

The Abdus Salam International Centre for Theoretical Physics



2037-19

Introduction to Optofluidics

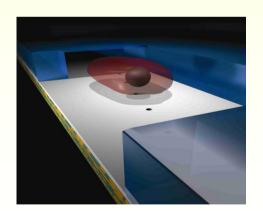
1 - 5 June 2009

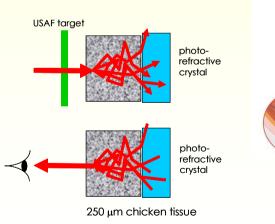
Optofluidic Microscope

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Optofluidic Microscope

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Acknowledgment

My group:

Jigang Wu (microscope, probe) Emily McDowell (turbidity suppression) Xiquan Cui (microscope) Lapman Lee (microscope) Guoan Zhang (microscope) Jian Ren (probe) Sean Pang (microscope) Yin Min Wang (microscope) Meng Cui (turbidity suppression)

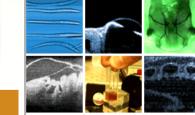
Collaborators:

Demetri Psaltis (EPFL) Paul W. Sternberg (Caltech) Snow Tseng (NTU, Taiwan)



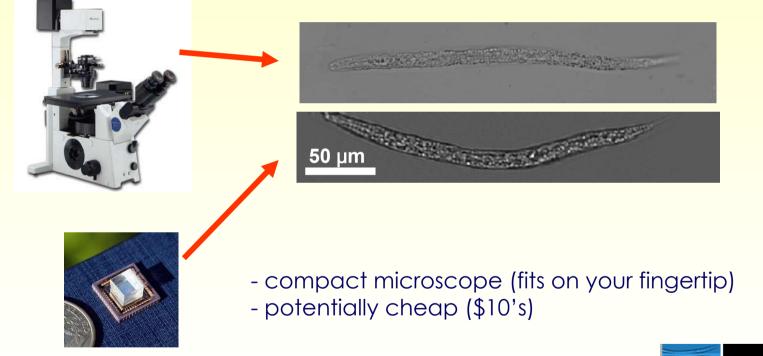
Funding Sources: DARPA Optofluidic Center, NIH, NSF, Coulter Foundation, Caltech Grubstake Program, Industrial Collaborators



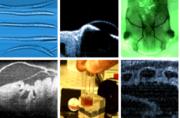


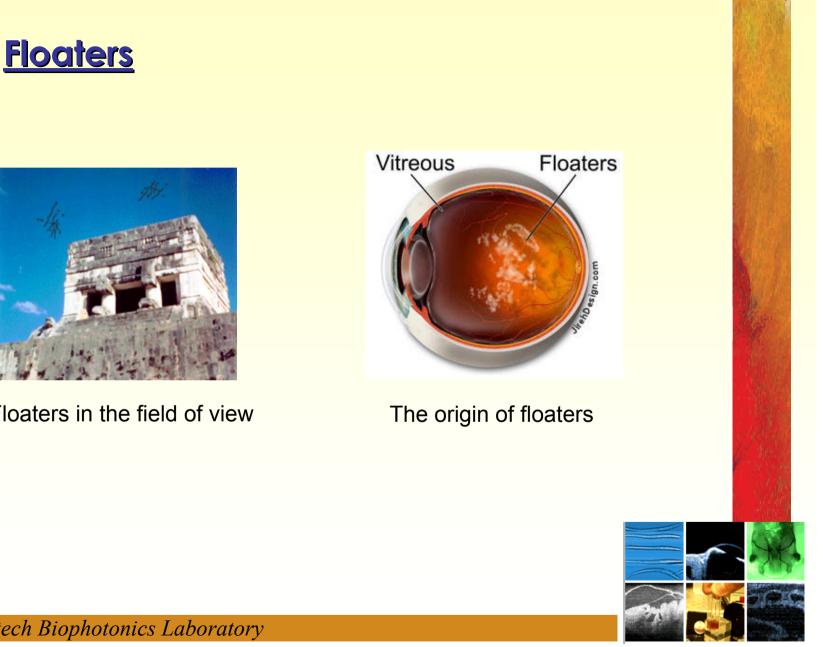
<u>Redesigning the microscope</u>

We abandon the conventional microscopy design and uses a novel aperture array for high resolution imaging Images of C. elegans



Xin Heng, David Erickson, Larry R. Baugh, Zahid Yaqoob, Paul W. Sternberg, Demetri Psaltis, and Changhuei Yang. 'Optofluidic microscopy: A Method for Implementing High Resolution Optical Microscope On A Chip,'Lab on a Chip 6, 1274 (2006).

