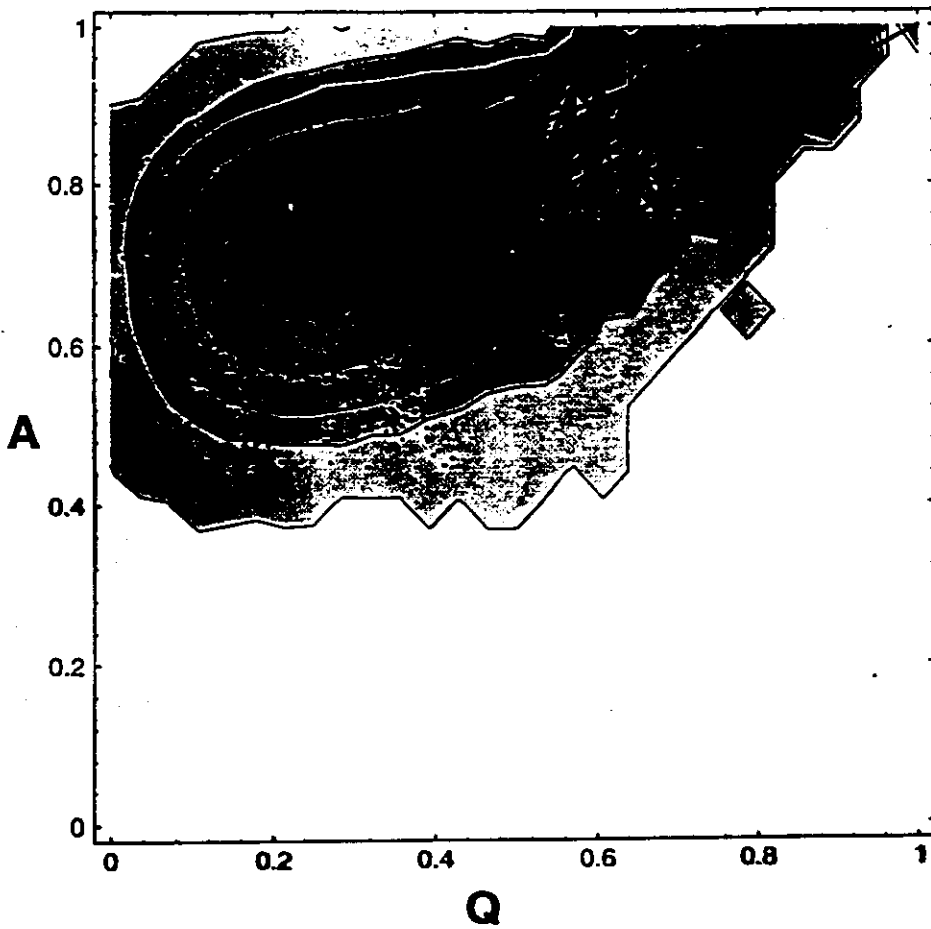


~ 100,000 steps

Free Energy and Folding Traj vs Q

Seq012, T=1.51 (T_f)

