



**The Abdus Salam  
International Centre for Theoretical Physics**



**2038-20**

**Conference: From DNA-Inspired Physics to Physics-Inspired Biology**

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**How Proteins Find Their Targets on DNA**

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*USA*



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Department of Chemistry

How Proteins Find Their Targets on  
DNA

# COLLABORATION:

**Prof. A.A. Kornyshev**, Department of Chemistry, Faculty of Physical Sciences, Imperial College London, UK.

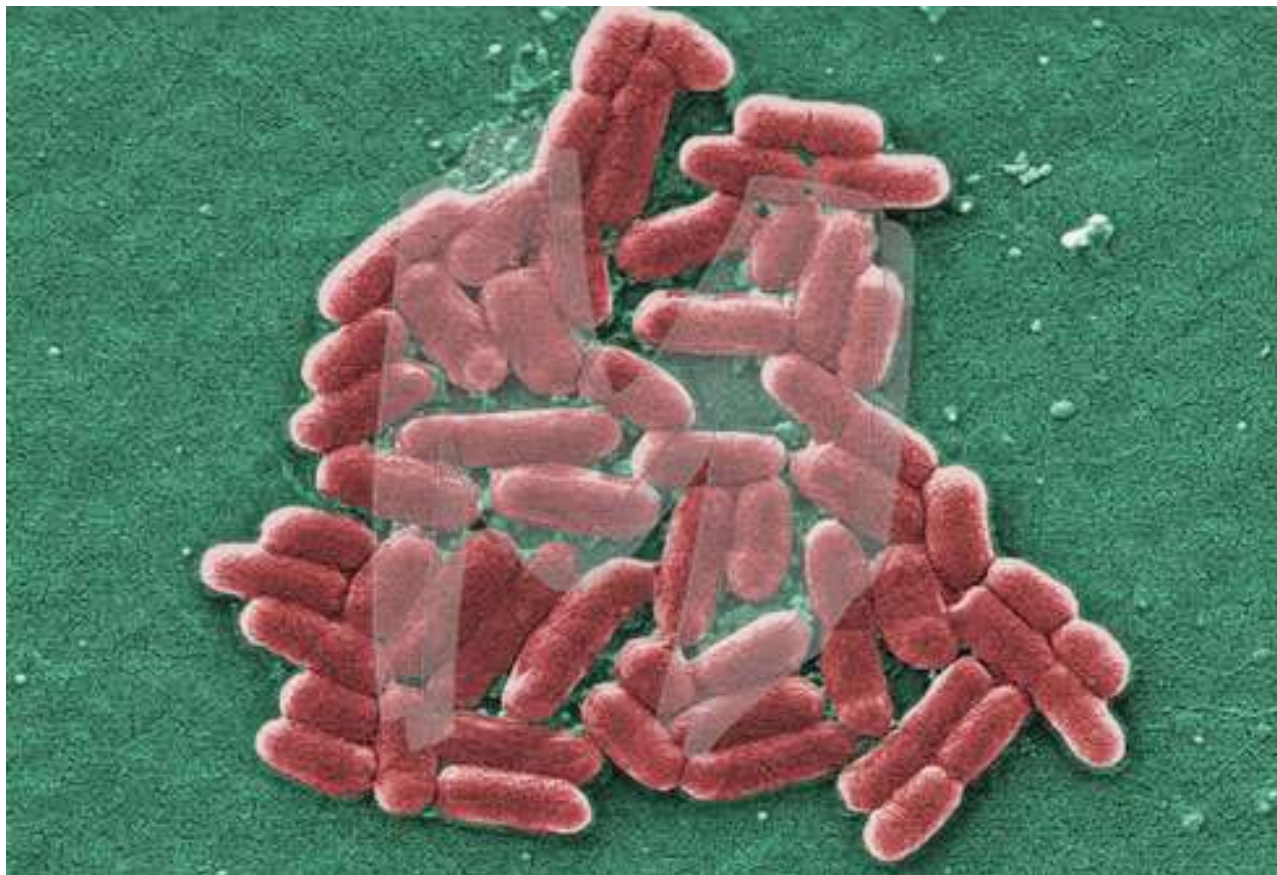
**Dr. A.G. Cherstvy**, Research Center, Juelich, Germany.

