



The Abdus Salam  
International Centre  
for Theoretical Physics

# Thermal Lens Microscopy Combined with Intracavity Spectroscopy

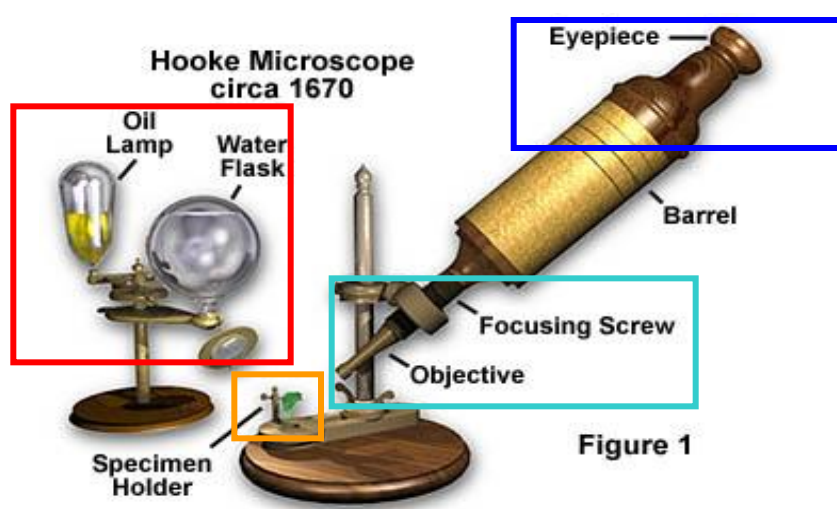
Humberto Cabrera

Preparatory School to the Winter Colleague on Optics: Optical Frequency Combs

- Introduction
- Basic components of the microscope and their functions
- Thermal lens microscopy
- Thermal lens microscopy combined with intracavity spectroscopy
- Typical examples and applications

# Microscope Components

- Ocular
- Objective
- Condenser
- Numerical Aperture
- Optical Filters



# Basic components and their functions

- (1) **Eyepiece (ocular lens)**
- (2) Revolving nose piece (to hold multiple objective lenses)
- (3) **Objective lenses**
- (4) And (5) **Focus knobs**
  - (4) Coarse adjustment
  - (5) Fine adjustment
- (6) **Stage** (to hold the specimen)
- (7) **Light source** (lamp)
- (8) **Condenser lens** and diaphragm
- (9) Mechanical stage (move the specimen on two horizontal axes for positioning the specimen)



# The Concept of Magnification

## Magnification of the Microscope

■  $M_{\text{Microscope}} = M_{\text{Objective}} \times M_{\text{Eyepiece}} \times M_{\text{Intermediate Factor}}$

M = Magnification

■ Example: Objective = 60 x

Eyepiece = 10 x

Intermediate Factor = 1 x

Overall M = 600 x



# The characteristics of objectives

## 60x Plan Apochromat Objective

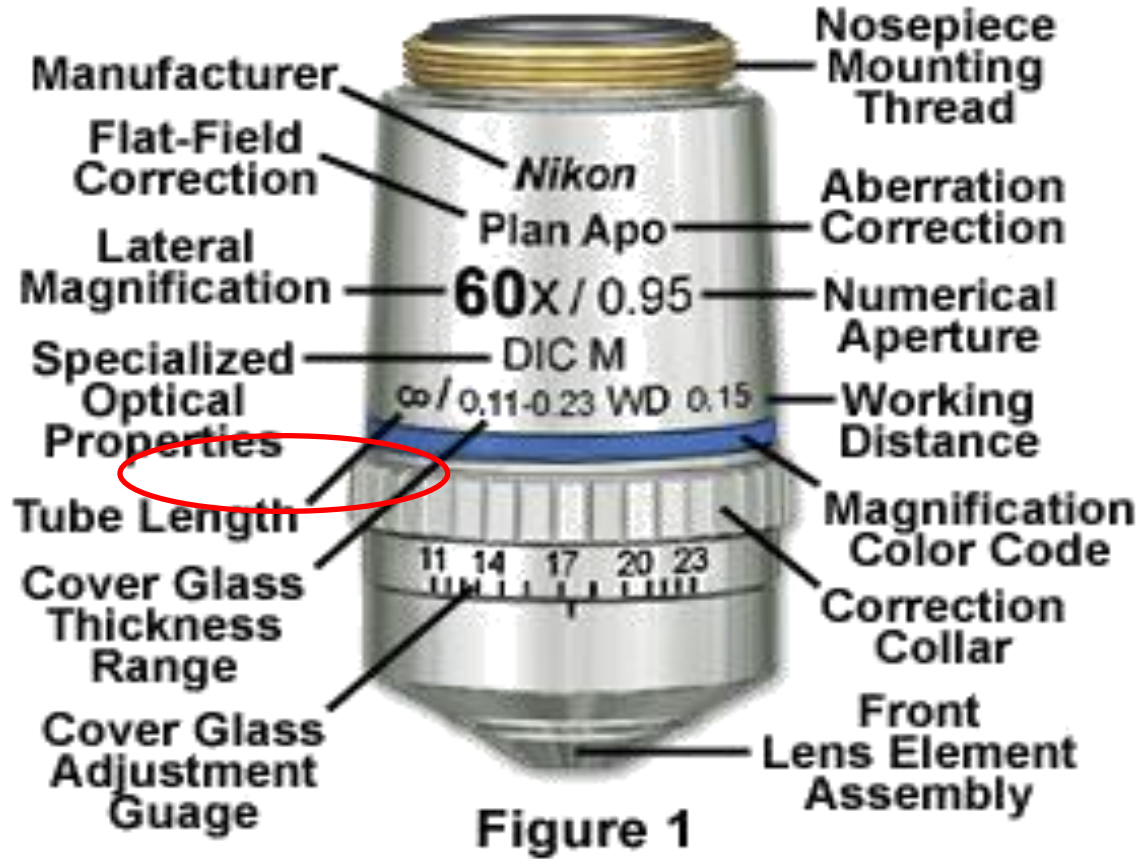
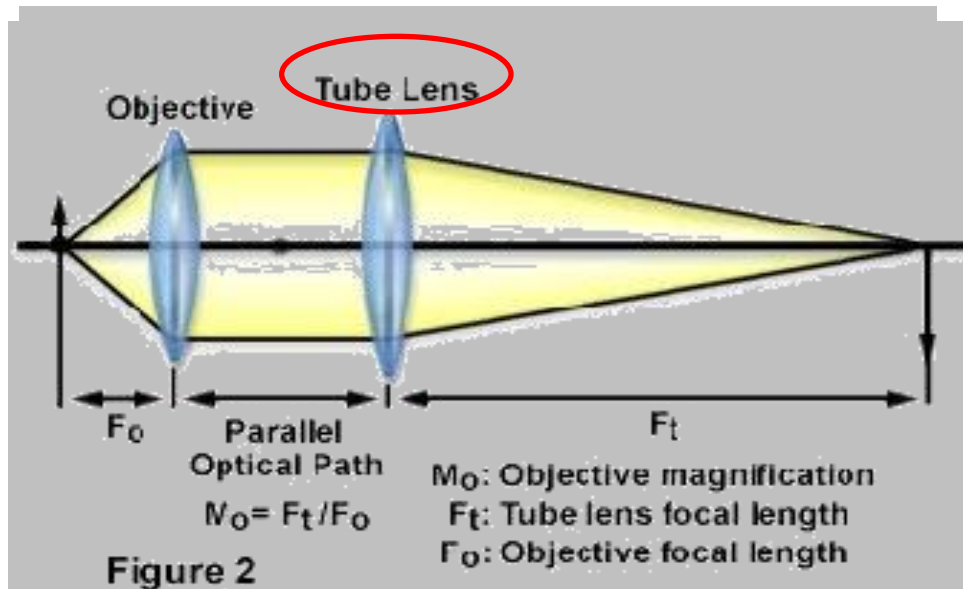
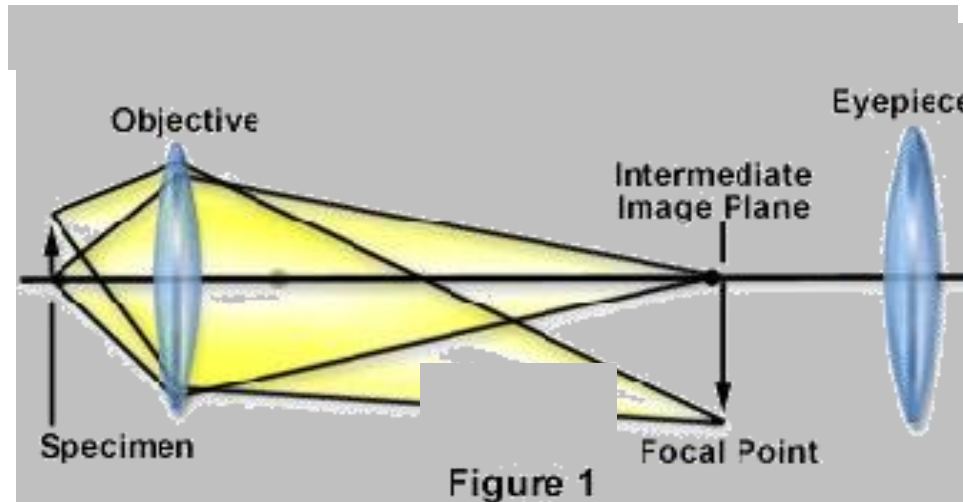