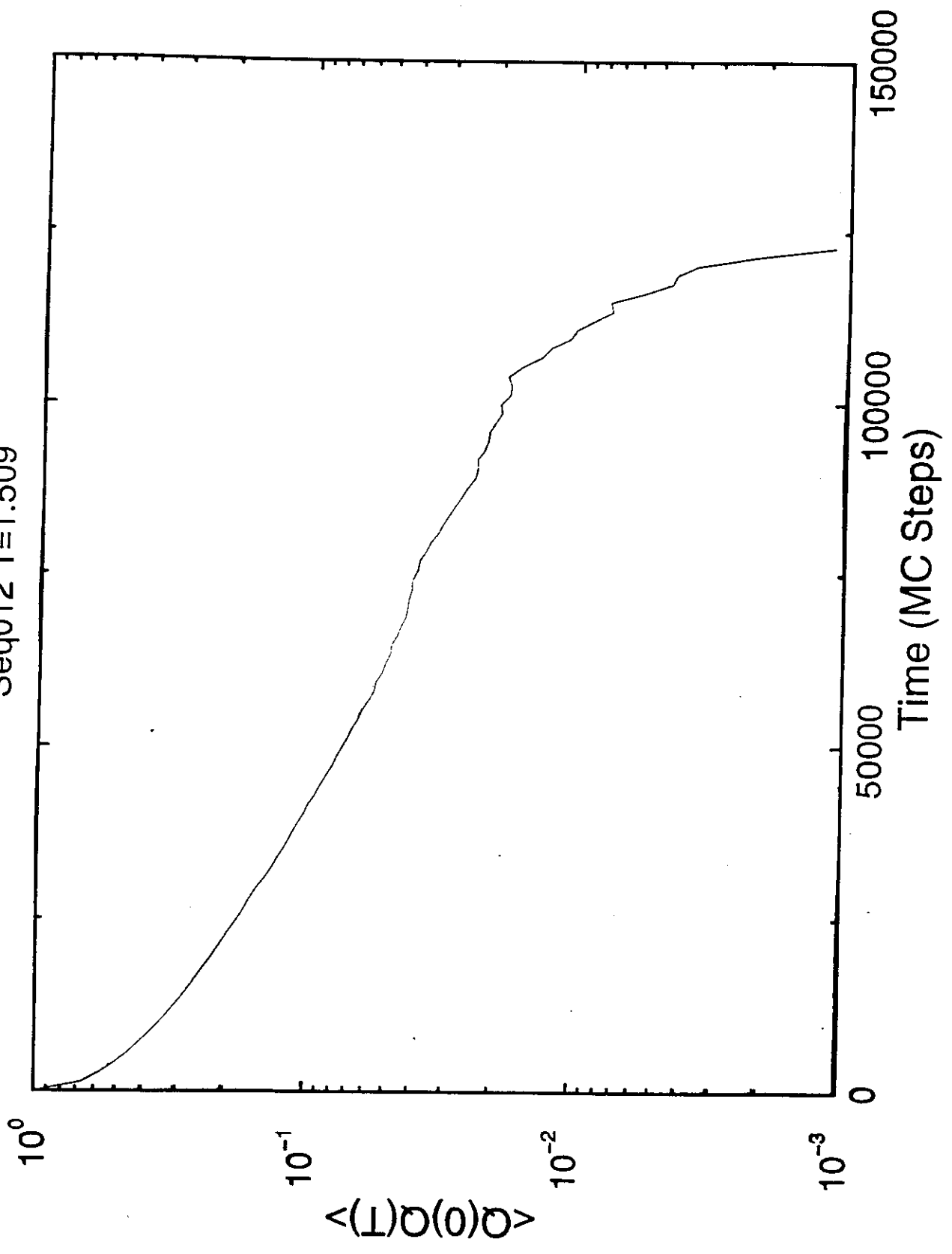




Very sharp
3000 steps

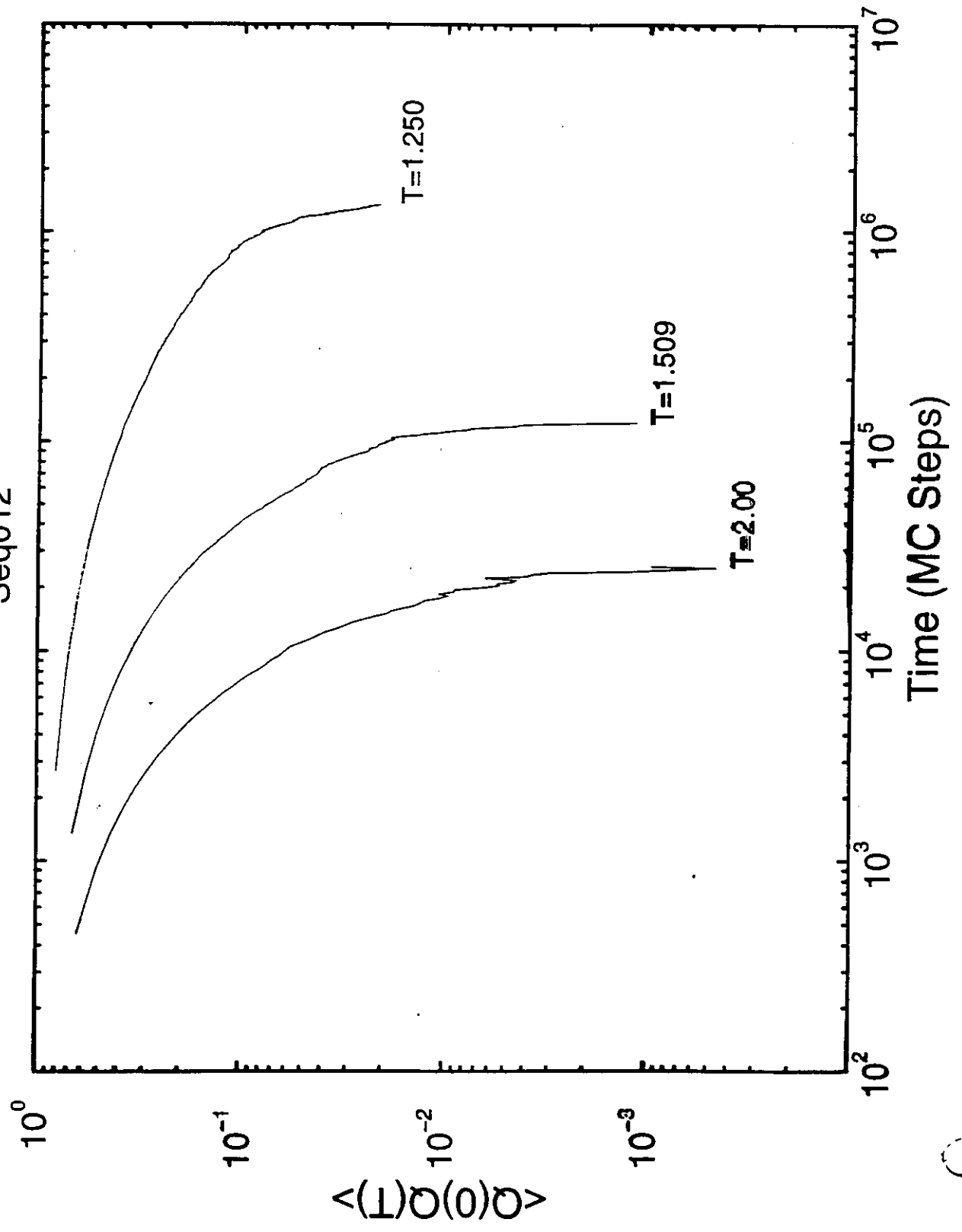
Autocorrelation Function (Q)