**Rahul K. Das**, graduate student in my group

# Protein-DNA Interactions

Play fundamental role in all biological processes

Example #1: switching genes on and off in *Ecoli* bacteria

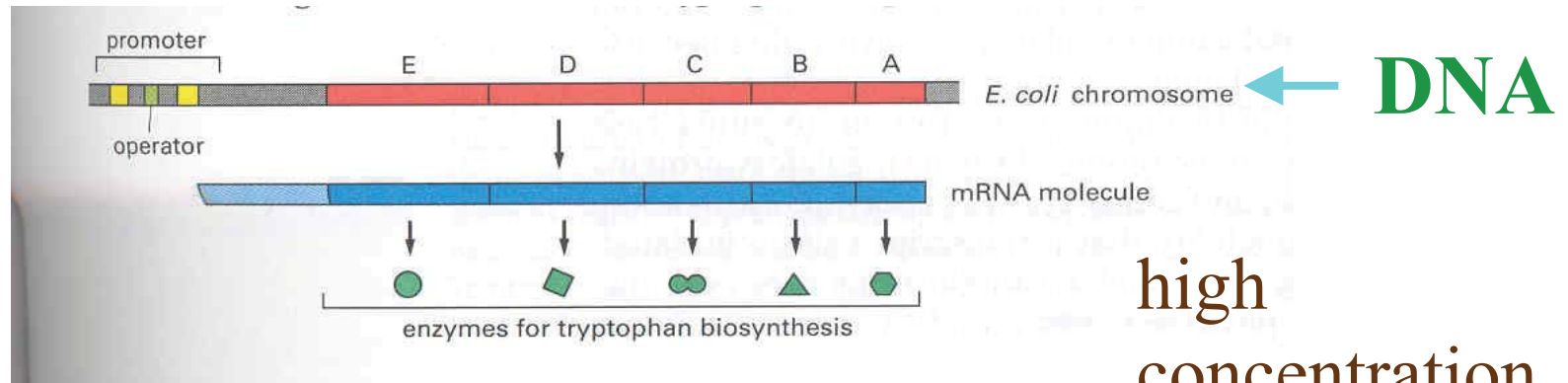


Large numbers of *Ecoli* bacteria live in human intestines

Bacteria needs to produce proteins which are made from aminoacids

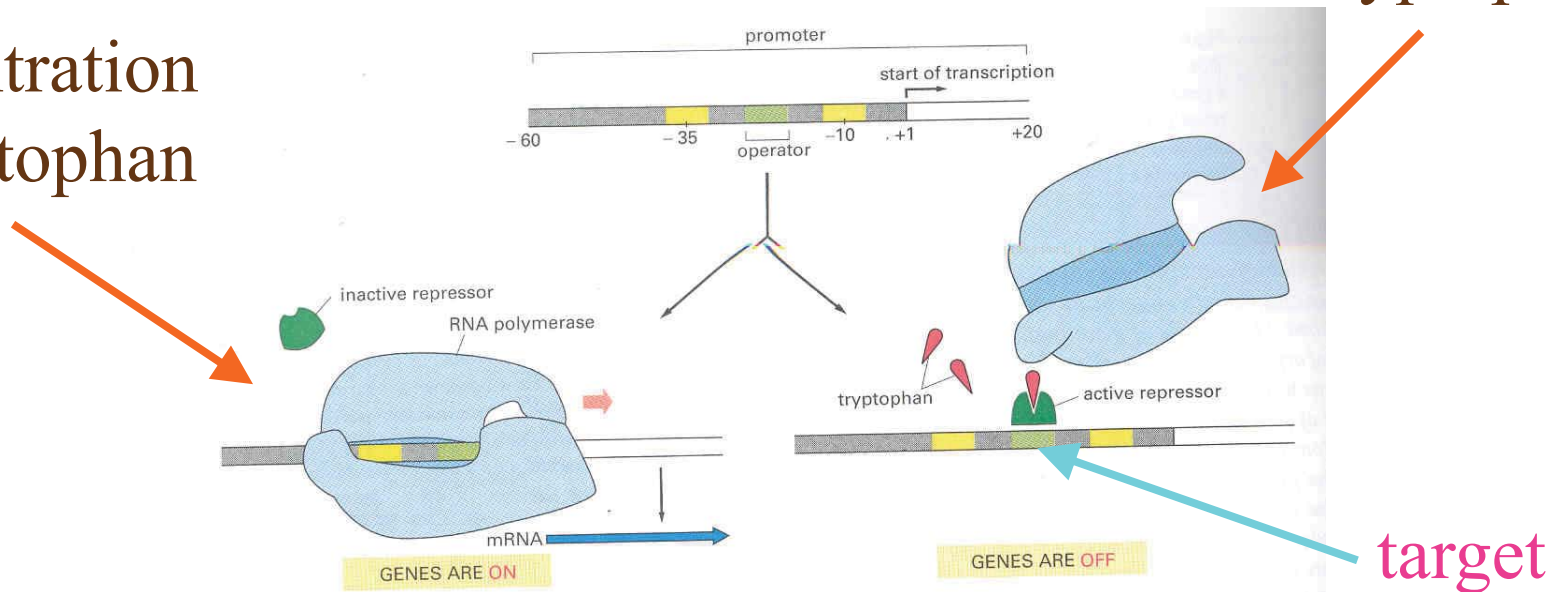
# Gene Switch in Bacteria

## Production of the amino acid tryptophan



high  
concentration  
of tryptophan

low  
concentration  
of tryptophan



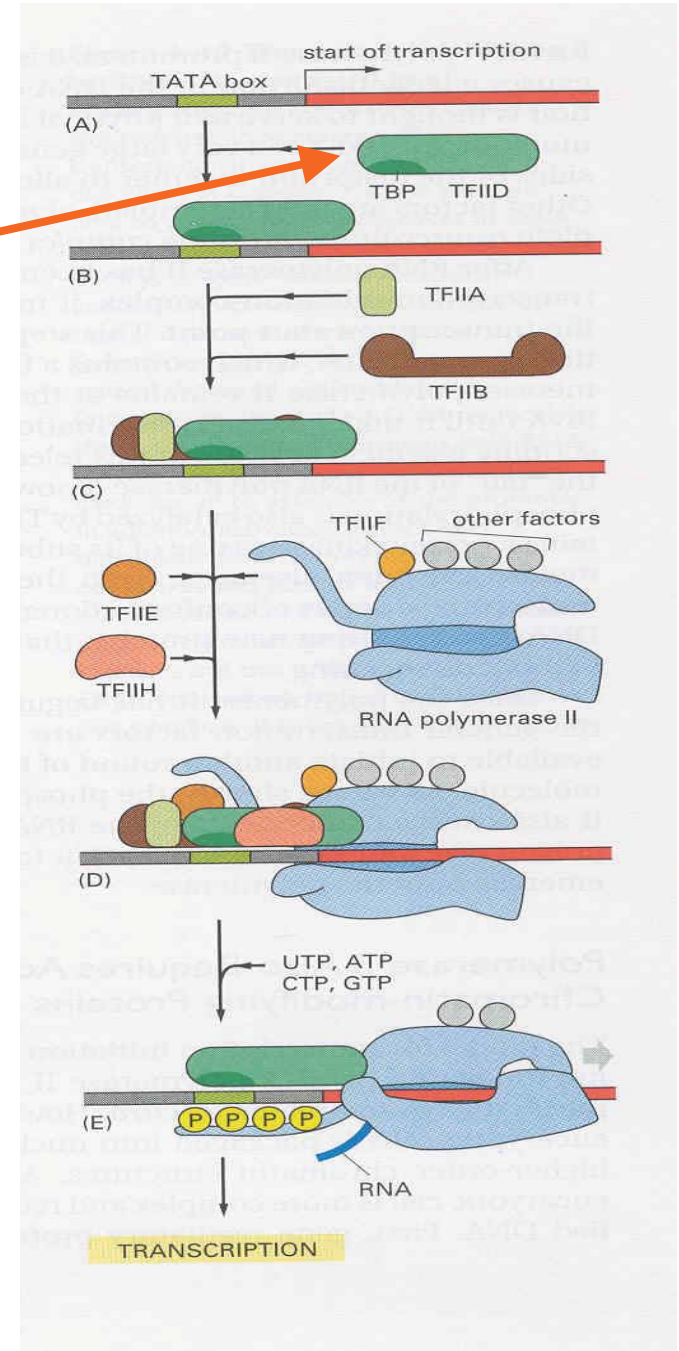
# Transcription

## Example #2:

transcription proteins must find a special sequence (called **TATA box**), 25 bp from the binding site of RNA Polymerase before transcription starts

**Critical step in protein-DNA interactions:**

**protein finding and recognizing the target on DNA**



# Protein-DNA Interactions

**Experimental Observations:** Lac repressor protein finds its target with association rate  $k_{\text{ass}} \sim 10^{10} \text{ M}^{-1} \text{ s}^{-1}$  (!!!)

Riggler et al., *J. Mol. Biol.* **53** 401-417 (1970)

100-1000 times larger than the maximal rate by three-dimensional diffusion-controlled search (from Debye-Smoluchowski theory)

The phenomenon is called **Facilitated Diffusion**

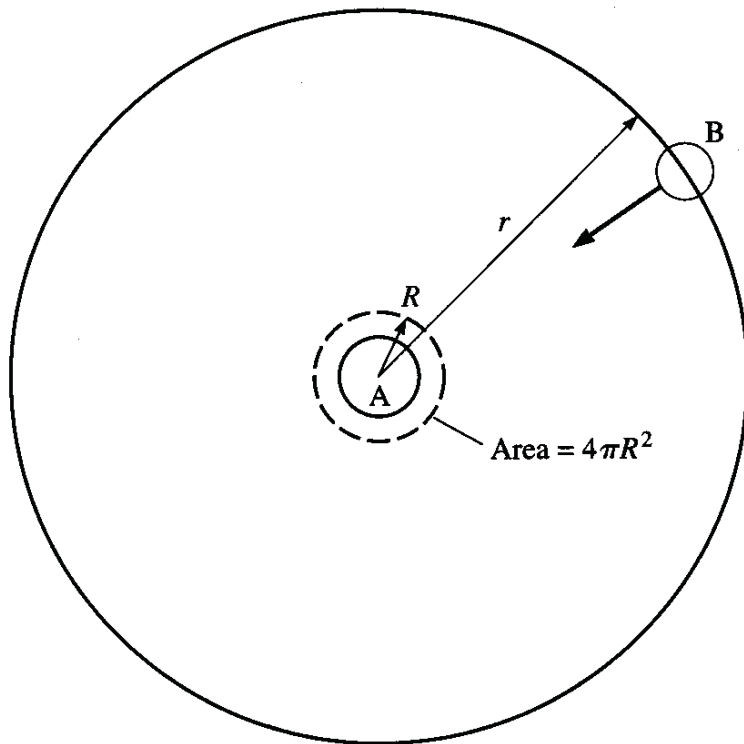
Generated huge controversy – still unresolved

# Diffusion-Controlled Rate Constants

Compound B reacts with compound A  
after approaching it within the radius  $R$

Debye-Smoluchowski

$$k_{\max} = 4\pi(D_A + D_B)R$$



For protein-DNA interactions it is assumed that DNA does not move, i.e.,  $D_A=0$ , target size is few bases -  $R \sim 1$  nm, for a typical protein  $D_B \sim 10^{-11} \text{ m}^2 \text{ s}^{-1}$

$$k_{\max} \sim 10^8 \text{ M}^{-1} \text{ s}^{-1}, \text{ compare with } k_{\text{exp}} \sim 10^{10} \text{ M}^{-1} \text{ s}^{-1}$$

# Possible Resolution?

## An end to 40 years of mistakes in DNA–protein association kinetics?

Stephen E. Halford<sup>1</sup>

The DNA–Proteins Interaction Unit, Department of Biochemistry, School of Medical Sciences, University of Bristol, Bristol BS8 4DL, U.K.

### Abstract

Proteins that bind to specific sequences in long DNA molecules have to locate their target sites amid myriad alternative sequences, yet they do so at remarkably rapid rates, sometimes approaching  $10^{10} \text{ M}^{-1} \cdot \text{s}^{-1}$ . Hence, it has been asserted widely that binding to specific DNA sites can surpass the maximal rate for 3D (three-dimensional) diffusion through solution and that this could only be accounted for by a reduction in the dimensionality of the search for the target in effect by 1D (one-dimensional) diffusion (or ‘sliding’) along the DNA contour. It will be shown here that there is, in fact, no known example of a protein binding to a specific DNA site at a rate above the diffusion limit, and that the rapidity of these reactions is due primarily to electrostatic interactions between oppositely charged molecules. It will also be shown that, contrary to popular belief, reduced dimensionality does not, in general, increase the rate of target-site location but instead reduces it. Finally, it will be demonstrated that proteins locate their target sites primarily by multiple dissociation/reassociation events to other (nearby or distant) sites within the same DNA molecule, and that 1D diffusion is limited to local searches covering  $\sim 50$  bp around each landing site.

# Possible Resolution? NO!!!

S.E. Halford, *Biochem Soc. Trans.* 2009, 37, 343-348

Original experiments have been carried out in low-salt conditions:

10 mM of KCl, Tris/HCl, and magnesium acetate

transactions

isotrans.org

## An end to 40 years of mistakes in DNA-protein association kinetics?

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### Abstract

Proteins that bind to specific sequences in long DNA molecules have to locate their target sites amid myriad alternative sequences, yet they do so at remarkably rapid rates, sometimes approaching  $10^{10} \text{ M}^{-1} \cdot \text{s}^{-1}$ . Hence, it has been asserted widely that binding to specific DNA sites can surpass the maximal rate for 3D (three-dimensional) diffusion through solution and that this could only be accounted for by a reduction in the dimensionality of the search for the target in effect by 1D (one-dimensional) diffusion (or 'sliding') along the DNA contour. It will be shown here that there is, in fact, no known example of a protein binding to a specific DNA site at a rate above the diffusion limit, and that the rapidity of these reactions is due primarily to electrostatic interactions between oppositely charged molecules. It will also be shown that, contrary to popular belief, reduced dimensionality does not, in general, increase the rate of target-site location but instead reduces it. Finally, it will be demonstrated that proteins locate their target sites primarily by multiple dissociation/reassociation events to other (nearby or distant) sites within the same DNA molecule, and that 1D diffusion is limited to local searches covering  $\sim 50$  bp around each landing site.

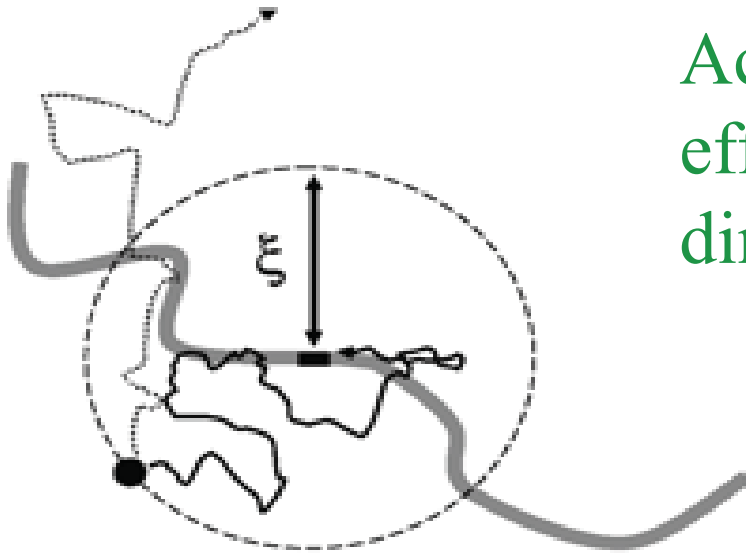
Debye length – separates region where electrostatics is important

In this system it is  $\sim 1$ -2 nm, comparable with the target size.