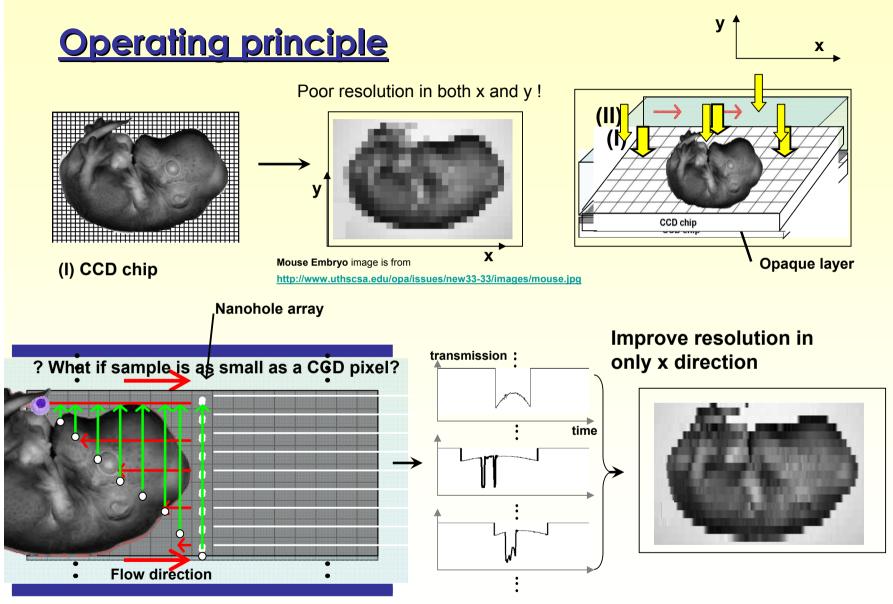






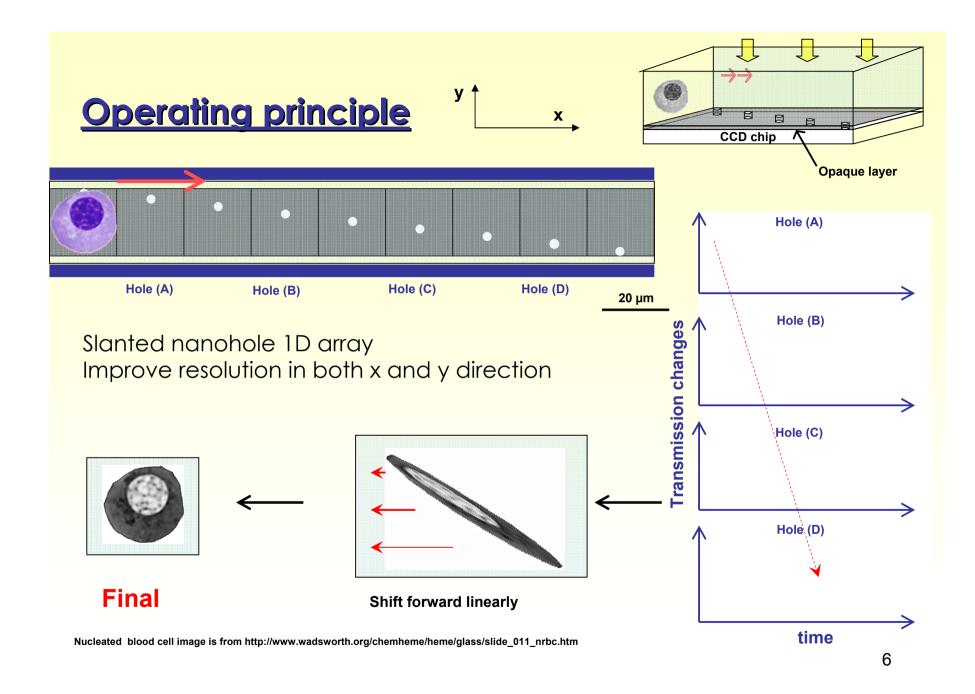
Floaters in the field of view

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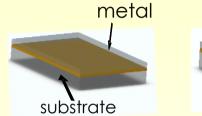
(II) Same CCD but with a column of nanoholes

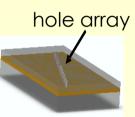
Nucleated blood cell image is from http://www.wadsworth.org/chemheme/heme/glass/slide_011_nrbc.htm

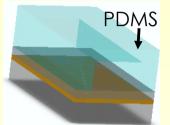


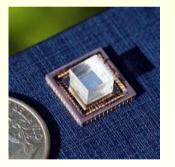
Fabricating the OFM

Fabrication Steps:

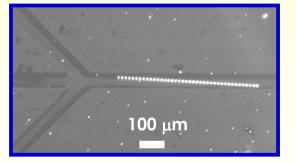






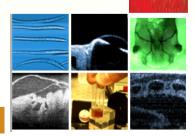


On-chip OFM

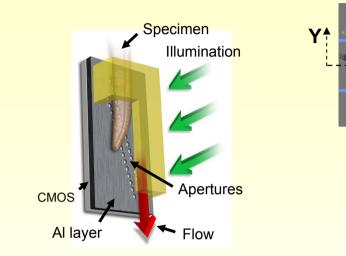


Transmission microscope image of the hole array

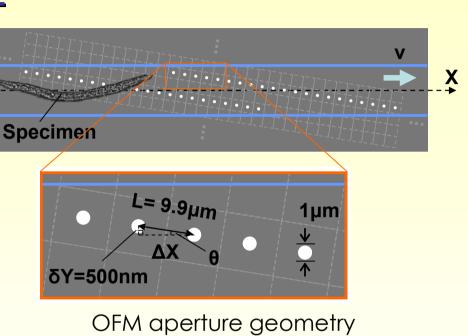
Xin Heng, David Erickson, Larry R. Baugh, Zahid Yaqoob, Paul W. Sternberg, Demetri Psaltis, and Changhuei Yang. 'Optofluidic microscopy: A Method for Implementing High Resolution Optical Microscope On A Chip,'Lab on a Chip 6, 1274 (2006).

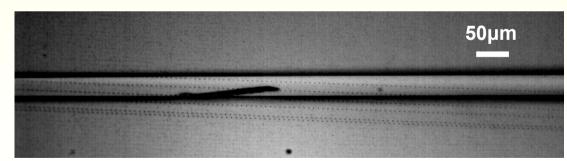


Imaging Strategy

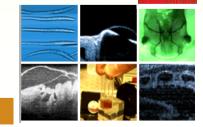


OFM imaging geometry

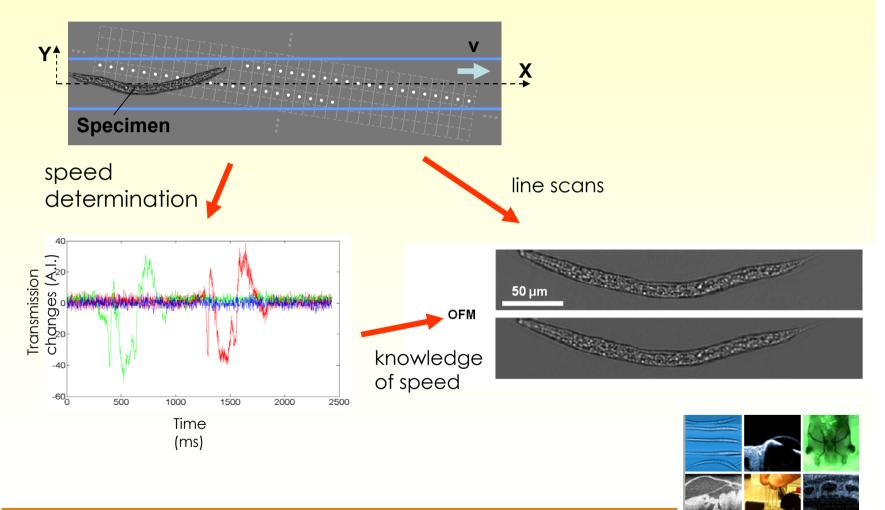




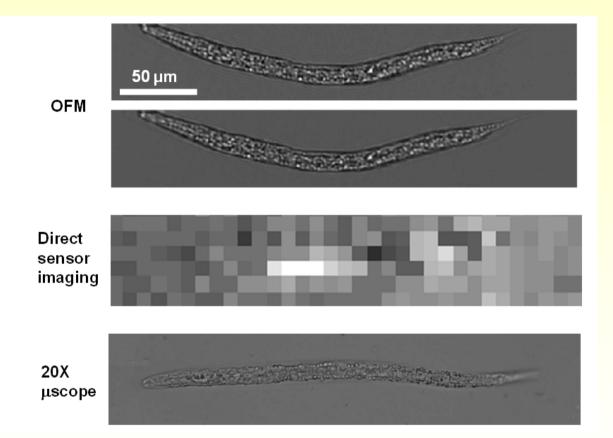
C. elegans moving across the hole array



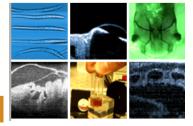
Imaging Strategy



Imaging C. elegans



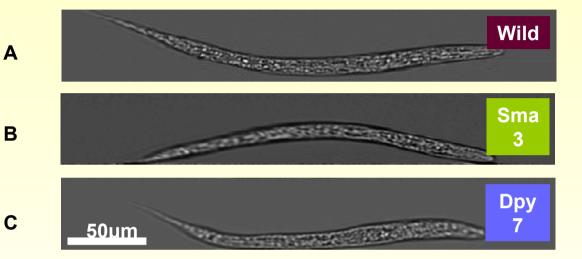
OFM prototype capabilities: Each worm takes 2.5 sec to image Optimal worm processing rate: ~ 2 worms/sec