Figure 1



# Objectives configurations



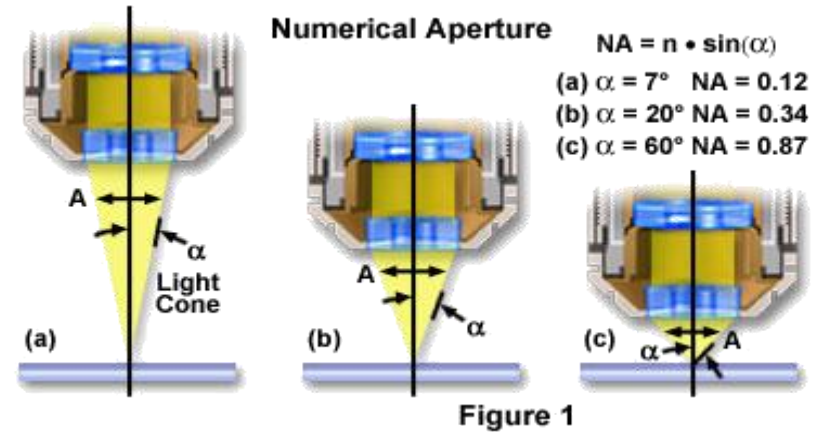
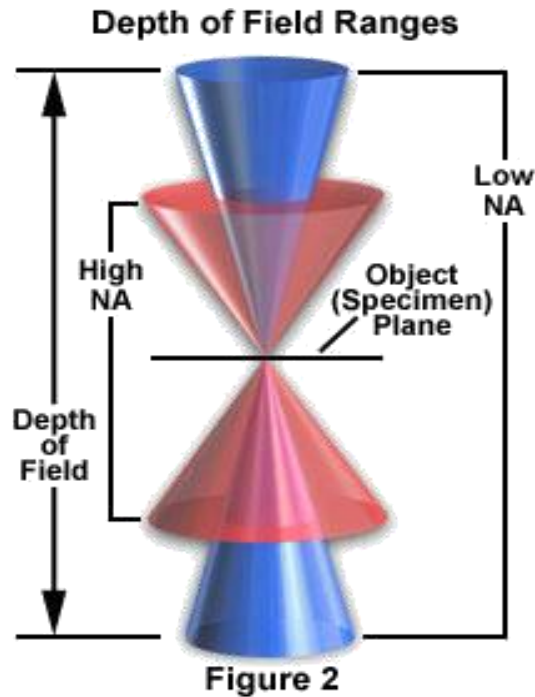