**Electrostatics does not play role in the fast search!**

# Facilitated Diffusion: Current Theoretical Views

Protein search for the target is viewed as a sequence of 3D excursions and 1D sliding along the DNA



Acceleration due to increase in the effective target length or lowering dimensionality; assumed that  $D_1 \sim D_3$

$$k_{\max} = 4\pi(D_A + D_B)R$$

S.E. Halford and J.F. Marko, *Nucl. Acids Res.* **32**, 340 (2004)

Experiments support facilitated diffusion picture:

D.M. Gowers, G.G. Wilson and S.E. Halford, *PNAS USA*, **102**, 15883 (2005)

# Problems:

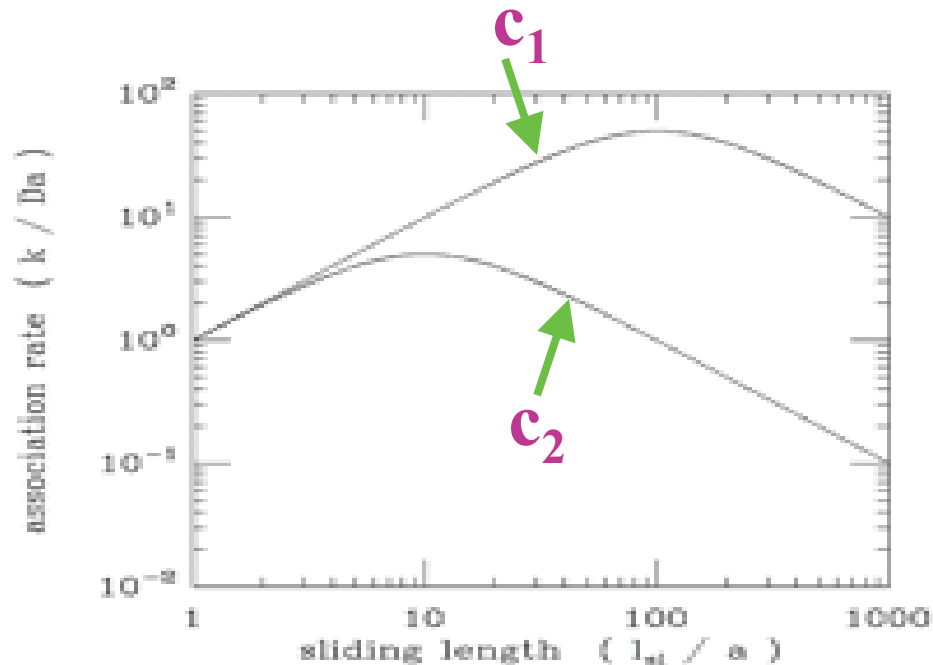
Current theories predict:  $D_1 \sim D_3 \sim 10^{-11} \text{ m}^2 \text{ s}^{-1}$ , and  $\tau(1D) \sim \tau(3D)$

Single-molecule experiments:  $D_1 \sim 10^{-13} - 10^{-16} \text{ m}^2 \text{ s}^{-1}$ ,

$\tau(1D)/\tau(3D) > 10 - 100$



*Phys. Rev. Lett.*, **97**, 048302 (2006); *Science*, **316**, 1191 (2007)



$$c_1 < c_2$$

**What is wrong with this picture?**

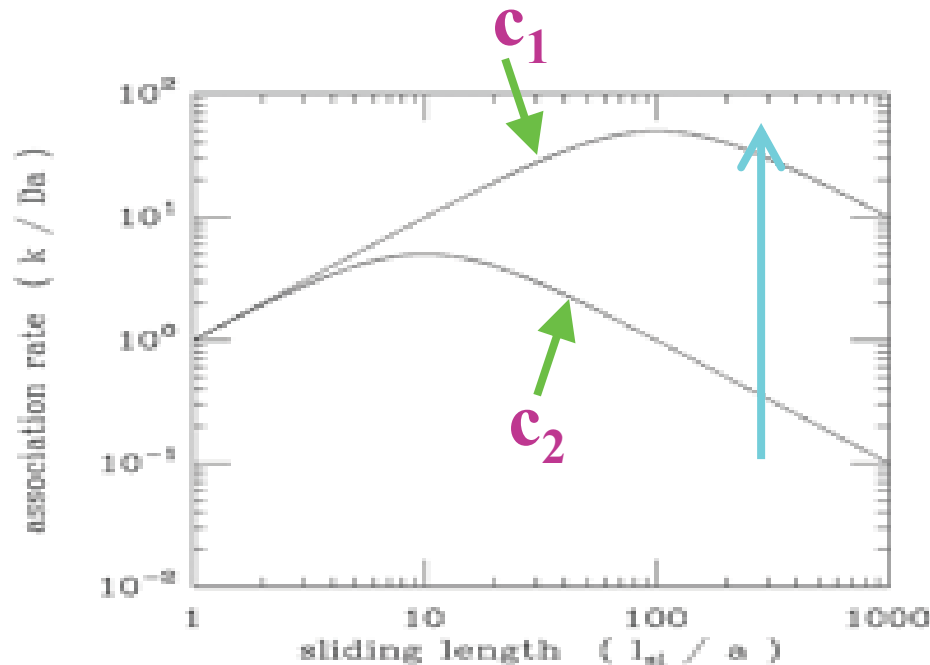
S.E. Halford and J.F. Marko, *Nucl. Acids Res.* **32**, 340 (2004)

# Problems:

Current theories predict:  $D_1 \sim D_3 \sim 10^{-11} \text{ m}^2 \text{ s}^{-1}$ , and  $t_1 \sim t_3$

Single-molecule experiments:  $D_1 \sim 10^{-13} - 10^{-16} \text{ m}^2 \text{ s}^{-1}$ ,  $t_1/t_3 > 10$

*Phys. Rev. Lett.*, **97**, 048302 (2006); *Science*, **316**, 1191 (2007)



$$c_1 < c_2$$

decreasing the concentration of proteins the reaction of association increases!

Contradicts Chemistry!

Unphysical behavior

S.E. Halford and J.F. Marko, *Nucl. Acids Res.* **32**, 340 (2004)

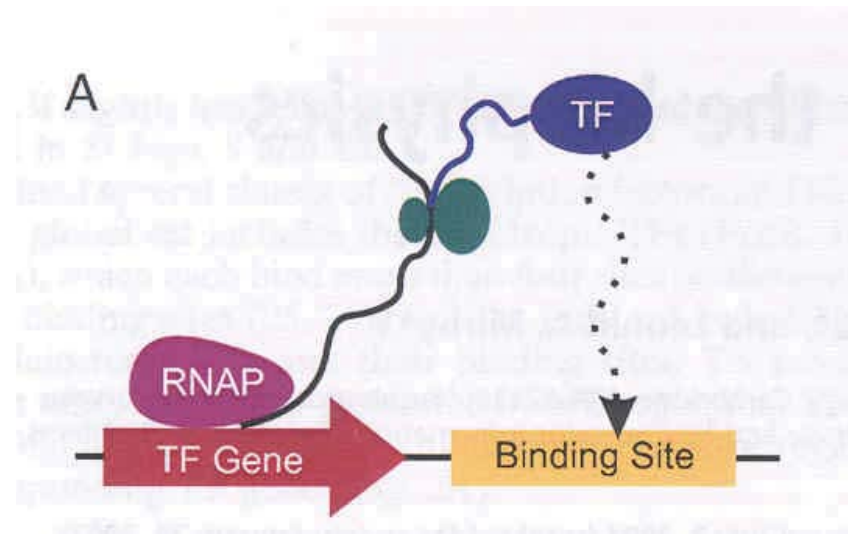
# Problems:

G. Kolesov et al., *PNAS USA*, **104**, 13948 (2007)

Search time for **transcription factors (TF)** proteins from theoretical estimates – **15-500 minutes (!!!)**, while from experiments **~1min**

## Colocalization mechanism:

Genes for TF and for their targets are close to enable fast search



However, 1) it does not work for eukaryotes; 2) does not work for proteins with many targets; 3) does not explain *invitro* experiments

# Our Goal:

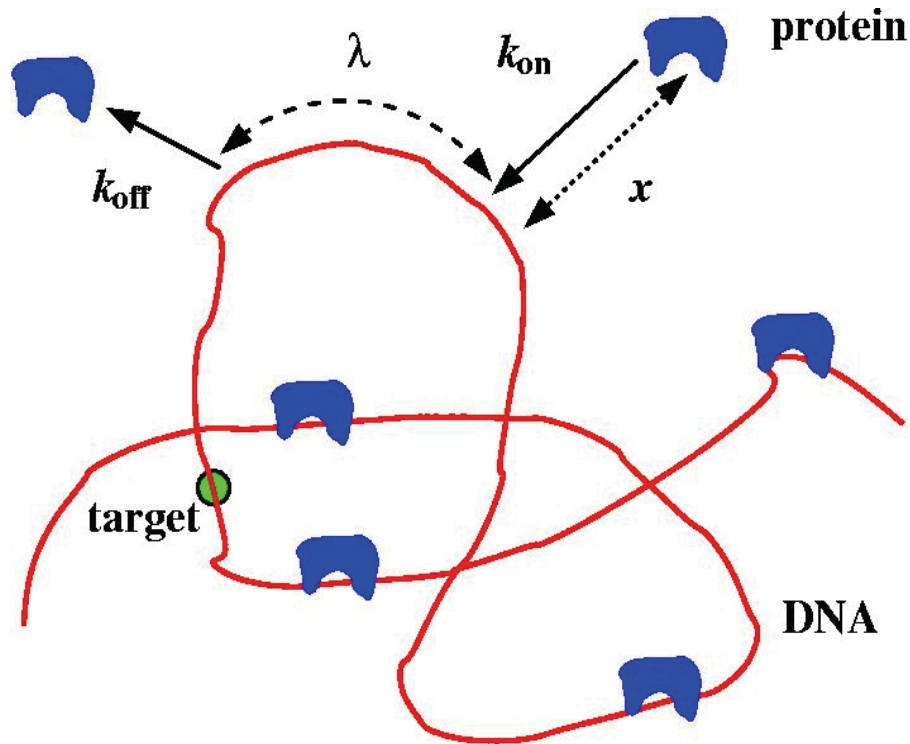
**To develop a simple model of protein search and recognition for targets on DNA consistent with experimental observations and basic laws of Chemistry and Physics**

Protein-DNA interactions – 2 steps:

- 1) Finding the target sequence
- 2) Recognizing the target

Successful theoretical picture must account for both stages of protein-DNA interactions

# Our Theoretical Approach

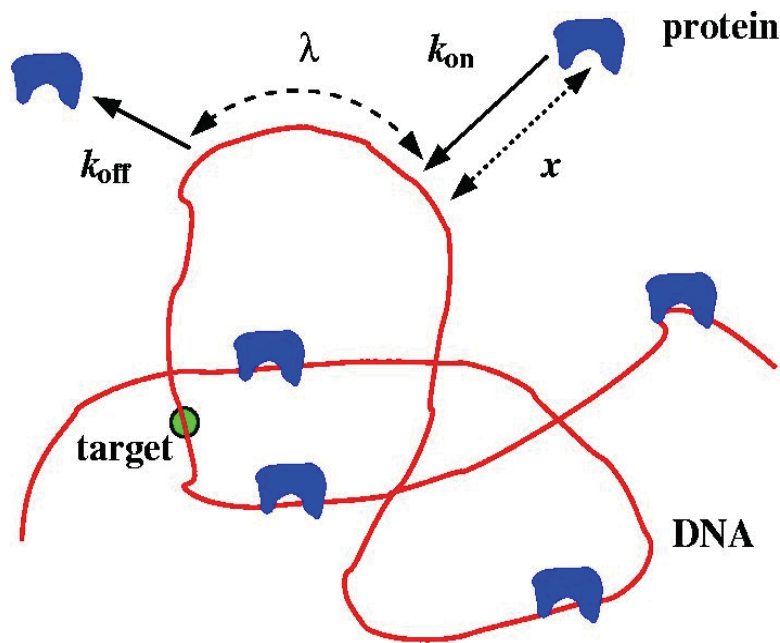


$L \sim 1 \mu\text{m}$  – length of DNA

$a \sim 1 \text{ nm}$  – target size

- 1) Reaching the target on DNA is viewed as a sequence of searching events (cycles)
- 2) Each protein on average binds/unbinds several times before finding the target
- 3) Each cycle consists of 3D and 1D tracks
- 4) Correlations
- 5) No assumption of equilibrium