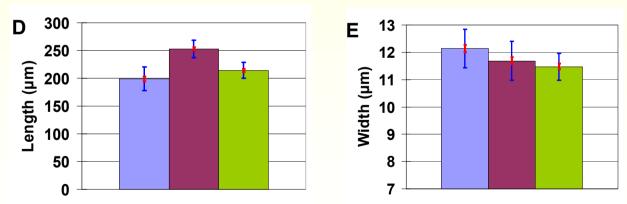


Automated Phenotype Characterization

Perform automated phenotype characterization

- a) automated c) computerized worm length and area
- measurement
- d) drop and go



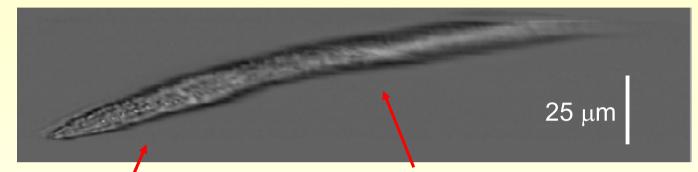


X Cui, L Lee, X Heng, W Zhong, PW Sternberg, D Psaltis & C Yang, PNAS Vol 105, 10670 (2008)

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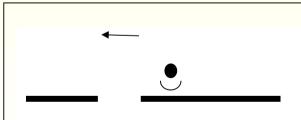
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Resolution Issues

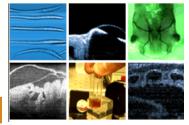


well resolved region that was in close proximity to the hole array blurry region that was not in proximity to the hole array

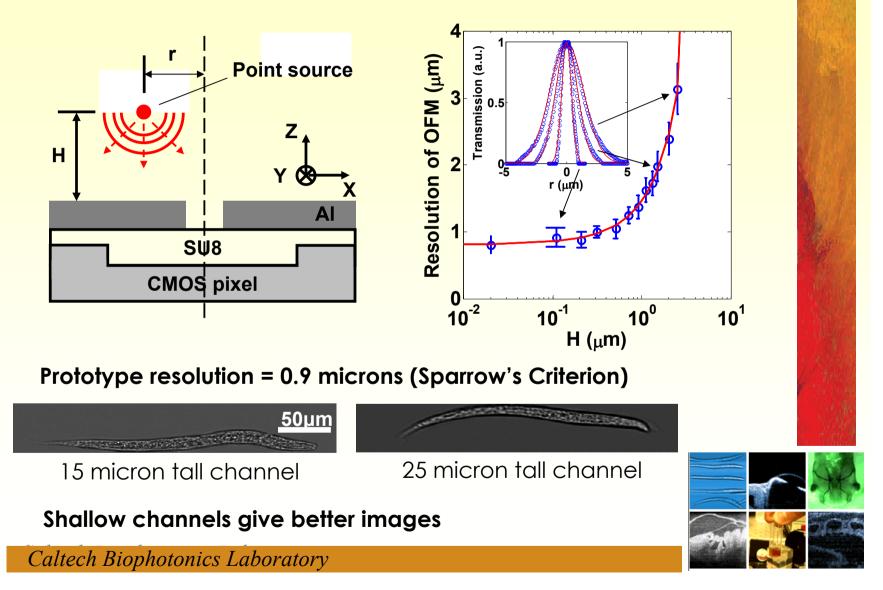
Proximity to hole array is required for good resolution.



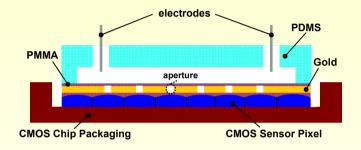
X. Heng, X. Cui, D. W. Knapp, J. Wu, Z. Yaqoob, E. J. McDowell, D. Psaltis, and C. Yang, Optics Express, Vol. 14, No. 22, pp. 10410-10425, October 2006.



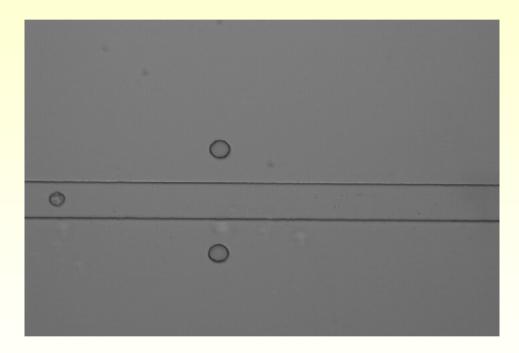
Resolution Issues



Electrokinetic Drive based Optofluidic Microscope (EK-OFM)



The flat velocity profile of EK flow reduces rotations of spherical/ellipsoidal cells in OFM.



50 µm

X Cui, L Lee, X Heng, W Zhong, PW Sternberg, D Psaltis & C Yang, PNAS Vol 105, 10670 (2008)