Seq012 T=1.509



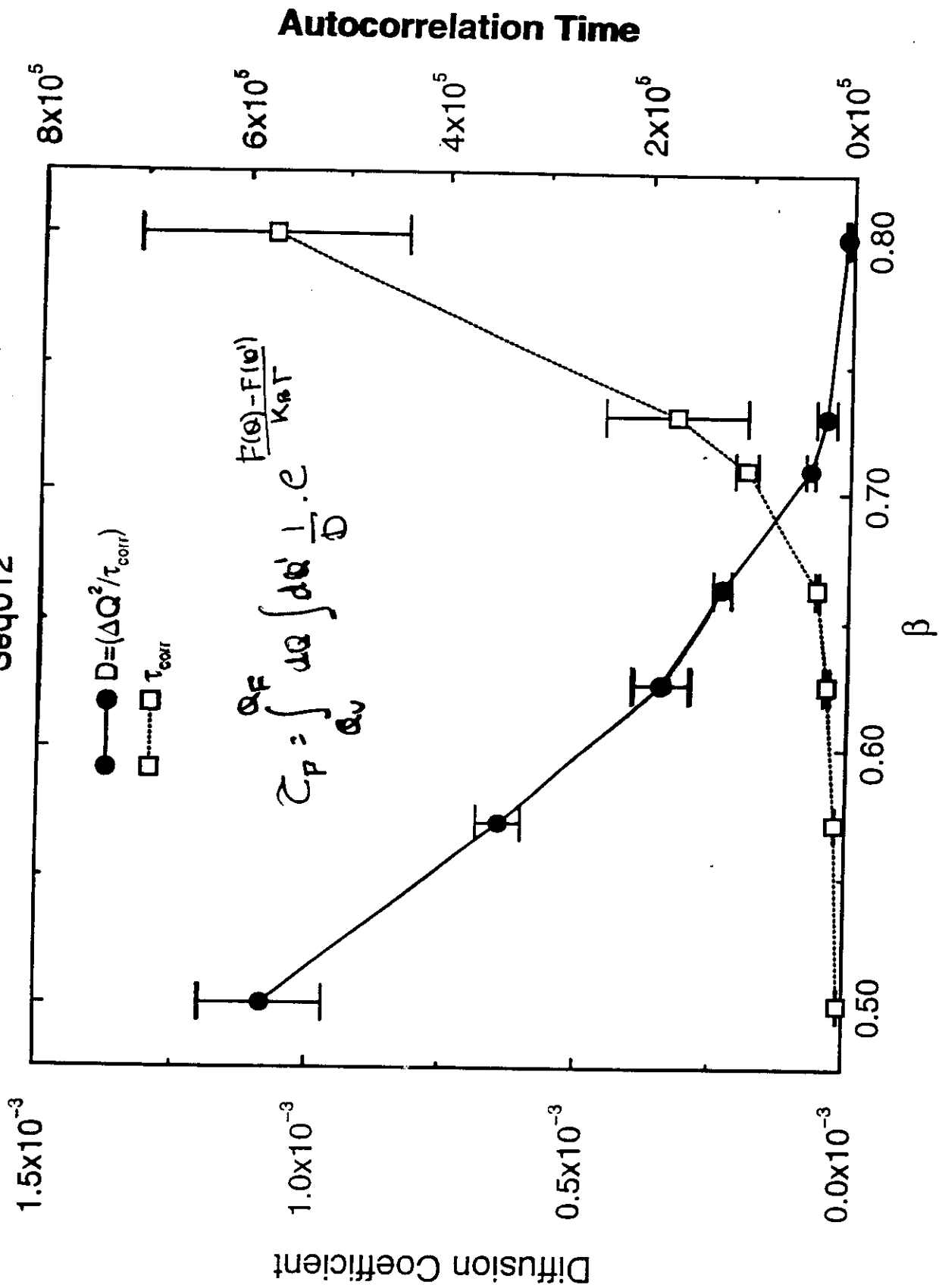
Autocorrelation Function (log-log)