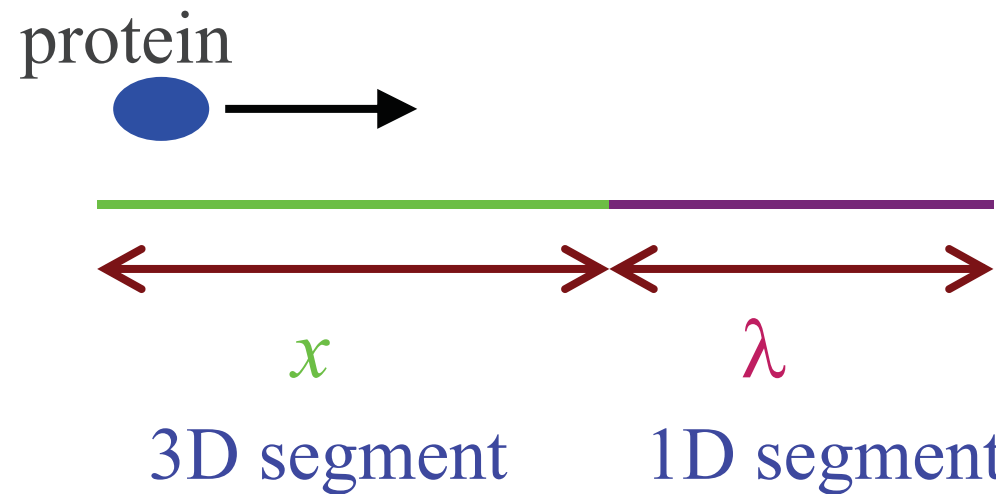
# Our Theoretical Approach



Mean first-passage time  
for one searching cycle

$$\tau_c = \int_0^{x+\lambda} \frac{\exp[\beta G(z)]}{D(z)} dz \int_0^z \exp[-\beta G(z')] dz'$$

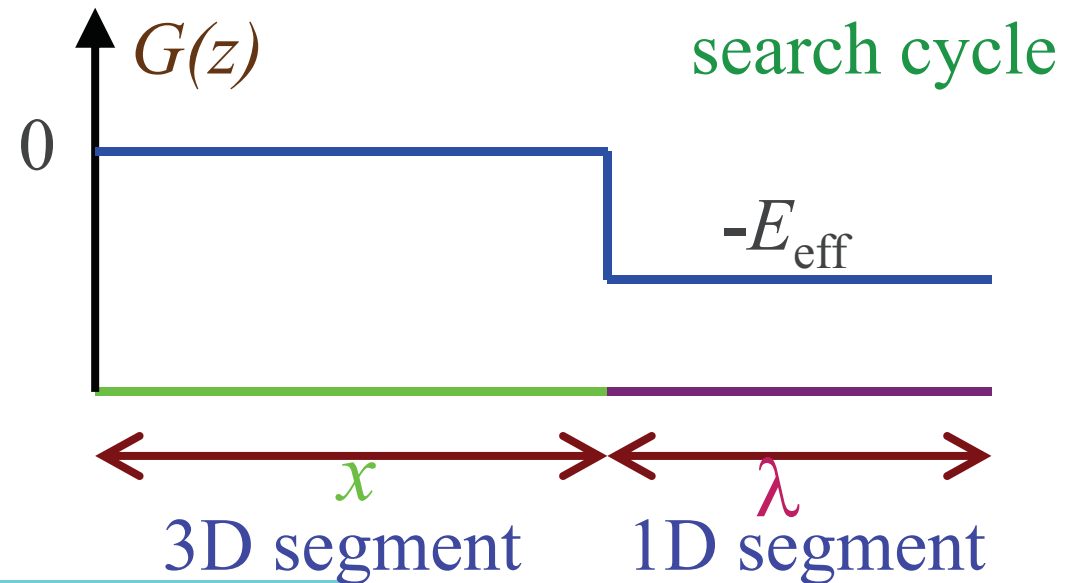
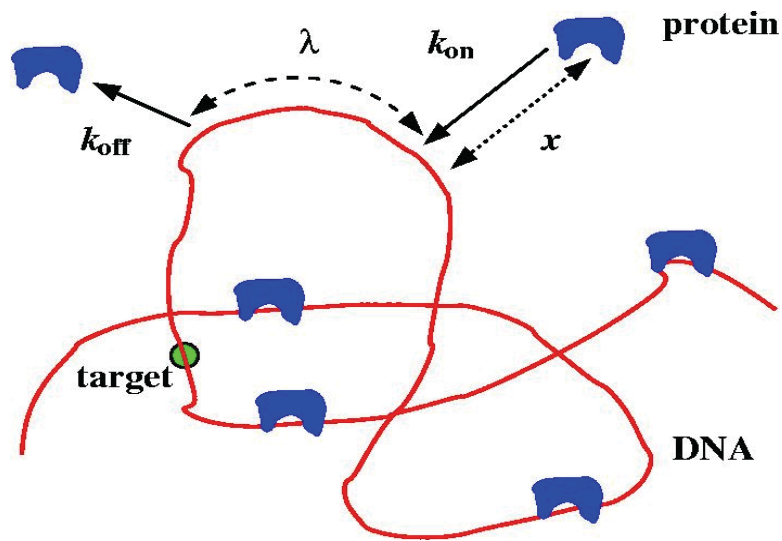
One search cycle – effective  
overall 1D motion



position-dependent  
diffusion constant

$$D(z) = \begin{cases} D_3, & 0 < z < x \\ D_1, & x < z < x + \lambda \end{cases}$$

# Our Theoretical Approach



$$\tau_c = \int_0^{x+\lambda} \frac{\exp[\beta G(z)]}{D(z)} dz \int_0^z \exp[-\beta G(z')] dz'$$

$E_{\text{ads}}$  – non-specific  
binding energy

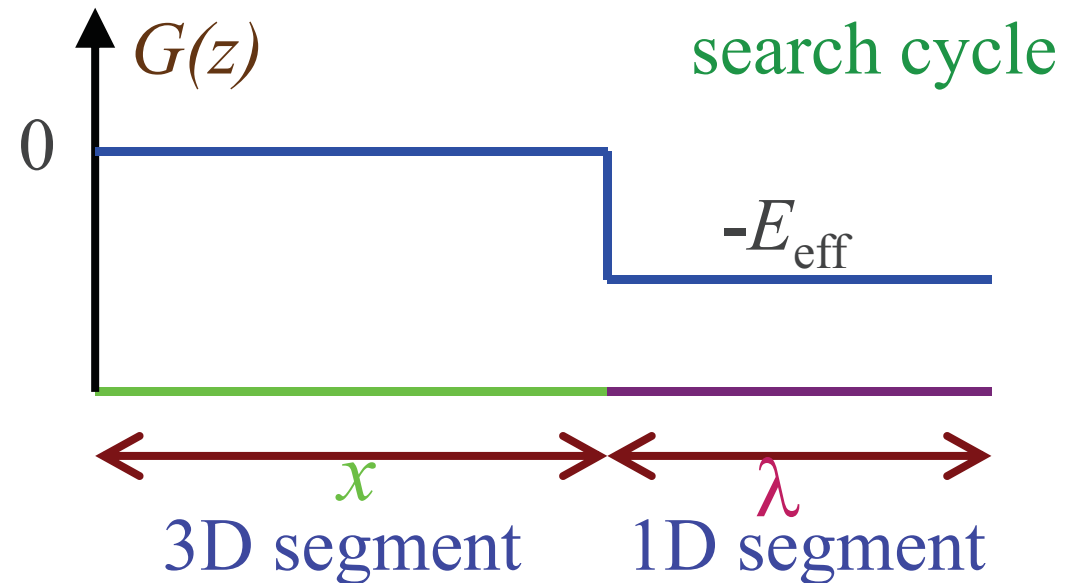
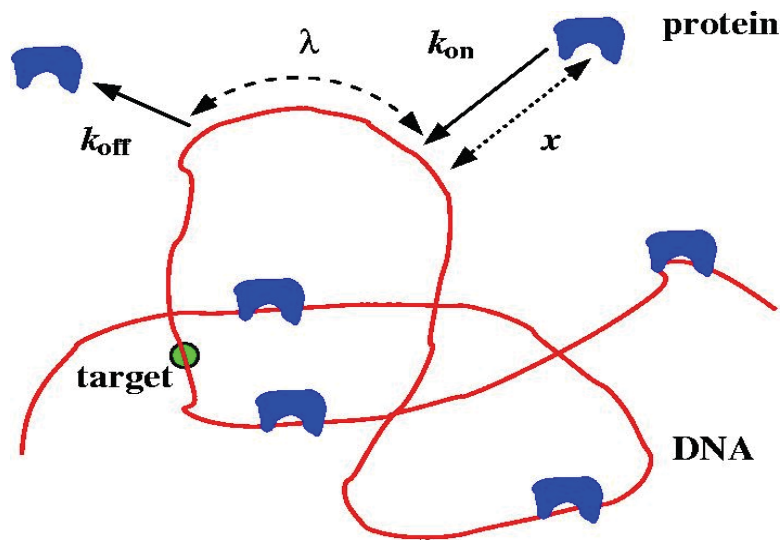
$y$ -equilibrium constant for  
binding/unbinding

$c_p$ -concentration of proteins in the  
bulk;  $c_{\text{ads}}$  – adsorbed to DNA

$$y = \frac{k_{\text{on}}}{k_{\text{off}}} = \exp\left(\frac{E_{\text{ads}}}{k_B T}\right)$$

$$y_{\text{eff}} = \frac{k_{\text{on}} c_p}{k_{\text{off}} c_{\text{ads}}} = \exp\left(\frac{E_{\text{eff}}}{k_B T}\right)$$