# The characteristics of objectives





# Numerical Aperture (N.A.)



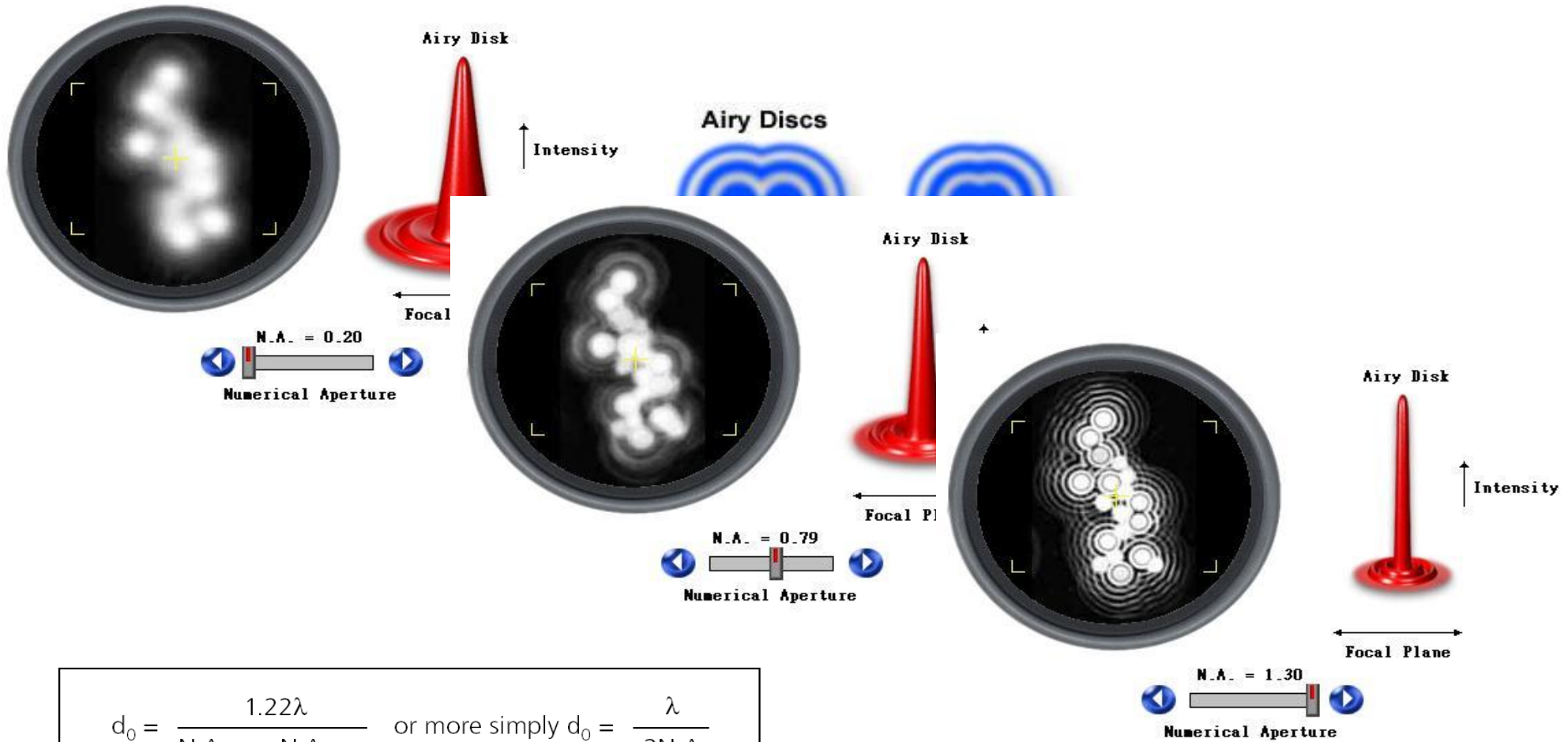
**Numerical Aperture = N.A. =  $n \cdot \sin \alpha$**

$\alpha$  is half the opening angle of the objective.

$n$  is the refractive index of the immersion medium used between the objective and the object.

( $n = 1$  for air;  $n = 1.51$  for oil or glass)

# Resolution



$$d_0 = \frac{1.22\lambda}{N.A._{obj.} + N.A._{Cond}} \quad \text{or more simply } d_0 = \frac{\lambda}{2N.A.}$$

$\lambda$  = wavelength of light, e.g. 550 nm (green)

● **Resolving power, the limit up to which two small objects are still seen separately.**

# Factors Affecting Resolution

- Resolution ( $d_{\min}$ ) improves (smaller  $d_{\min}$ ) if  $\lambda \downarrow$  or  $n \uparrow$  or  $\alpha \uparrow$
- Assuming that  $\sin \alpha = 0.95$  ( $\alpha = 71.8^\circ$ )

Wavelength		Air ( $n=1$ )	Oil ( $n=1.515$ )
Red	650 nm	0.42 $\mu\text{m}$	0.28 $\mu\text{m}$
Yellow	600 nm	0.39 $\mu\text{m}$	0.25 $\mu\text{m}$
Green	550 nm	0.35 $\mu\text{m}$	0.23 $\mu\text{m}$
Blue	475 nm	0.31 $\mu\text{m}$	0.20 $\mu\text{m}$
Violet	400 nm	0.27 $\mu\text{m}$	0.17 $\mu\text{m}$

Resolution<sub>air</sub>
Resolution<sub>oil</sub>

- (The eye is more sensitive to blue than violet)

# Two sets of conjugate planes in the light microscope

Understanding the reciprocal relationship between the two sets of conjugate planes is crucial for properly understanding:

- Image formation
- Image resolution
- How phase-contrast and DIC work

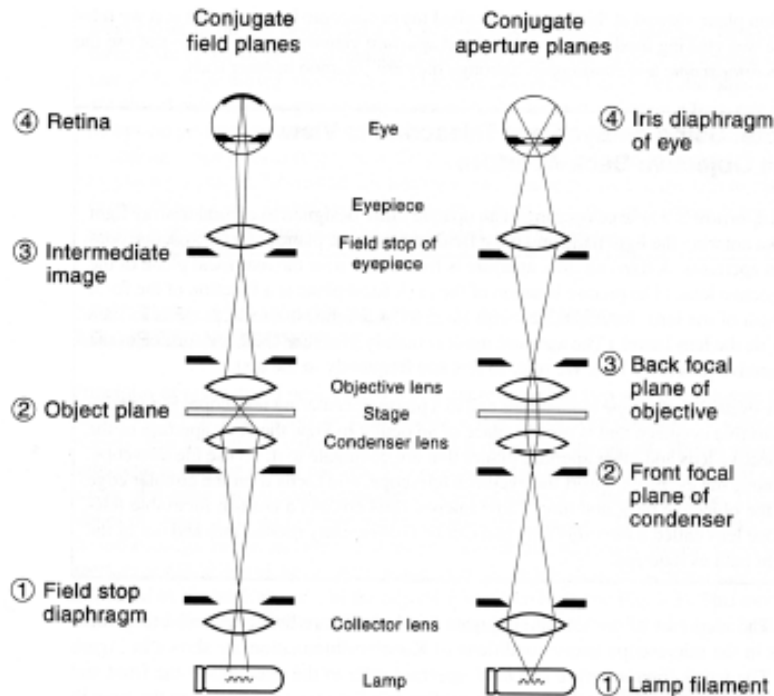


Figure 1-4

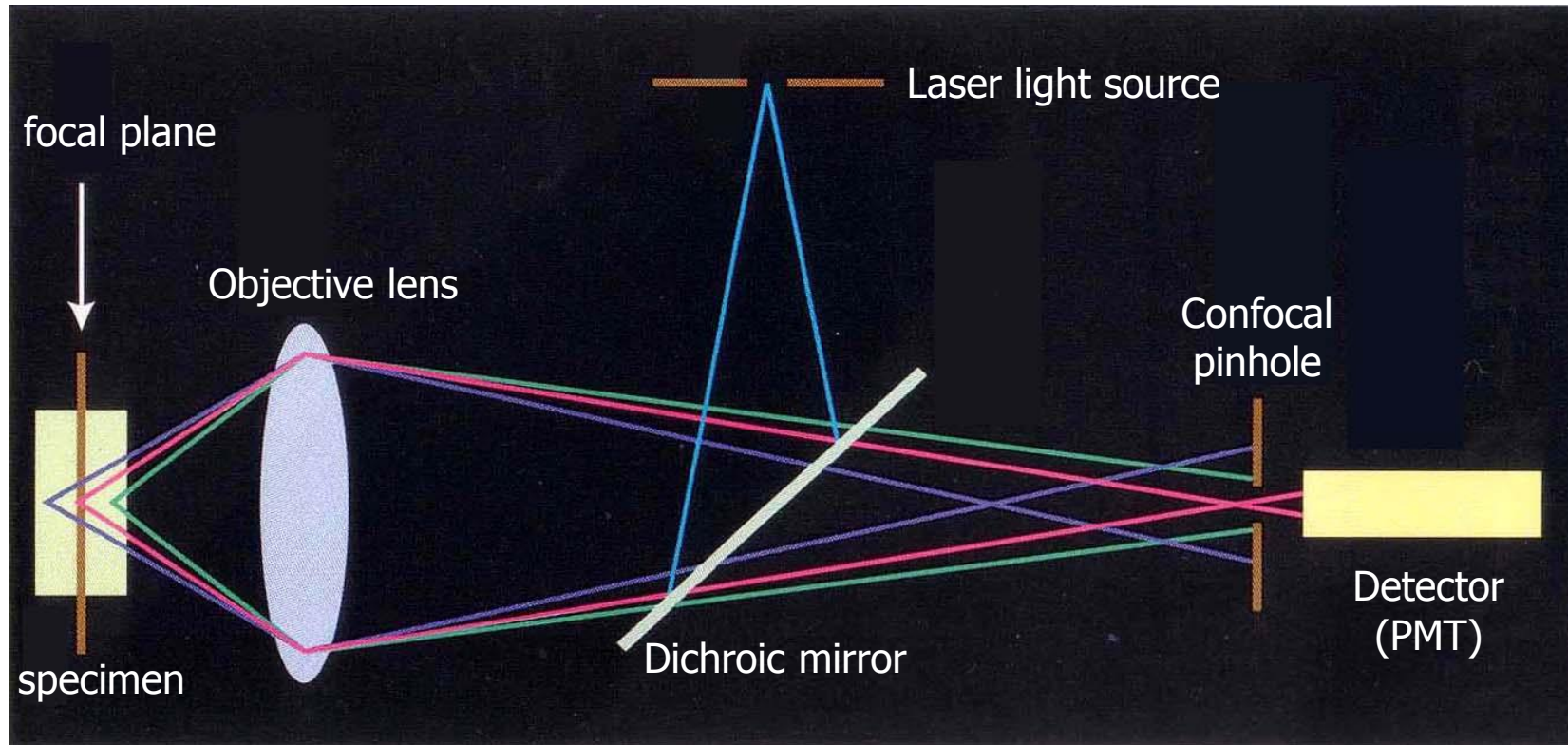
The locations of conjugate focal planes in a light microscope adjusted for Koehler illumination. Note the locations of four conjugate field planes (left) and four conjugate aperture planes (right) indicated by the crossover points of rays in the diagrams. The left-hand diagram shows that the specimen or object plane is conjugate with the real intermediate image plane in the eyepiece, the retina of the eye, and the field stop diaphragm between the lamp and the condenser. The right-hand drawing shows that the lamp filament is conjugate with aperture planes at the front focal plane of the condenser, the back focal plane of the objective, and the pupil of the eye.

Conjugate planes are "parfocal" with each other

When something is in focus in one set of conjugate planes, it is "maximally out-of-focus" in the other set of planes

These two sets are often called "reciprocal" or "transform" planes (with respect to each other)

# Laser Scanning Microscope (Confocal System)



- Light emitted from the focal plane
- Light emitted from the out-of-focus region

# Confocal Aperture

Decreasing the pinhole size rejects more out of focus light, therefore improving contrast and effective z resolution.

Decreasing the pinhole will increase x,y resolution (1.3x wide field)

Decreasing pinhole size decreases the amount of the Airy disk that reaches the detector. This results in less light from each point being collected

Generally, collecting the diameter of 1 Airy disk is considered optimal. This collects about 85% of light from a sub-resolution point.

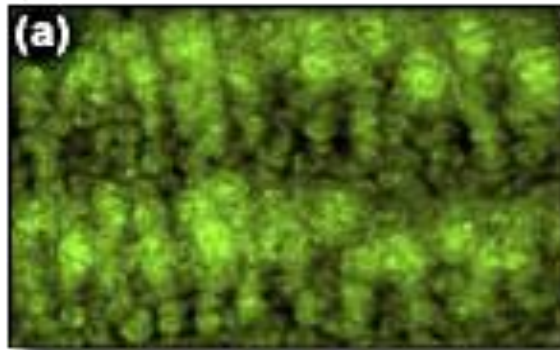
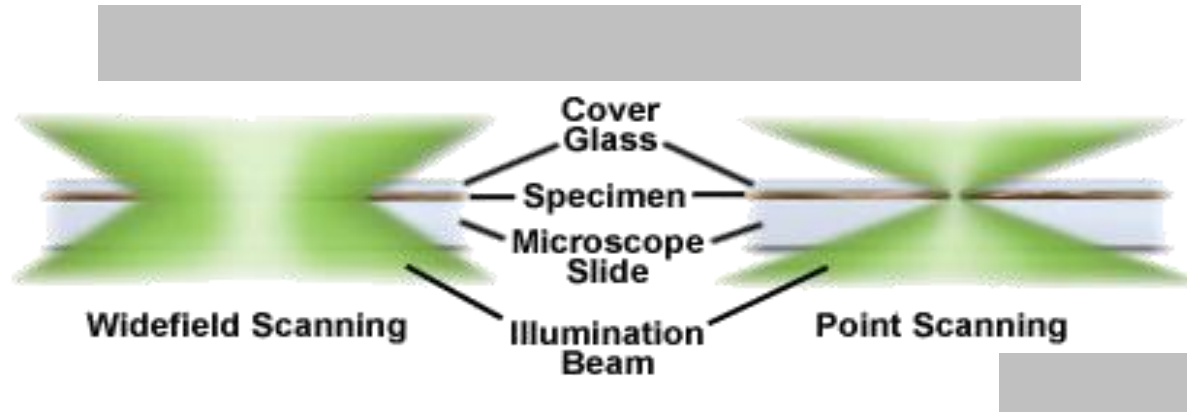
## Limits:

**Open pinhole:** nearly wide field resolution (still some confocality)

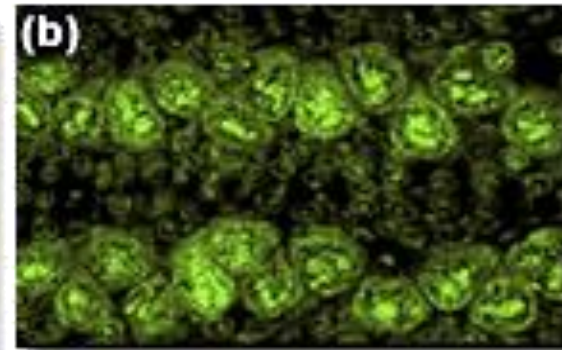
**Closed:** no image



# Wide field versus confocal scanning



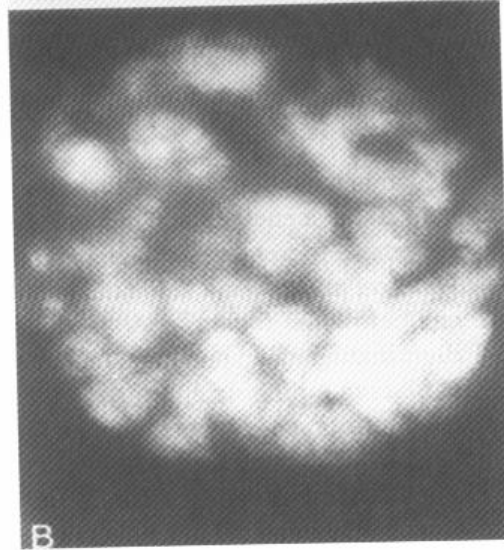
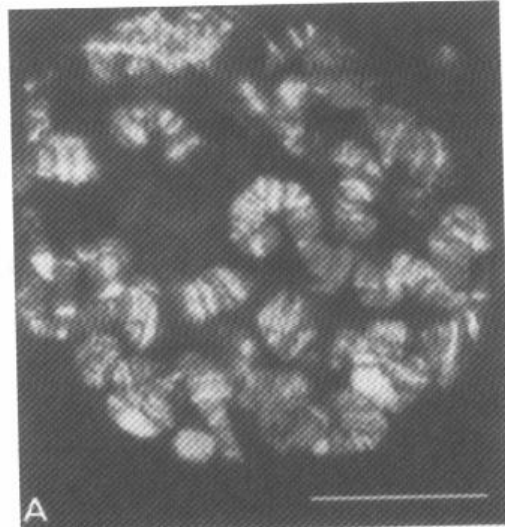
**Wide Field**



**Confocal**



# WF vs C - Fluorescence Imaging



## Confocal

Greatly reduces  
Out of focus blur

## Wide-field

Brighter but  
No sectioning

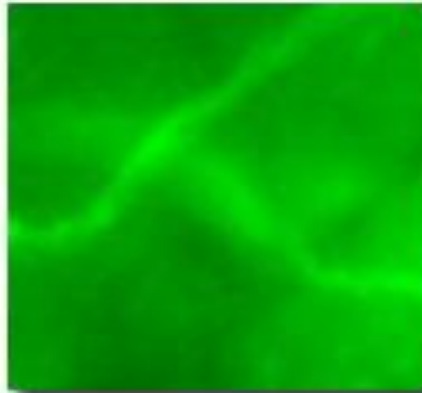
# More examples

## Confocal and Widefield Fluorescence Microscopy

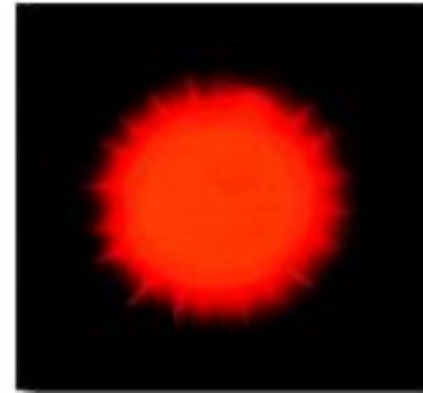
widefield



(a)

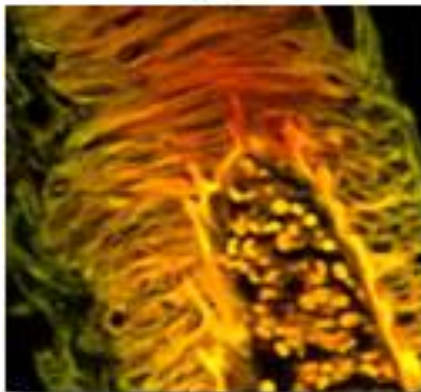


(b)

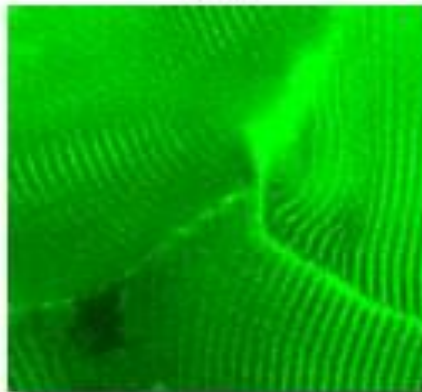


(c)

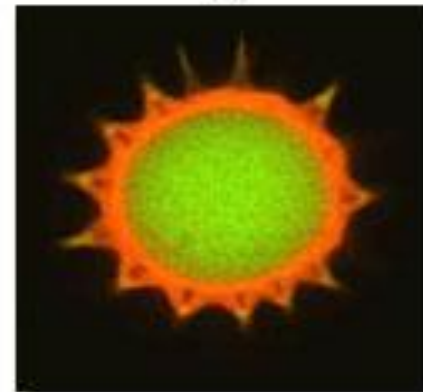
confocal



(d)



(e)



(f)