Seq012



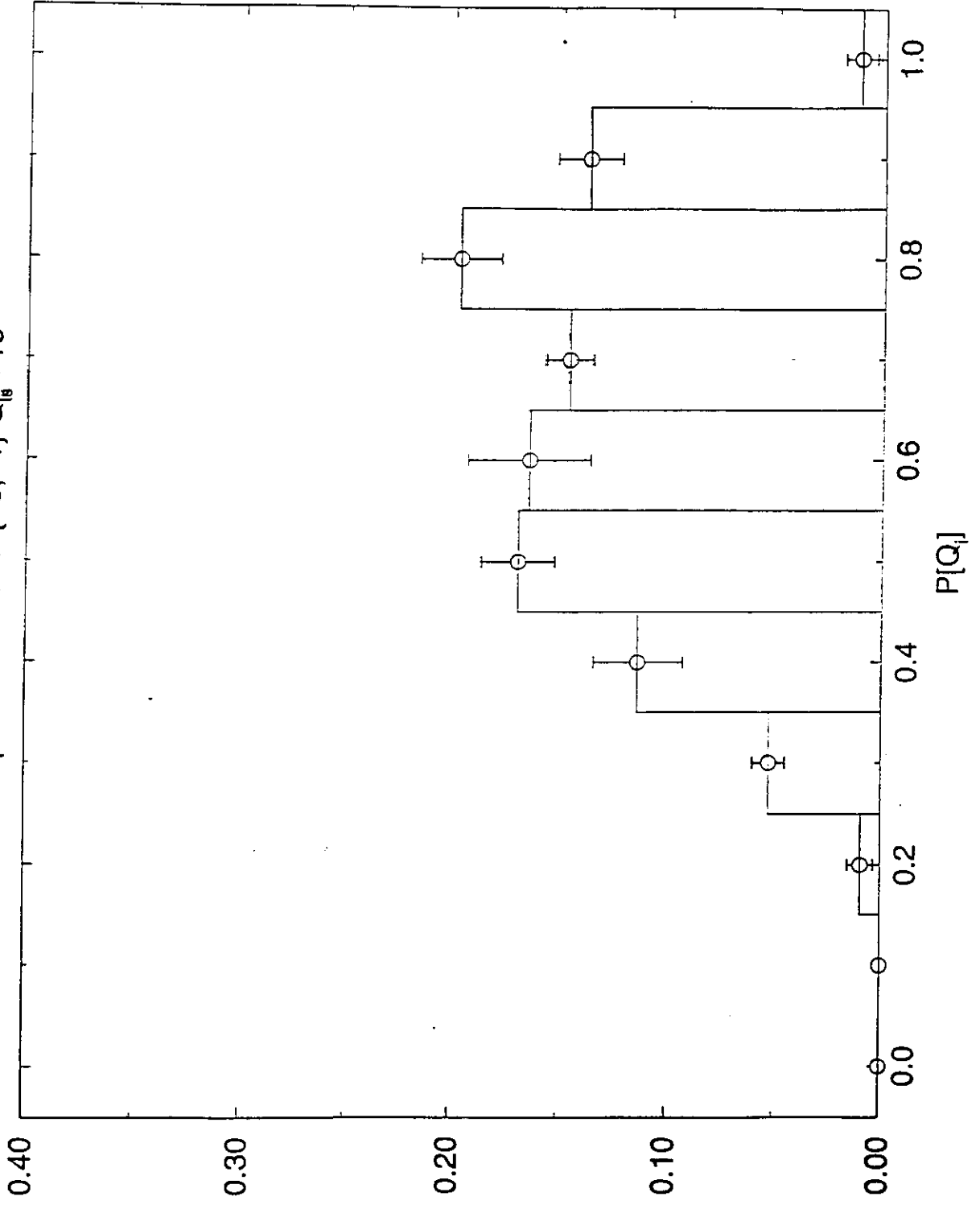
D and τ_{corr} vs Inverse Temperature

Seq012

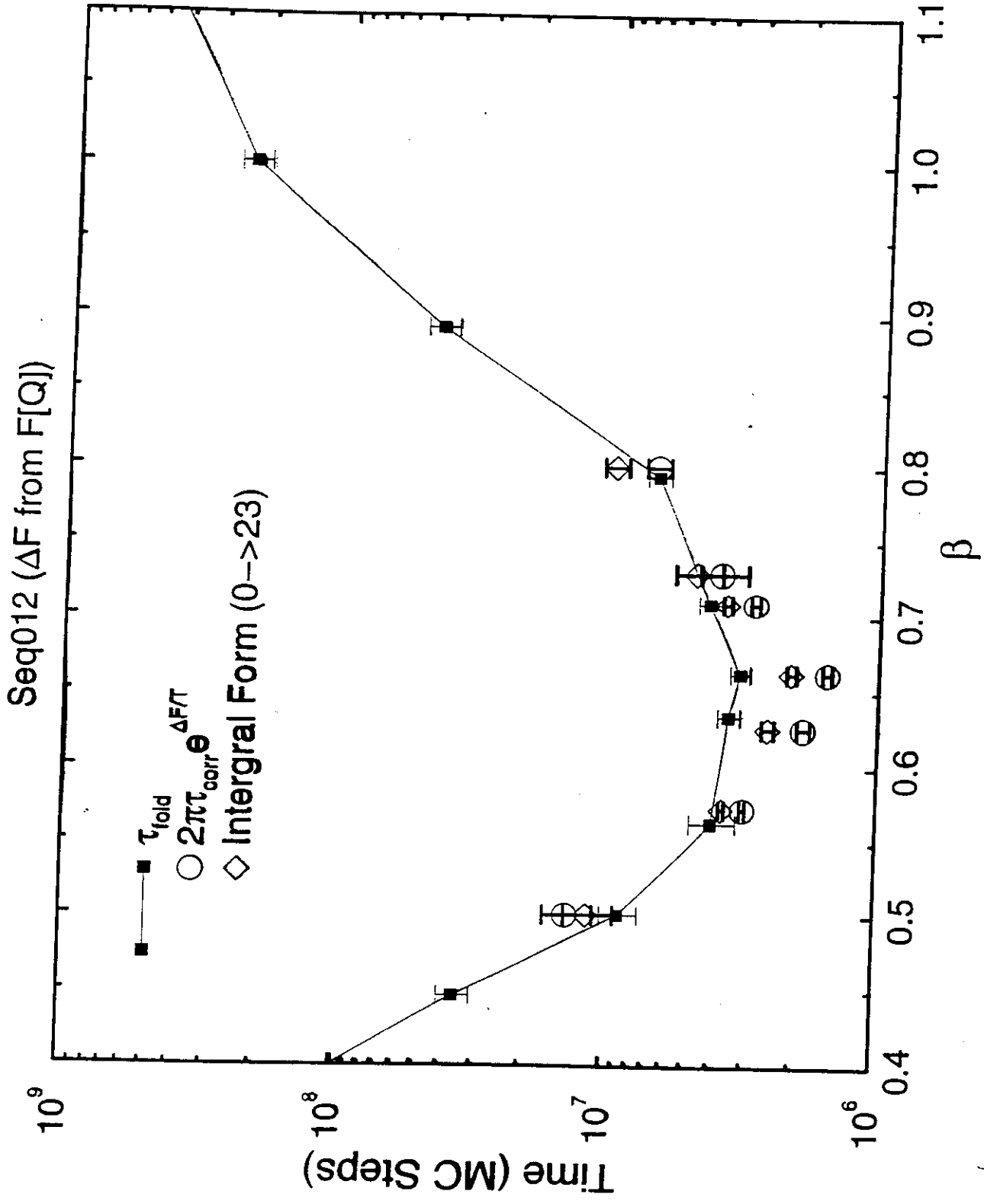


Histogram of Transition State Contact Probability