# Our Theoretical Approach



Mean first-passage time for one searching cycle

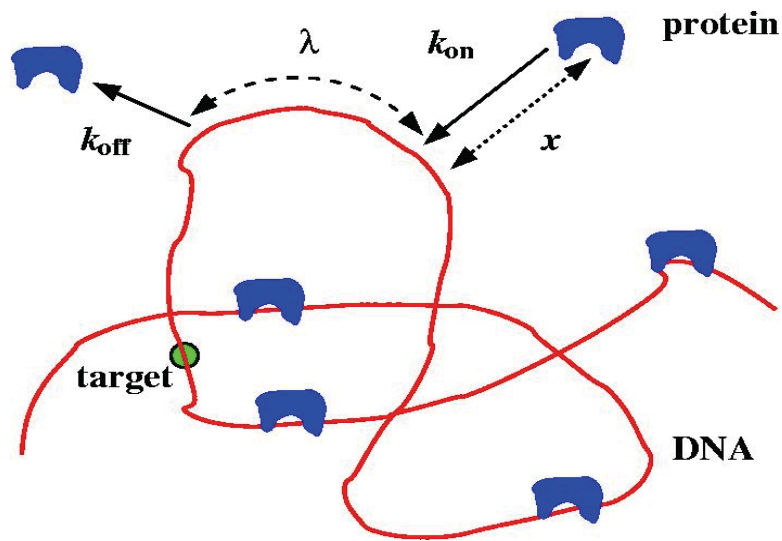
$$\tau_c = \frac{x^2}{2D_3} + \frac{\lambda^2}{2D_1} + \frac{x\lambda}{D_1 y_{eff}}$$

time spent by  
the protein only  
on 3D segment

time spent by  
the protein only  
on 1D segment

correlation  
term – reflects  
forward-  
backward  
trajectories

# Our Theoretical Approach



Consider 1 DNA molecule

$n_{\text{ads}}$  - number of adsorbed proteins per DNA

$\lambda n_{\text{ads}}$  - length of DNA scanned during one cycle

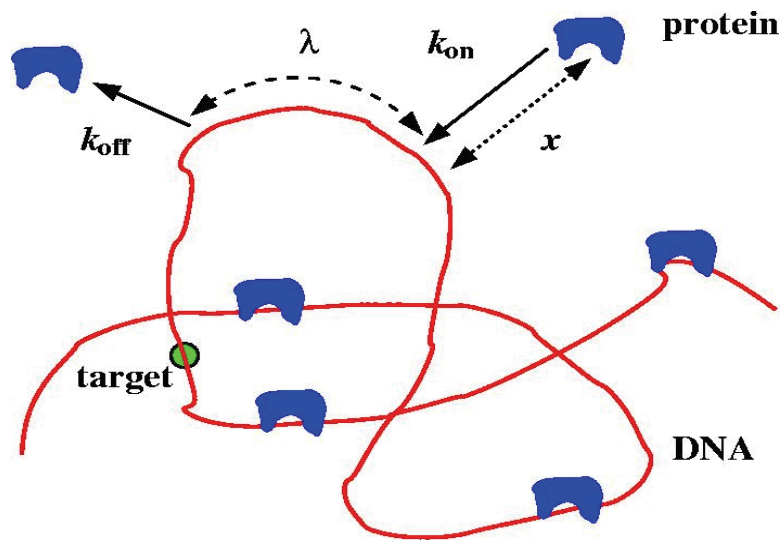
Total search time is a sum of one-cycle times

$L / \lambda n_{\text{ads}}$  - average number of cycles before finding the target

We assume low concentration of proteins on DNA and no memory in rebinding

$$\tau = \tau_c \left( \frac{L}{\lambda n_{\text{ads}}} \right)$$

# Our Theoretical Approach



$$\tau = \tau_c \left( \frac{L}{\lambda n_{ads}} \right)$$

$$S_j = p(1 - p)^{j-1}$$

Average number of search cycles



$$\langle j \rangle = \sum_{j=1}^{\infty} j S_j = \frac{1}{p} = \frac{L}{\lambda n_{ads}}$$

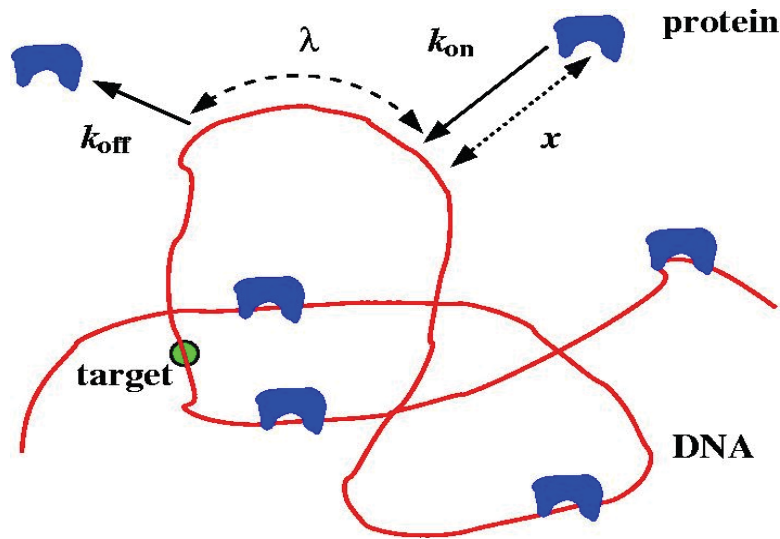
Why average number of search cycles is  $L / \lambda n_{ads}$  ?

$p = \lambda n_{ads} / L$  - probability to find a target after binding DNA

$\lambda n_{ads}$  - average distance scanned by all proteins on DNA in one cycle

$S_j$  - probability to find the target after  $j$ -th cycle

# Our Theoretical Approach



$$V = Lr^2$$

volume per DNA,

$r$ -effective DNA radius

$a$ -target size

$$c_p = \frac{n_p}{V}, \quad c_{ads} = \frac{n_{ads}}{V}$$

Total search time should be compared with Smoluchowski 3D search time

$$\tau_s = \frac{1}{2D_3ac_p} = \frac{Lr^2}{2D_3an_p}$$

**relative search time:**

$d=D_1/D_3$ -ratio of diffusion constants

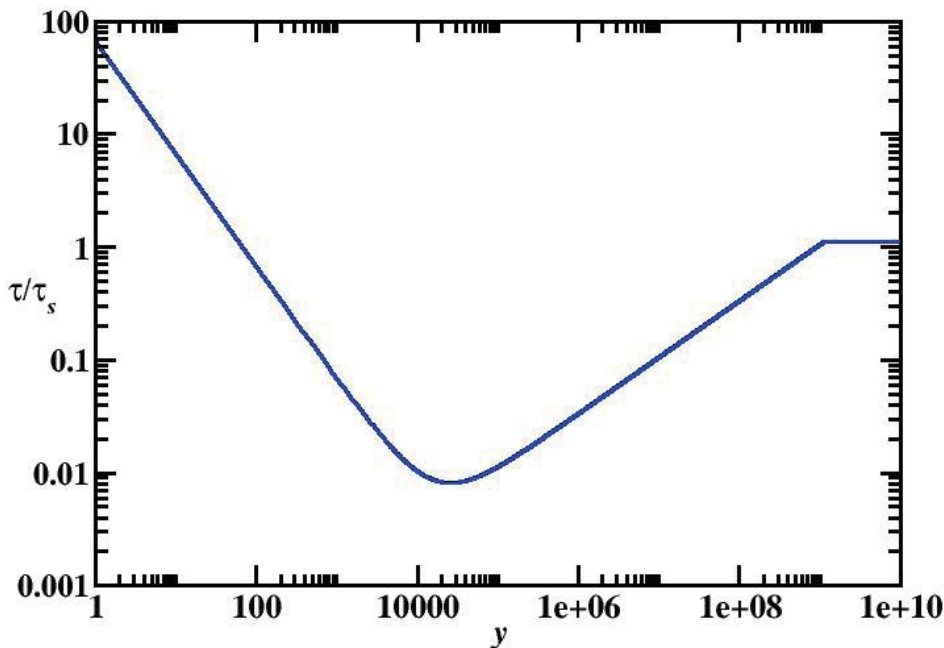
$y$ -equilibrium constant

$$\frac{\tau}{\tau_s} = \frac{a}{r} \left( \frac{1}{\sqrt{n_{ads} y d}} + \frac{n_p \sqrt{y}}{n_{ads}^{3/2} \sqrt{d}} + \frac{2}{\sqrt{n_p y d}} \right)$$

# Results

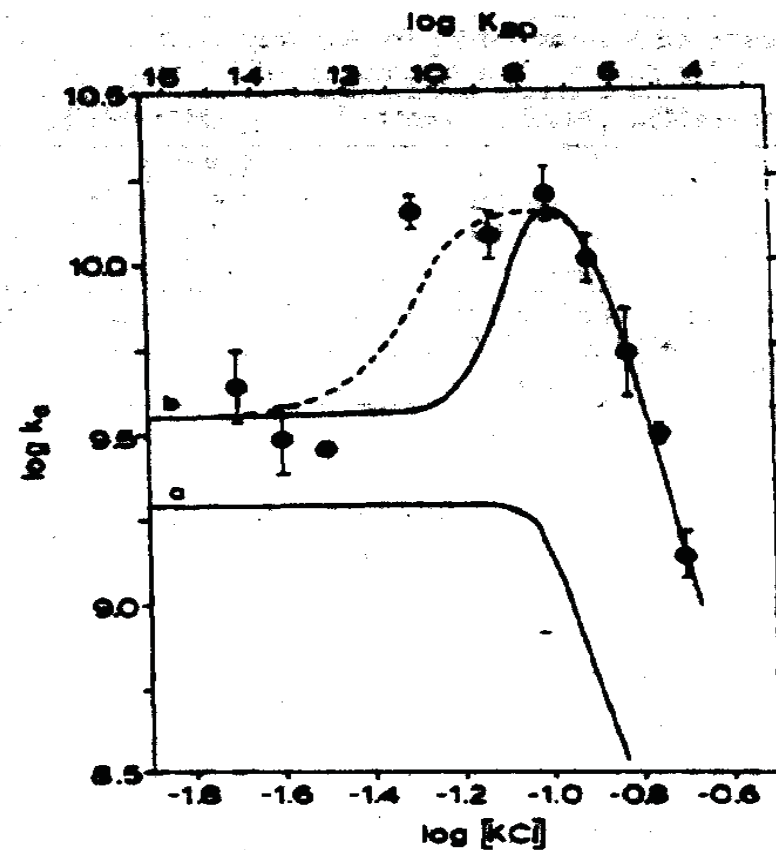
relative search time as a function  
of the adsorption strength