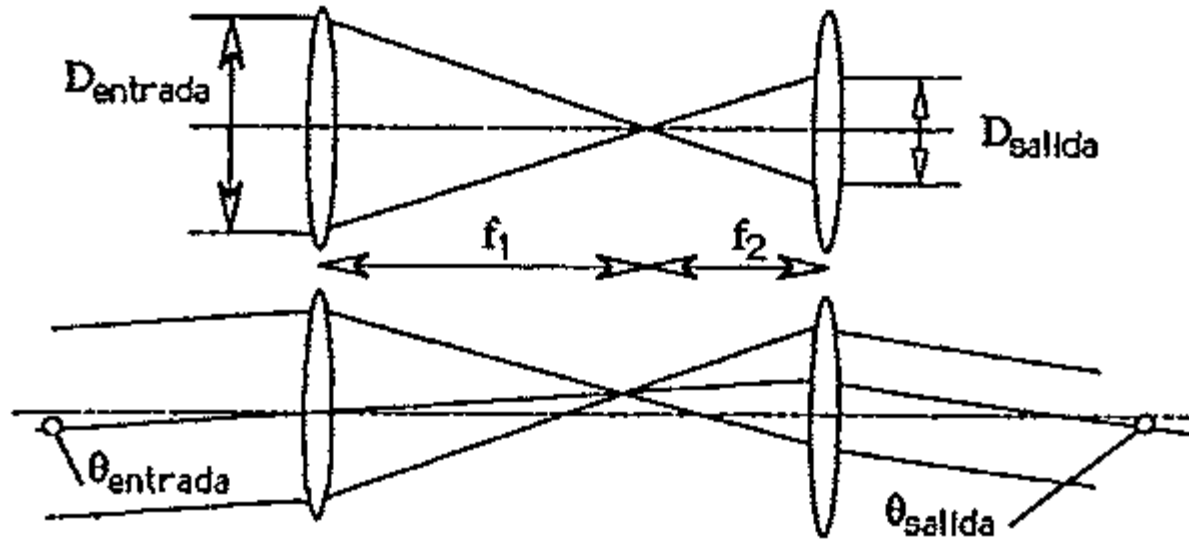
medulla

muscle

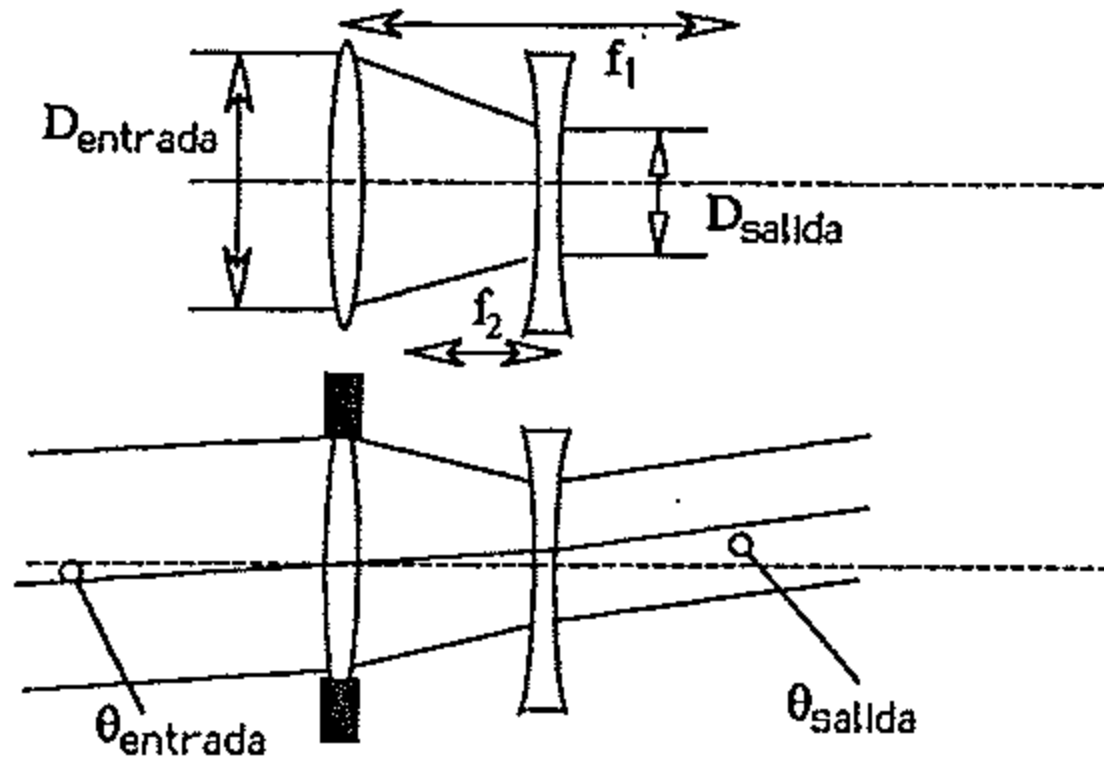
Figure 1

pollen

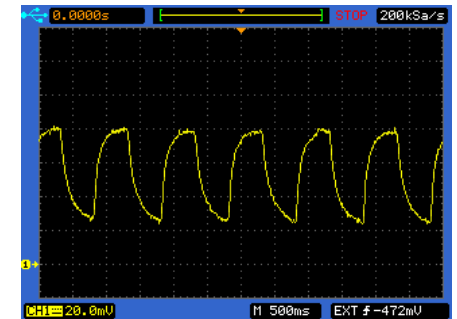
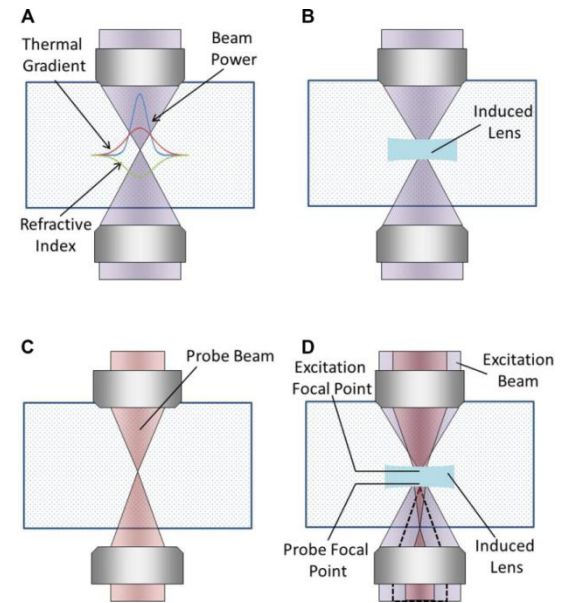
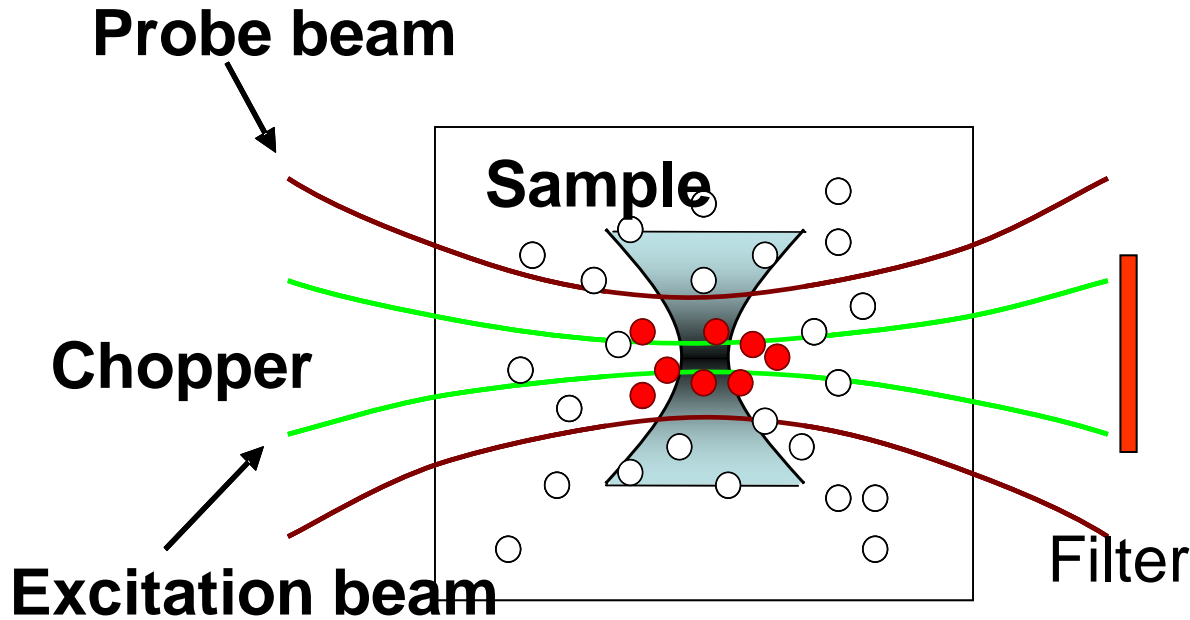
# Kepler Telescope