Seq012 T=1.509 H={-3,-1} Q₁₈ = 18

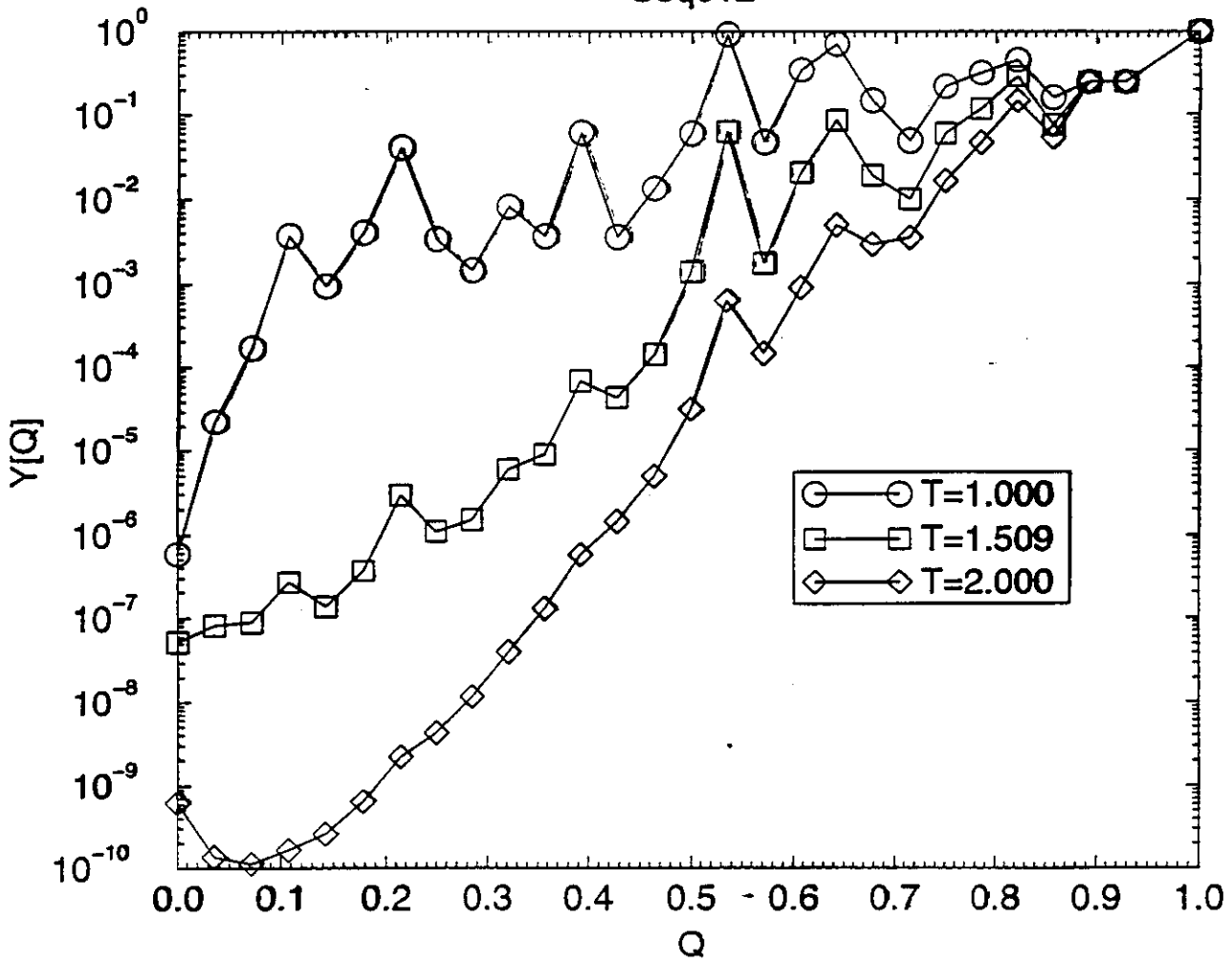


Kramers Rate vs MC MFPT



Y[Q] vs. Q

Seq012



Folding Time

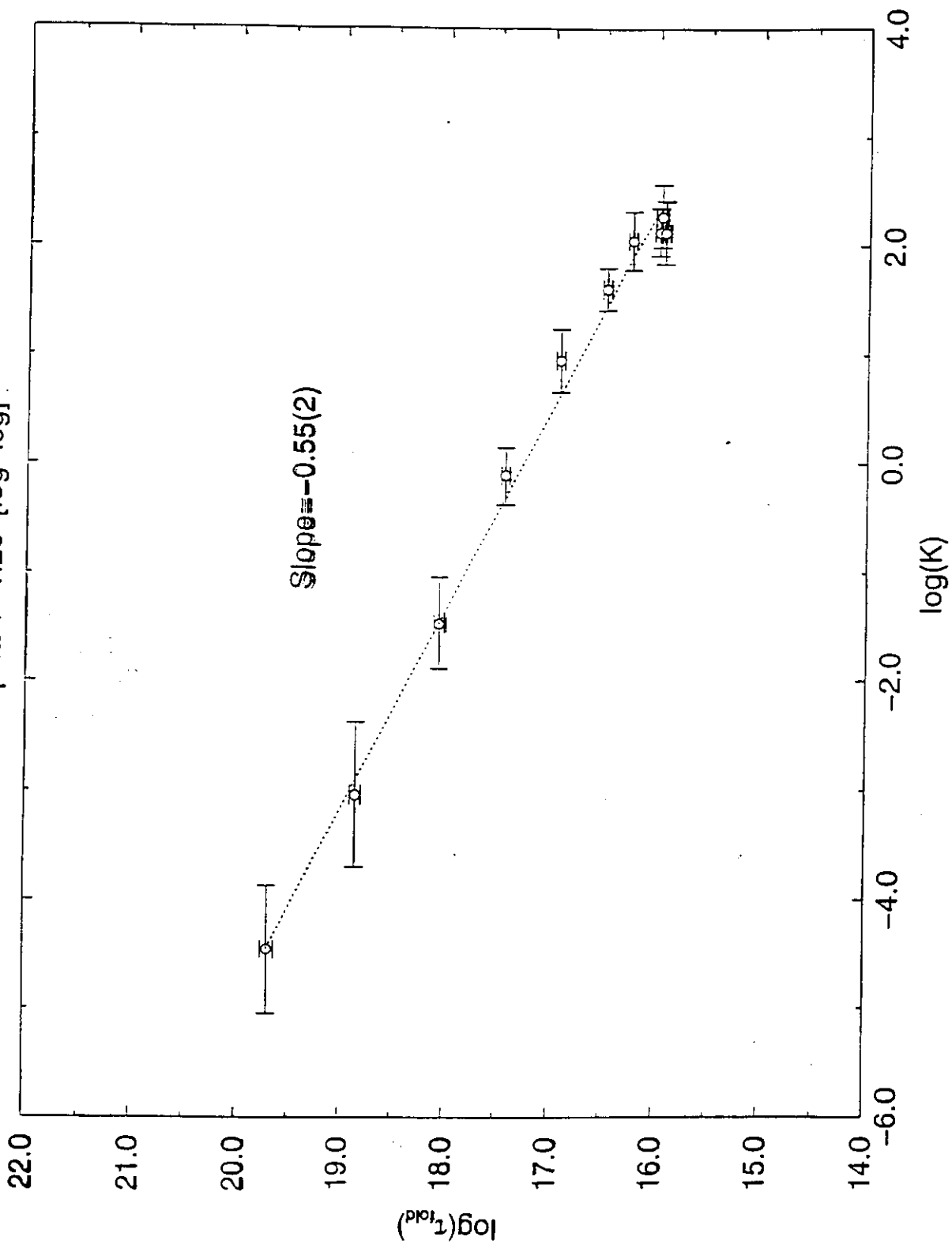
vs.