$$a=1\text{nm}, r=30\text{nm}, n_{\text{ads}}=1000, \\ n_p=1, d=0.001$$



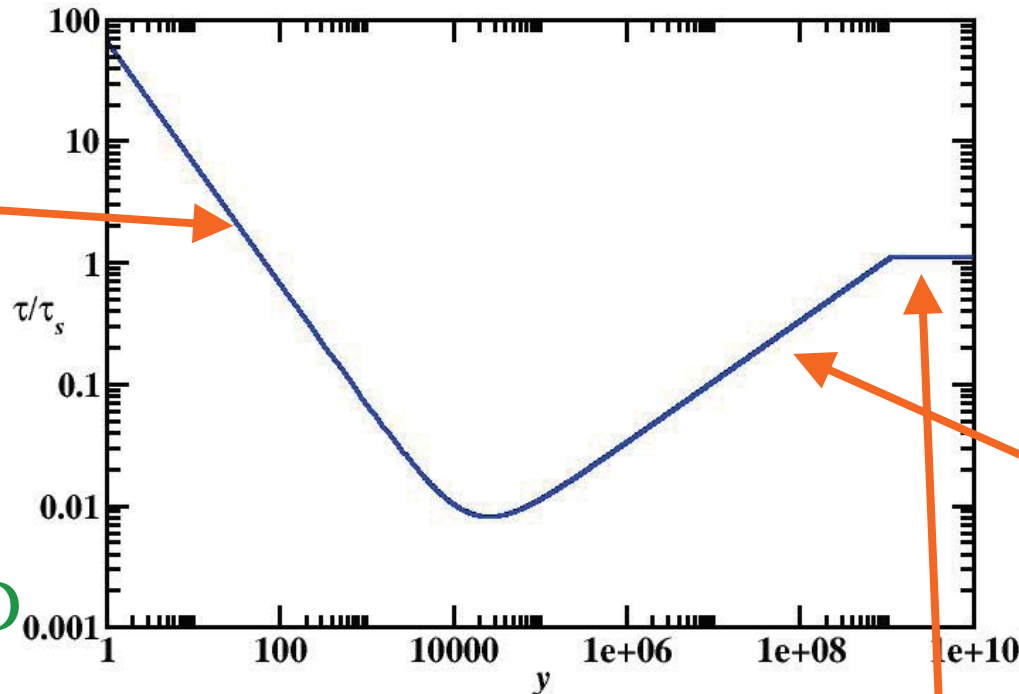
Interactions between  
proteins and DNA depend  
on ionic strength

*Biochemistry*, **20**, 6961 (1981)



# Results

at  $y \ll 1$  –  
strong  
repulsion  
between the  
protein and  
DNA, the  
protein is  
mainly in 3D  
motion, not  
enough time  
to scan DNA



at  $y \gg 1$  –  
strong  
attraction  
between the  
protein and  
DNA, the  
protein is  
mainly in 1D  
motion,  
which is very  
slow

$$y = \frac{k_{on}}{k_{off}} = \exp\left(\frac{E_{ads}}{k_B T}\right)$$

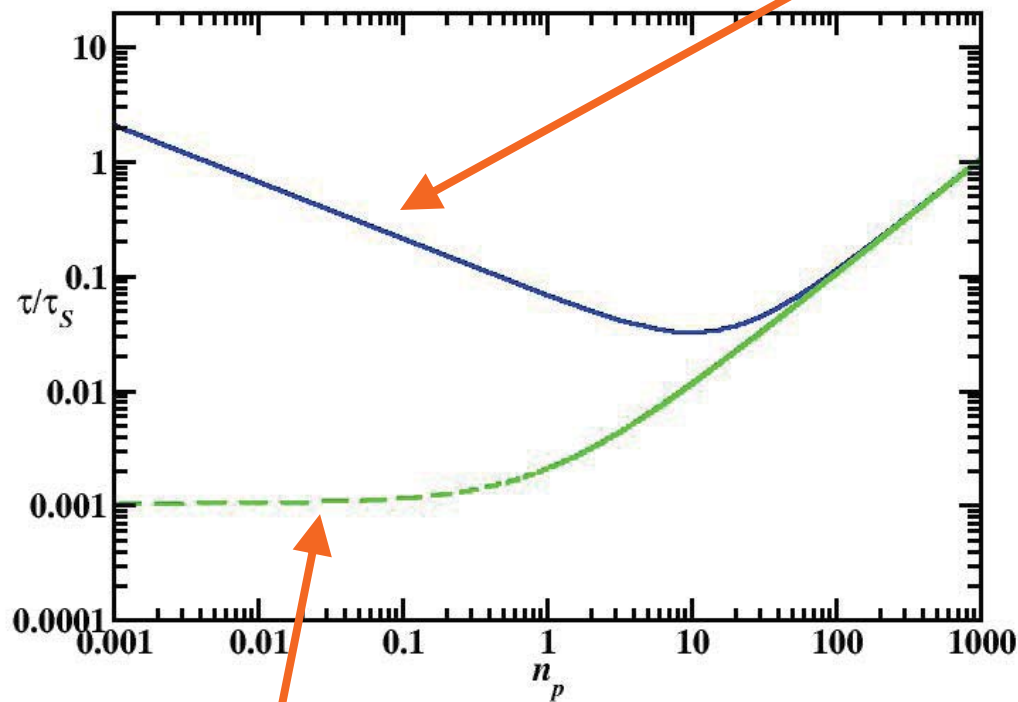
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sliding length is equal to  $L$

# Results

Relative search time as a function of protein concentration

Our theory with correlation term



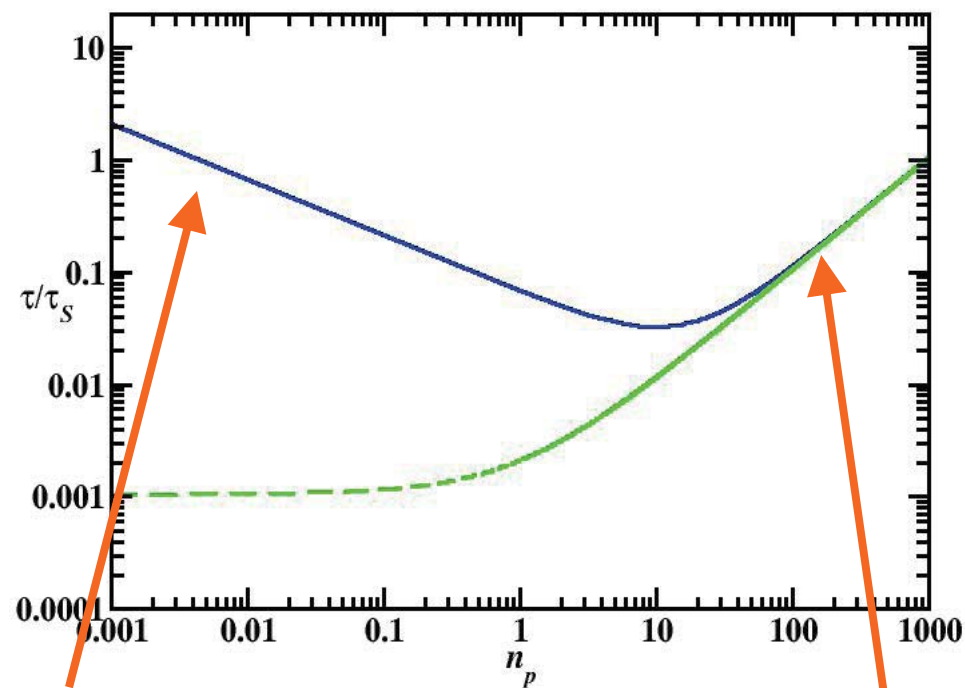
$$\frac{\tau}{\tau_S} = \frac{a}{r} \left( \frac{1}{\sqrt{n_{ads} y d}} + \frac{n_p \sqrt{y}}{n_{ads}^{3/2} \sqrt{d}} + \frac{2}{\sqrt{n_p y d}} \right)$$

Correlation term is critically important!

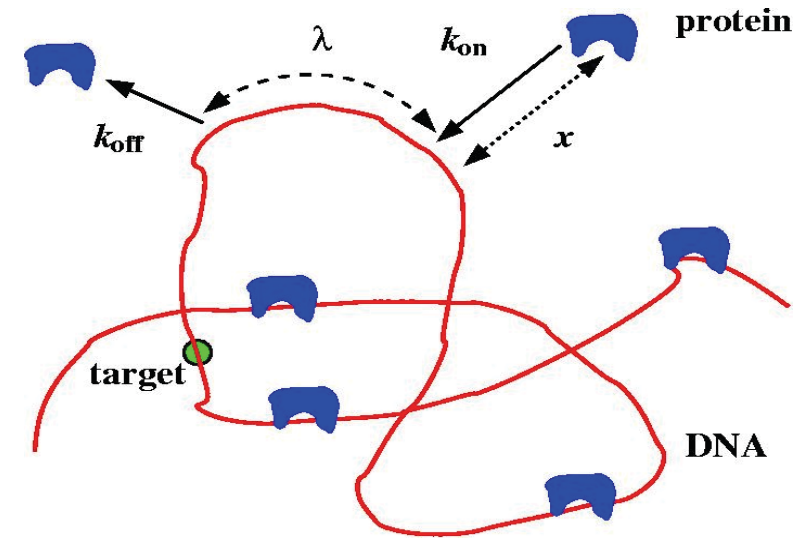
Old theories without correlation term

# Results

Relative search time as a function of solution protein concentration:



Reaching the DNA is a rate-limiting step, protein mostly binds/unbinds

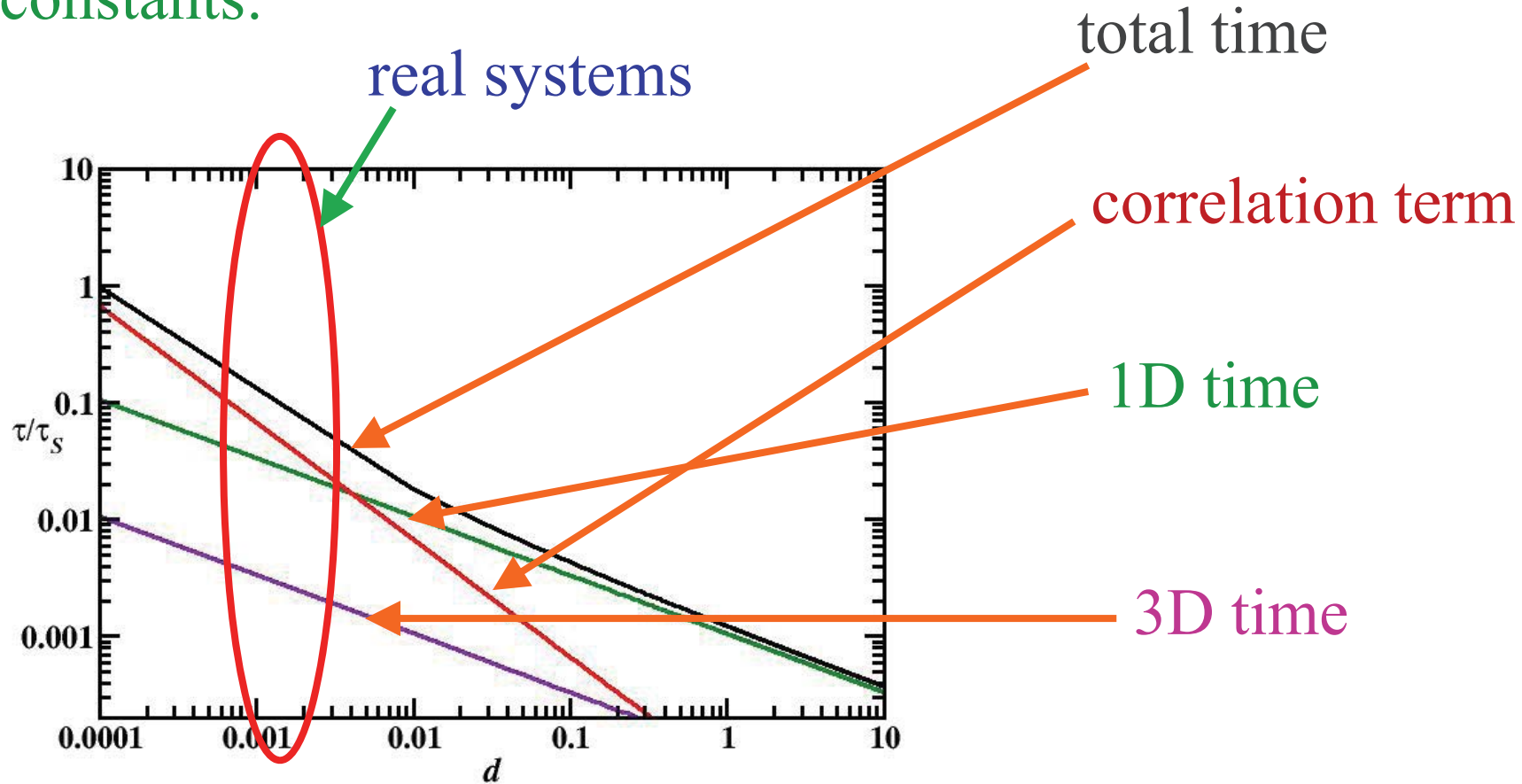


$$\frac{\tau}{\tau_S} = \frac{a}{r} \left( \frac{1}{\sqrt{n_{ads} y d}} + \frac{n_p \sqrt{y}}{n_{ads}^{3/2} \sqrt{d}} + \frac{2}{\sqrt{n_p y d}} \right)$$

concentration of proteins becomes so large that it is faster to reach the target only by 3D

# Results

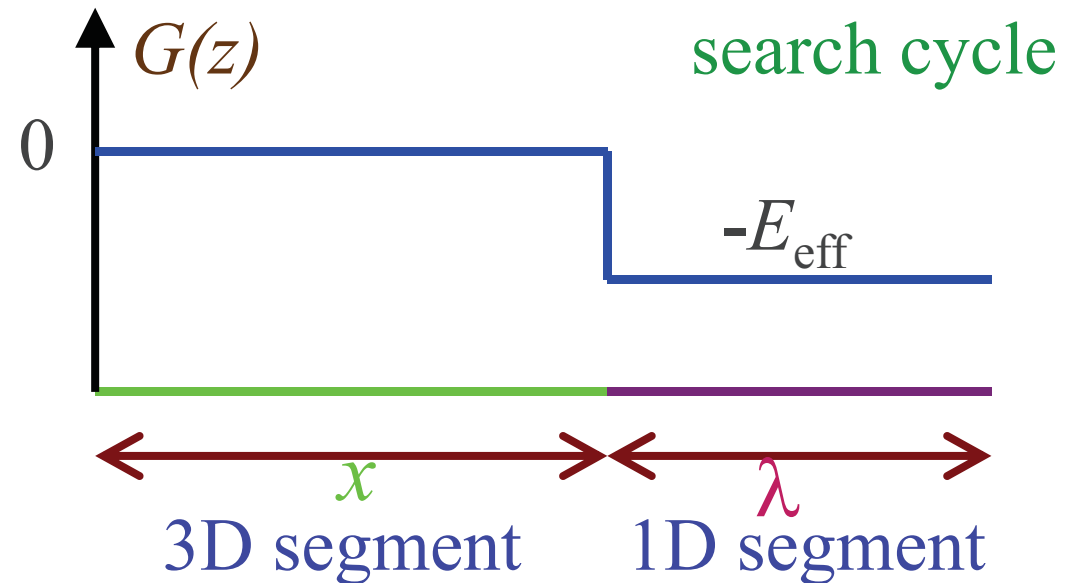
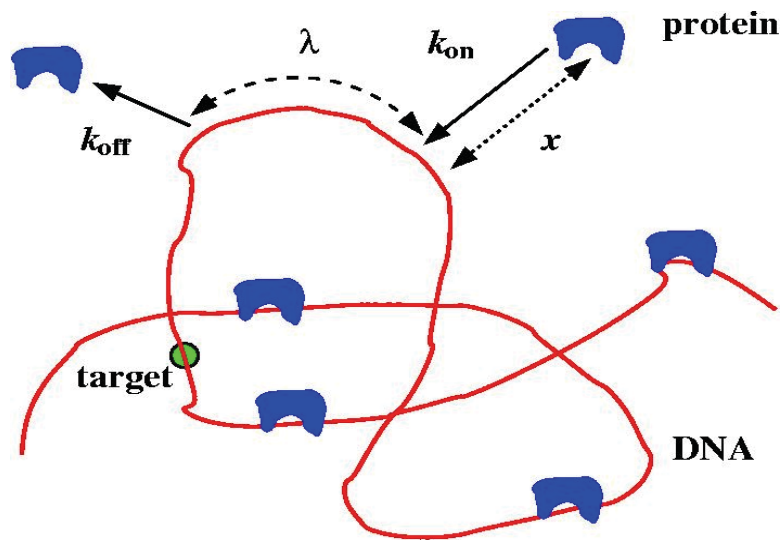
Relative search time as a function of the ratio of diffusion constants:



$$\frac{\tau}{\tau_s} = \frac{a}{r} \left( \frac{1}{\sqrt{n_{ads}} y d} + \frac{n_p \sqrt{y}}{n_{ads}^{3/2} \sqrt{d}} + \frac{2}{\sqrt{n_p} y d} \right)$$

$d = D_1/D_3$  - ratio of diffusion constants

# Results



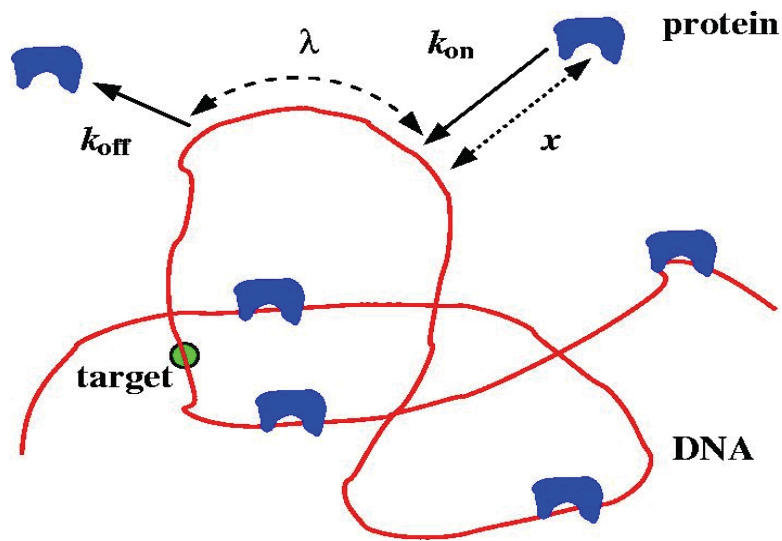
Our theory is non-equilibrium, equilibrium is a special case when  $E_{\text{eff}}=0$

For realistic values  $a=1\text{nm}$ ,  
 $r=30\text{nm}$ ,  $y=10$ ,  $n_p=1$ ,  $d=0.001$   
**No acceleration in our model**

$$\frac{\tau}{\tau_s} = \frac{a}{r} \frac{2}{y\sqrt{n_p}} \frac{\sqrt{d} + 1}{d}$$

facilitated diffusion – non-equilibrium process

# Our Theoretical Approach



## Our view of facilitated diffusion mechanism:

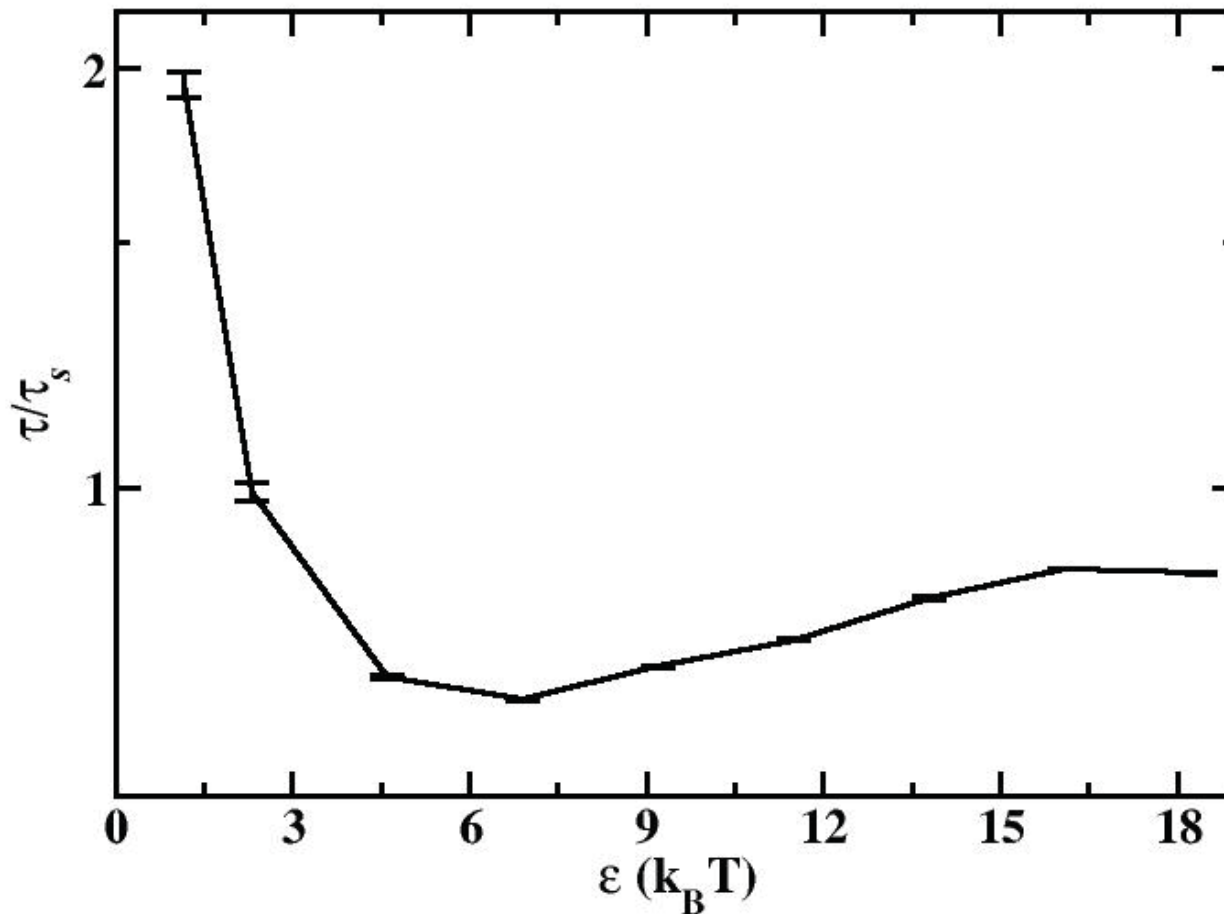
- 1) Non-equilibrium process
- 2) Fast 3D and slow 1D motions
- 3) Correlations are important
- 4) Acceleration due to increased local concentration of proteins because of non-specific binding