# Galileo Telescope



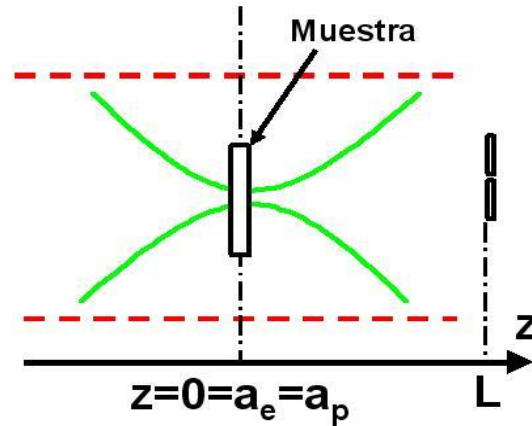
# Thermal lens effect and signal



$$I(r) = \frac{2P_e}{\pi w_e^2} e^{-2r^2/w_e^2}$$

# Physical mathematical model

$$z_p \gg L \gg z_e \quad ; \quad t \rightarrow \infty \quad ; \quad z_p \rightarrow \infty \quad ; \quad z = 0$$



$$S(z, t) = \Phi_0 \arctan \left\{ \frac{4m(z)\nu(z)t / t_c(z)}{\nu^2(z) + [1 + 2m(z)]^2 + [1 + 2m(z) + \nu^2(z)] 2t / t_c(z)} \right\}$$

$$I(r) = \frac{2P_e}{\pi w_e^2} e^{-2r^2/w_e^2}$$

$$D = \kappa / \rho C_p$$

$$t_c(z) = \omega_e^2(z) / 4D$$

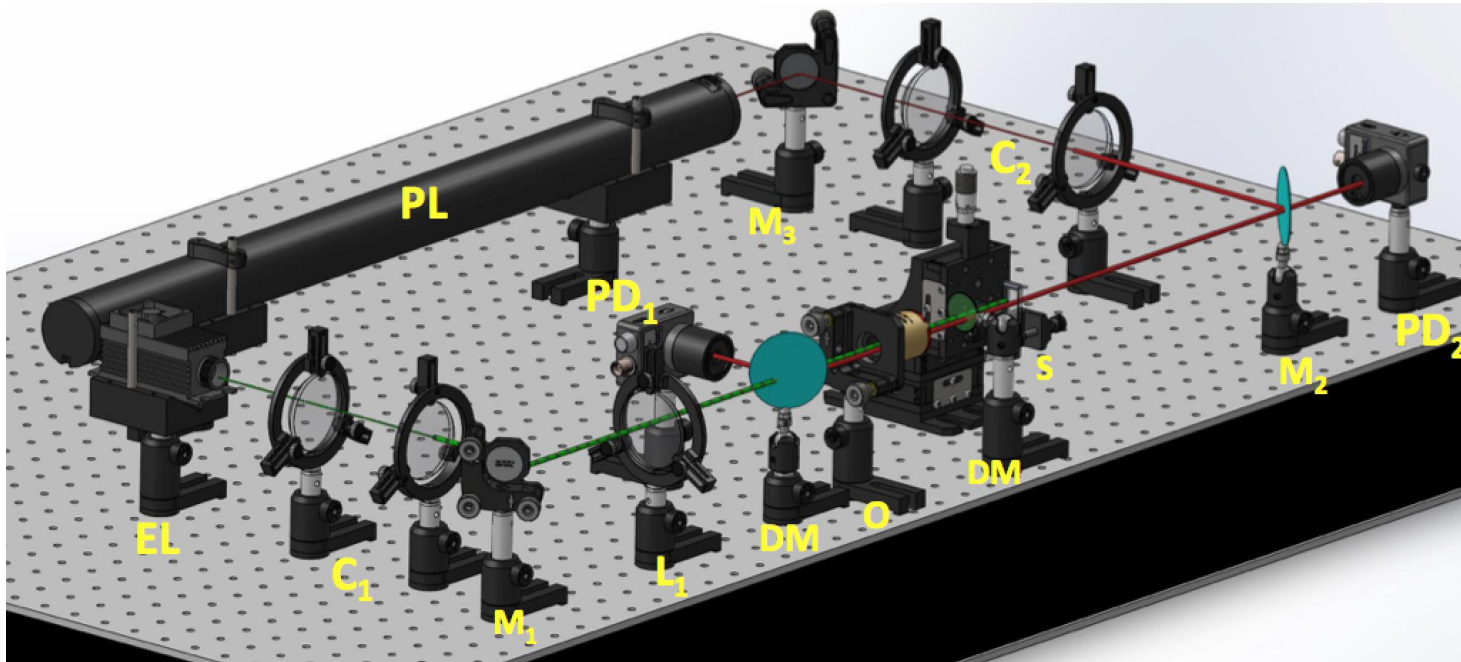
$$S_{max} = \pi \Phi_0 / 2$$

$$\Phi_0 = \frac{P_e \alpha l}{\lambda_p k} \frac{dn}{dT}$$

H Cabrera, J. Opt. Soc. Am. B, 23, 1408 (2006).