Denaturant Concentration

Theory and experiments

Folding Time vs Equilibrium Const.

Seq012 T=1.26 [log-log]



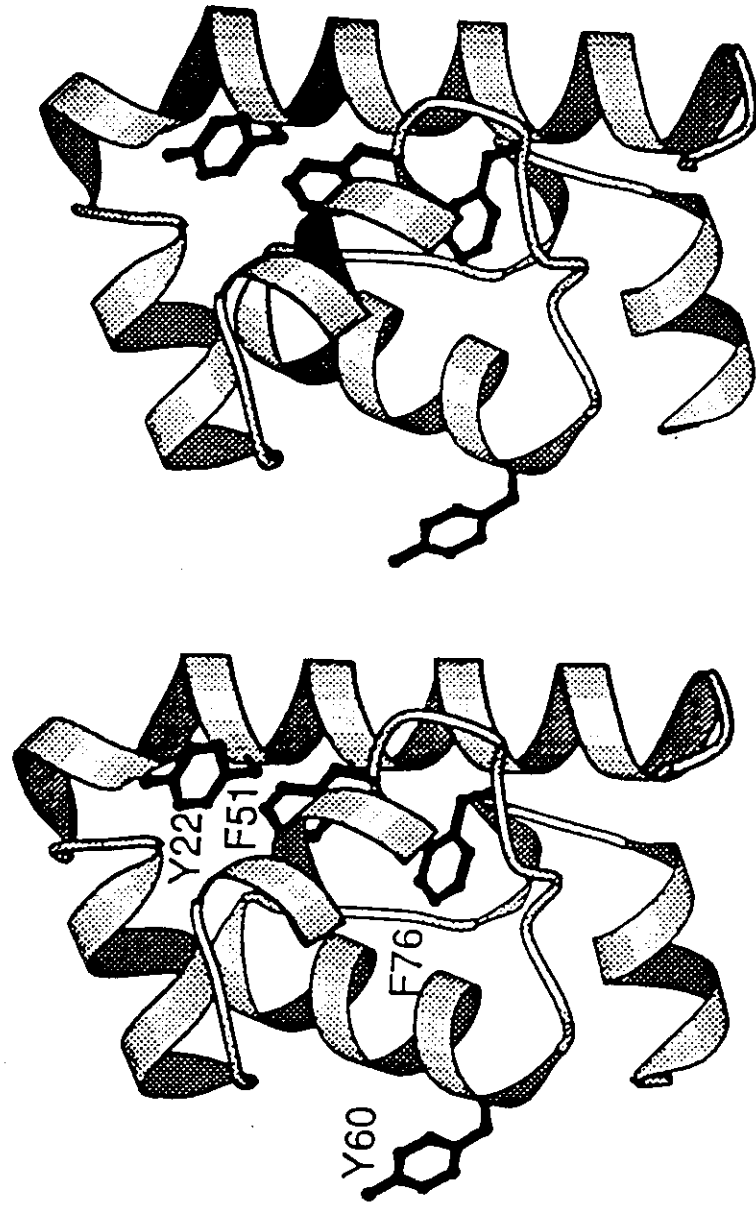


FIG. 1. Stereoview of λ_6 -85 coordinates from the protein-DNA cocrystal structure of Beamer and Pabo (27). Side chains of Tyr-22, Phe-51, Tyr-60, and Phe-76 are indicated. Figure was made with MOT.SCRIPT (29).

urea concentrations, each molecule samples both states several times during the acquisition of the NMR spectrum. This exchange process leads to a shift in peak position and line broadening. As shown in Fig. 3, the aromatic ^1H resonances of

The spectrum of a protein resonance in two-state exchange between native (N) and denatured (D) states is given by the following equation for the intensity as a function of frequency (13):