## Non-specific binding:

- 1) Slows down 1D diffusion
- 2) Increases concentration of proteins on DNA

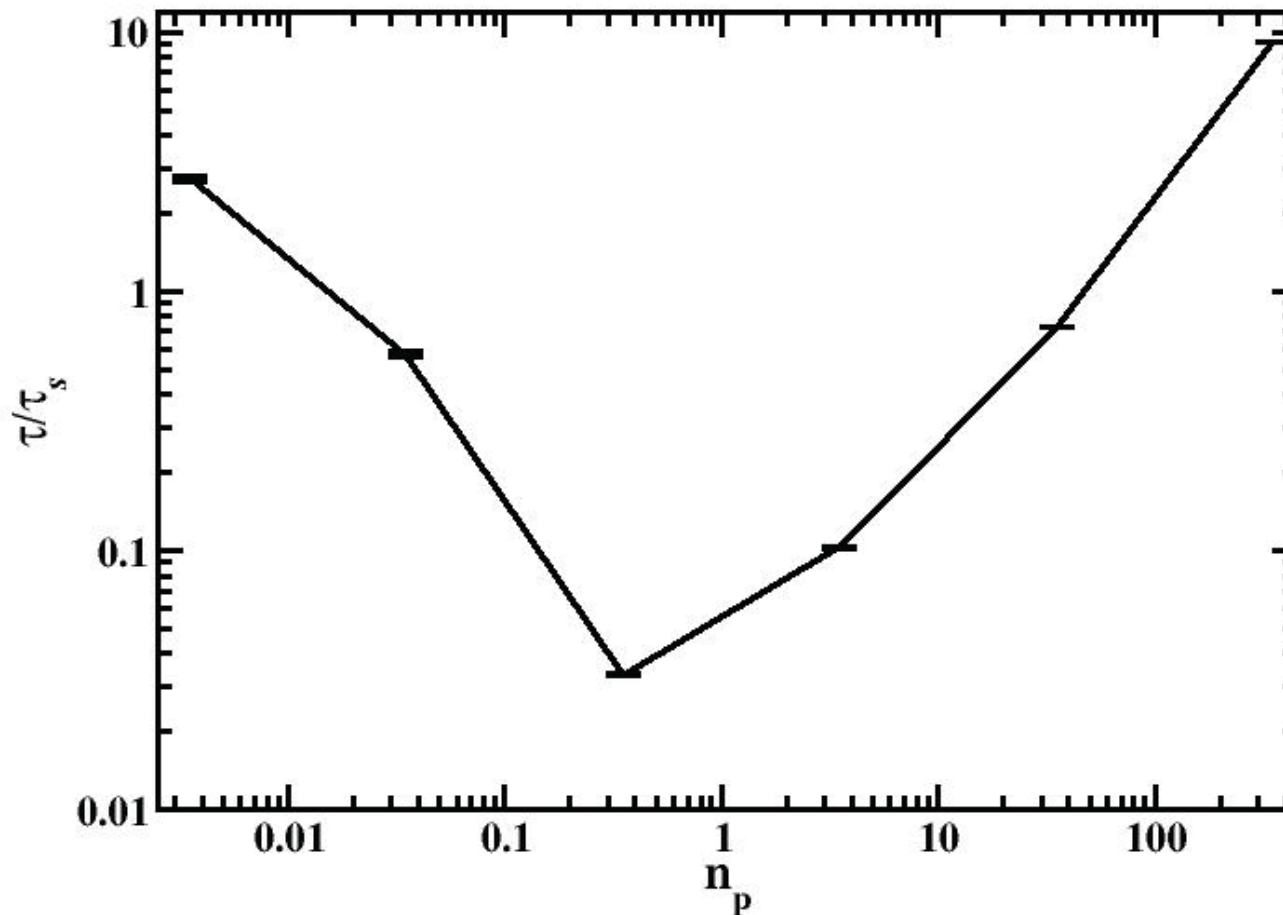
# Monte Carlo Simulations

Relative search time as a function of non-specific interaction with DNA



# Monte Carlo Simulations

Relative search time as a function of free proteins in solution



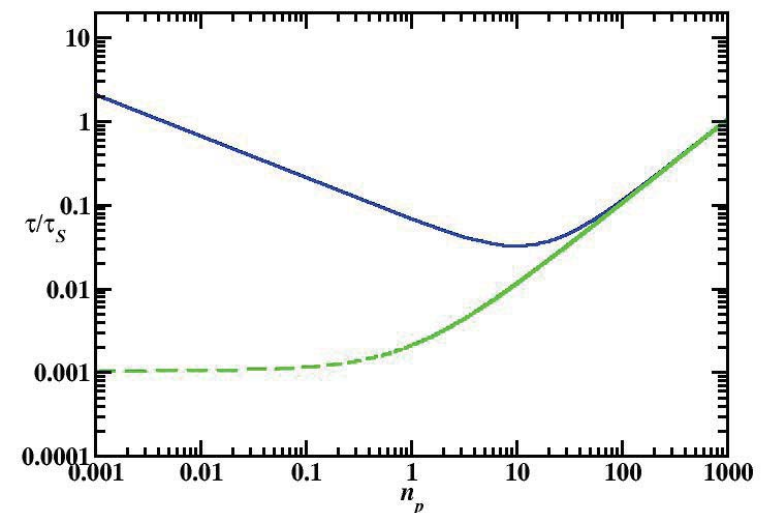
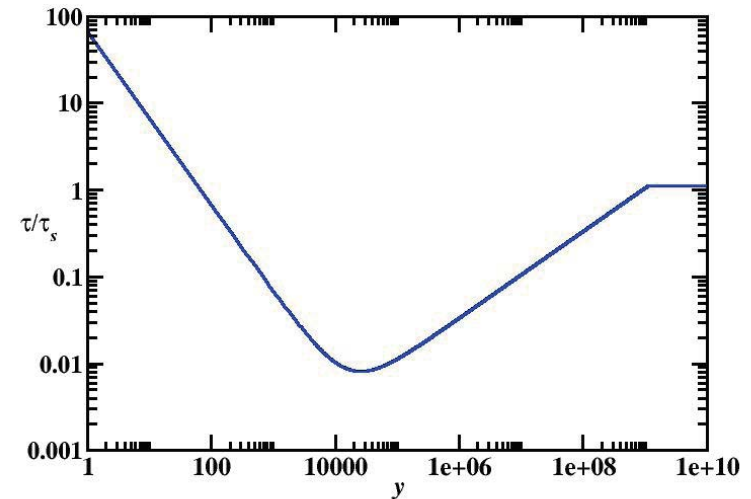
# Our Predictions:

1) **Facilitated diffusion mechanism** works for intermediate range of adsorption energies. It can be controlled by changing ionic strength

Agrees with experiments and computer simulations

2) **Facilitated diffusion mechanism** works for intermediate concentrations of proteins in the solution

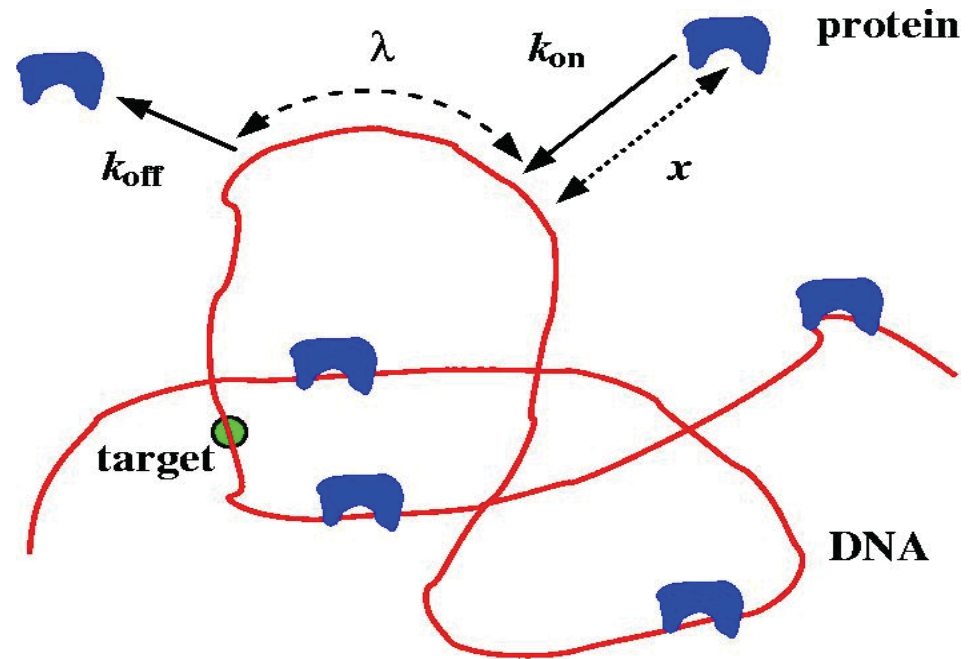
3)  $\tau(1D)/\tau(3D) \gg 1$  – agrees with



# Open Questions and Problems:

Our theory – approximate,  
improvements are needed

- 1) Correlations between search cycles
- 2) Correlations in binding positions after dissociation
- 3) Protein interactions with DNA that have several targets
- 4) Protein and DNA elasticity
- 5) DNA conformations
- 6) Effect of Temperature
- 7) Sequence dependence



# CONCLUSIONS:

- A theoretical approach of how proteins find and recognize their targets on DNA is developed
- **Facilitated diffusion** is a sequence of fast 3D and slow 1D motions
- Acceleration in facilitated diffusion is achieved due to the increased local concentration of adsorbed proteins (from non-specific binding)
- Our results and predictions consistent with basic laws of Chemistry and Physics
- In agreement with all currently available experimental observations

# Acknowledgements

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Some of the results are published in

*J. Phys. Chem. B* **112**, 4741-4750 (2008)

*J. Phys. Chem. B* **113**, 4242-4247 (2009)