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 ΔG_0 , $\delta \nu$, T_{2N} ,
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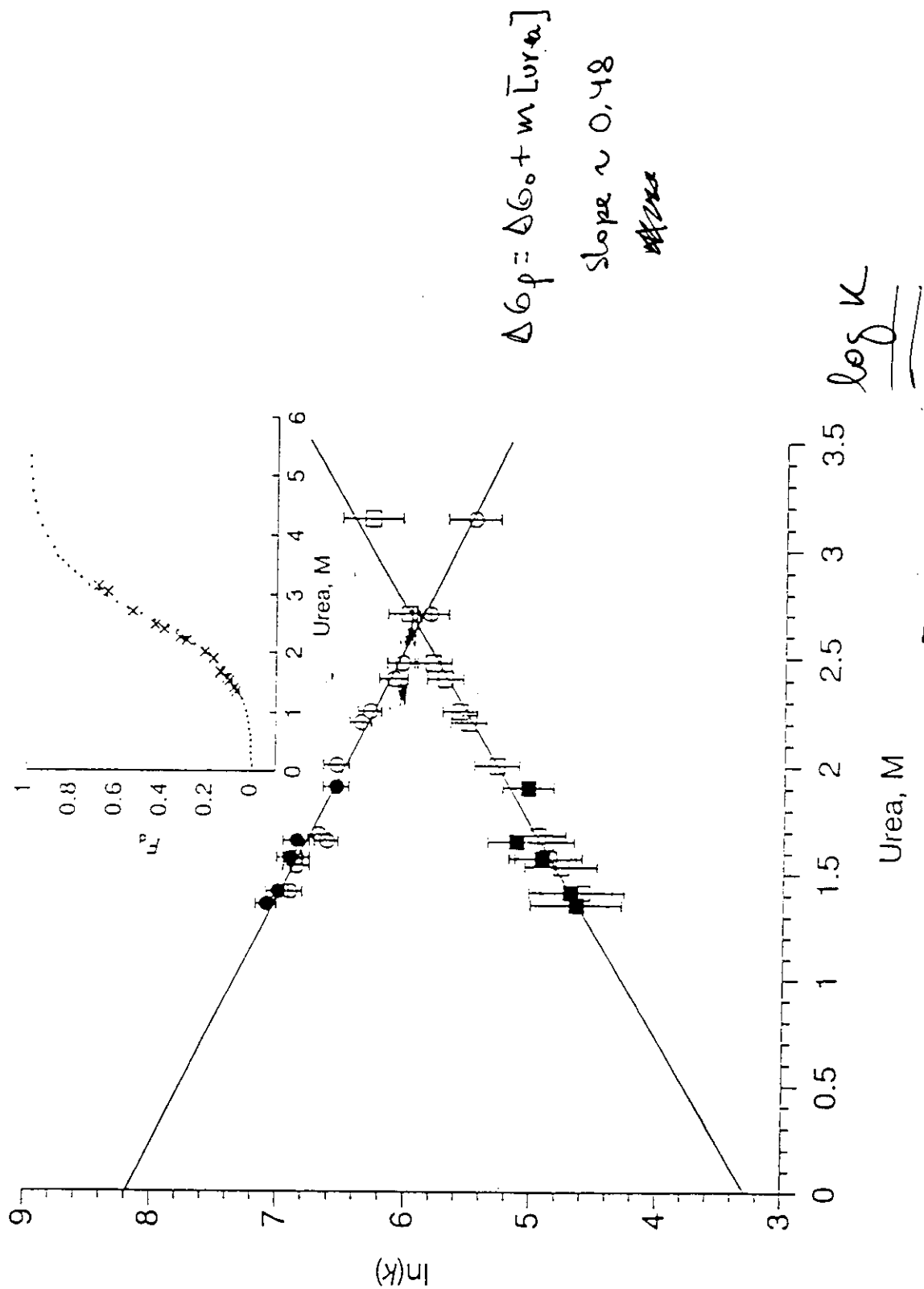


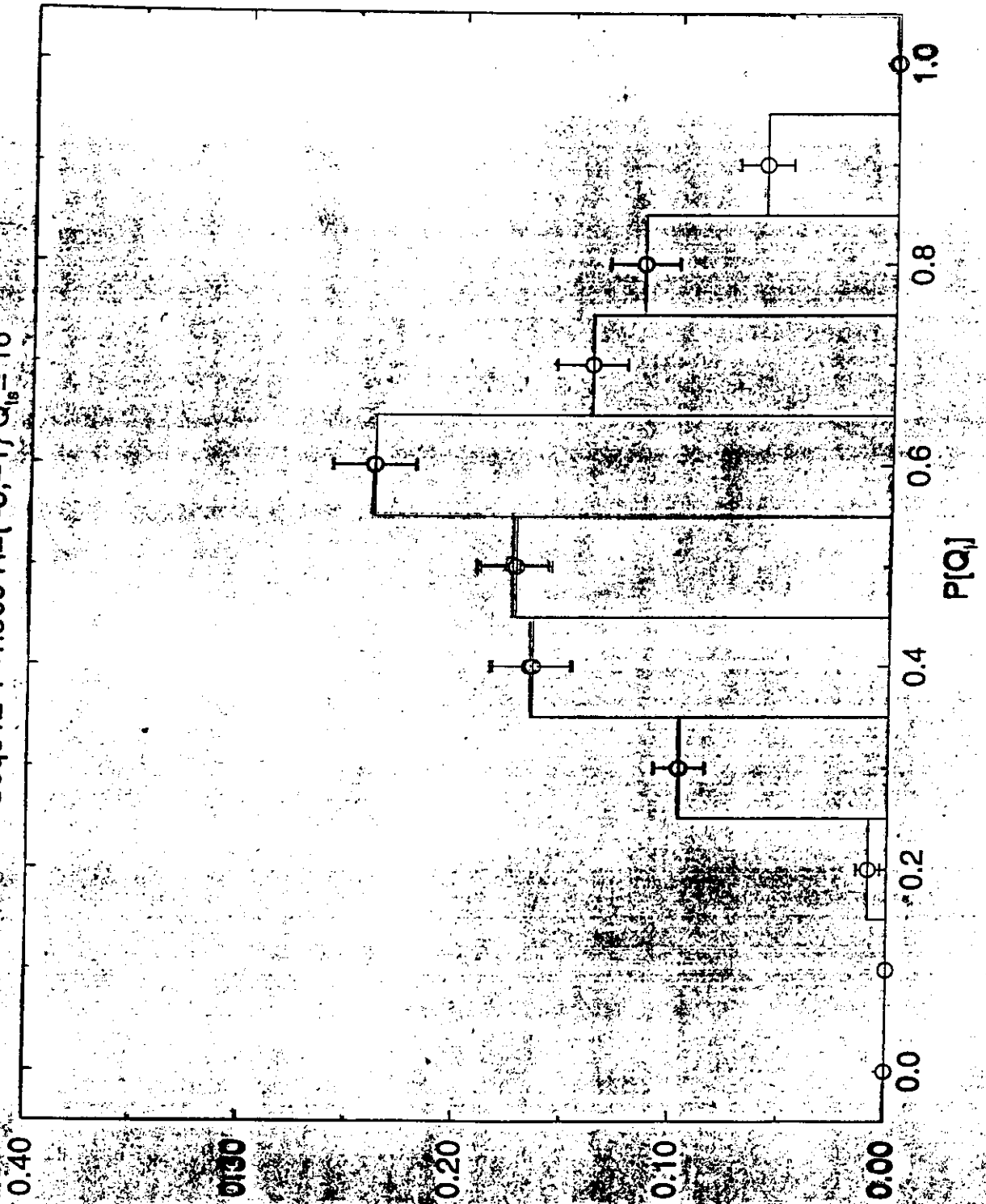
FIG. 5. Natural logarithm of the k_{6-85} folding (circles) and unfolding (squares) rate constants vs. urea concentration at 37°C. Open

Tyr-22 εH

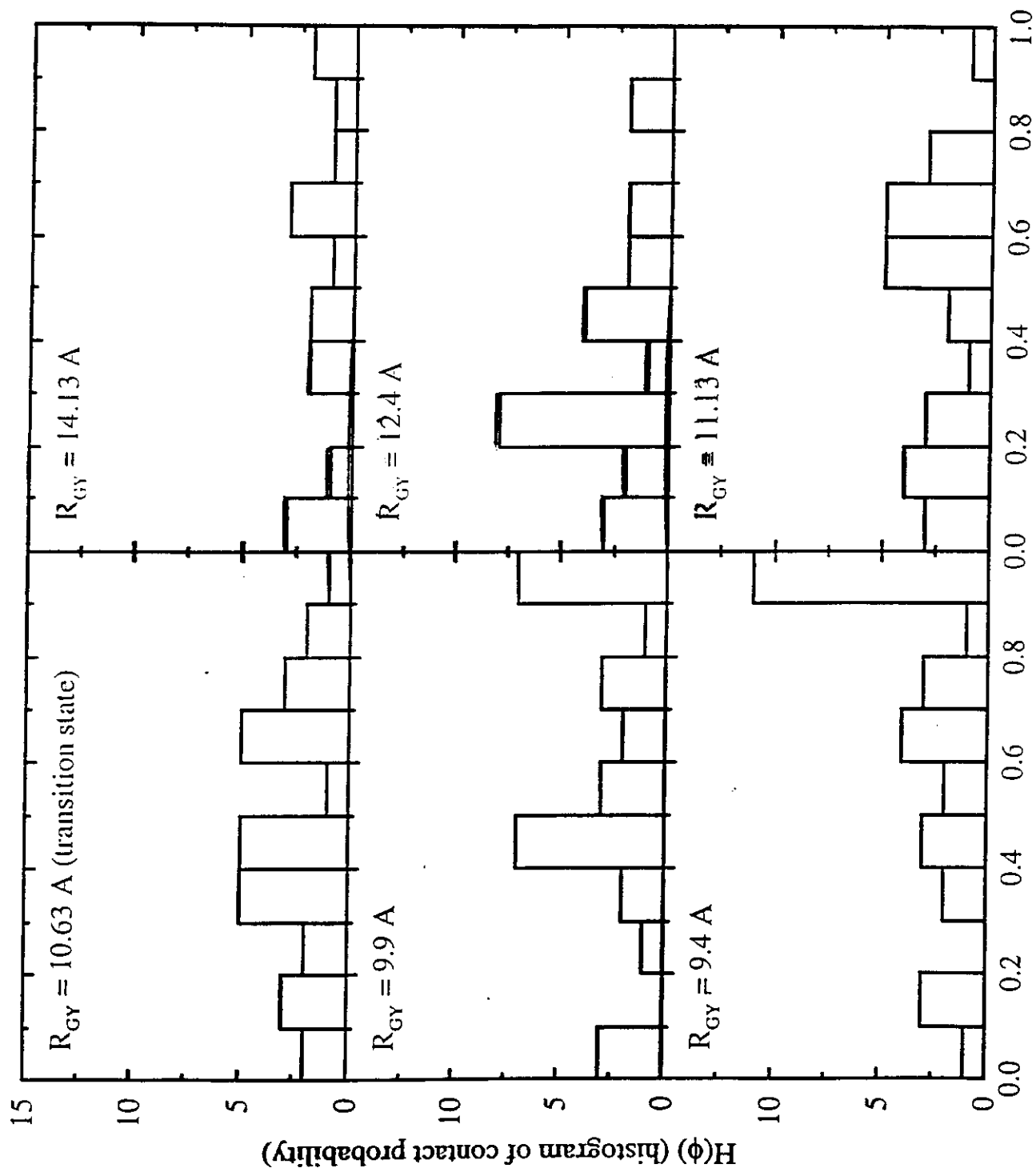
The Nature
of the
Transition State
Ensemble

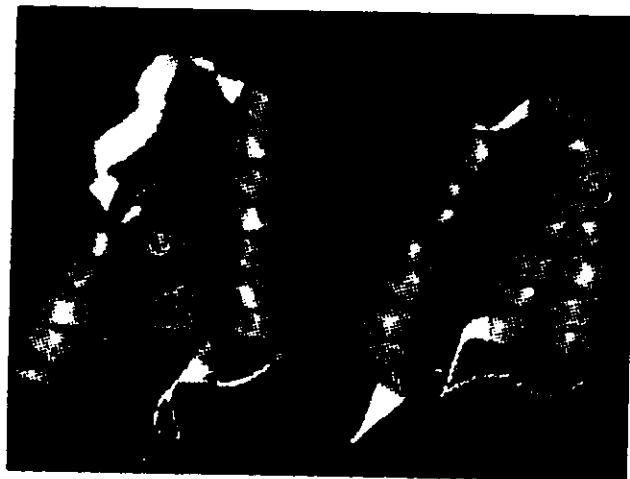
Histogram of Transition State Contact Probability

Seq012 T=1.509 H=(-3,-1) Q₁₆=16



Protein A contact probability histogram vs R_{GY} - Brooks





393

Protein folding
funnel