H. Cabrera, Appl. Phys. Lett. **94** 051103, (2009).

# Thermal lens microscopy set up



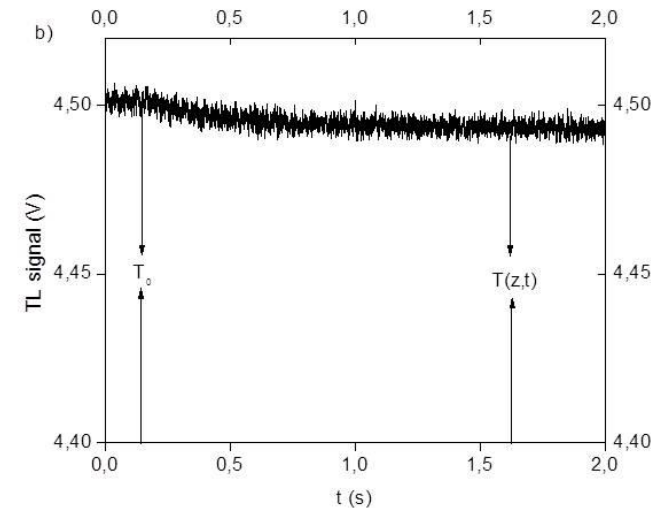
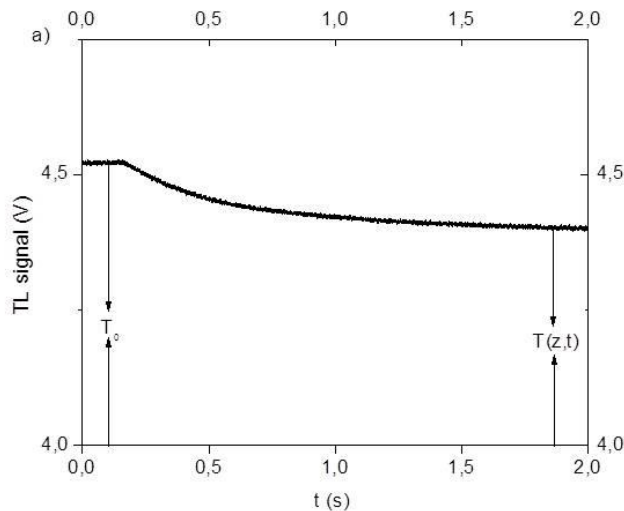
Schematic diagram of the TLM. S: sample;  $M_1 - M_3$ : mirrors; DM: dichroic mirror;  $L_1$ : lens; O: focusing objective lens; PD detector with pinhole and filter,  $C_1$  and  $C_2$  collimators; EL: excitation laser; PL: probe laser.



# Determination of the intracavity amplification factor $\eta$

$$S(z,t) = \frac{\pi P_e \alpha \eta l}{2\lambda_p k} \frac{dn}{dT}$$

$$S(z,t) = \frac{\pi P_e \alpha l}{2\lambda_p k} \frac{dn}{dT}$$



TL signal for ethanol sample as a function of time for an excitation power of 22 mW. The values of the probe beam aperture's transmission in the absence,  $T_0$ , and in the presence,  $T(z,t)$ , of the excitation beam are indicated, (a) TL signal for the sample inside the cavity and (b) TL signal for the sample without cavity.

## Experimental

$$S = -0.0265$$

$$dn/dT = -4 \times 10^{-4} \text{ } ^\circ\text{C}^{-1}$$

$$k = 1.67 \times 10^{-3} \text{ Wcm}^{-1} \text{ } ^\circ\text{C}^{-1}$$

$$\alpha = 6.8 \times 10^{-4} \text{ cm}^{-1}$$

$$l = 0.2 \text{ mm}$$

$$\lambda = 632.8 \text{ nm}$$

$$\eta = 14.9$$

$$S(z, t) = \frac{T(z, t) - T_0}{T_0}$$

$$S(z, t) = \frac{\pi P_e \alpha \eta l}{2 \lambda_p k} \frac{dn}{dT}$$

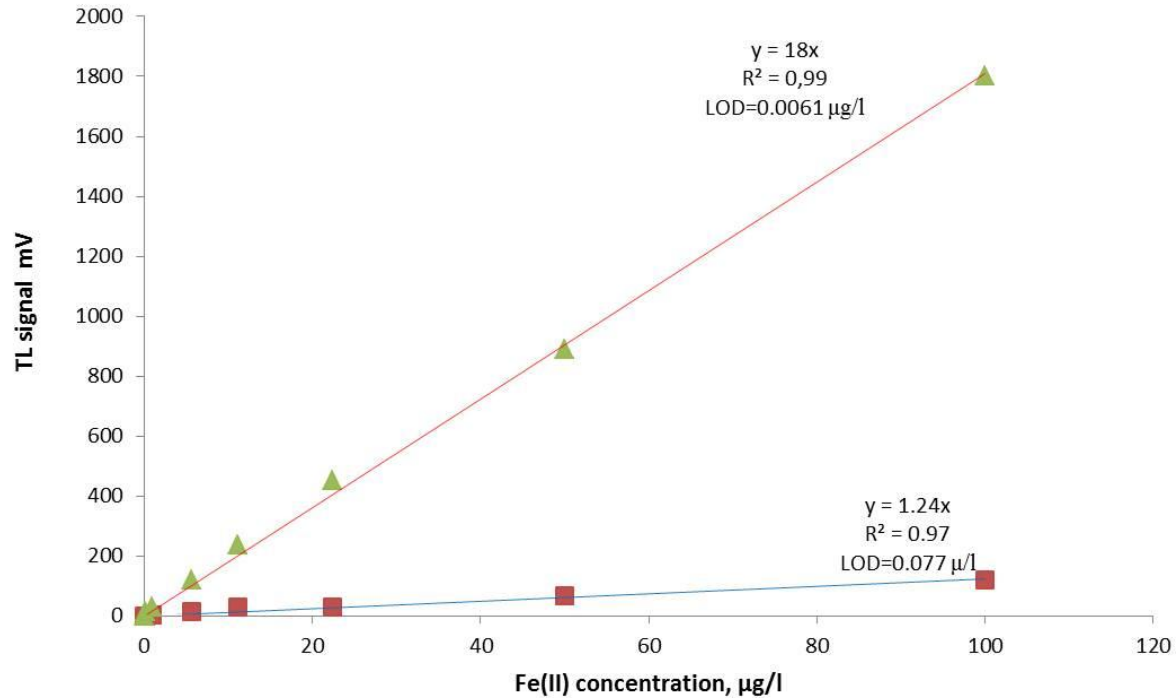
## Theoretical

$$R_M = 0.87$$

$$T_M = 0.13$$

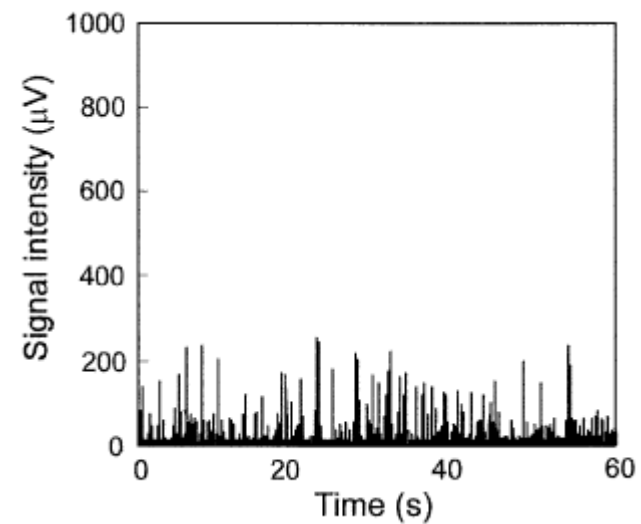
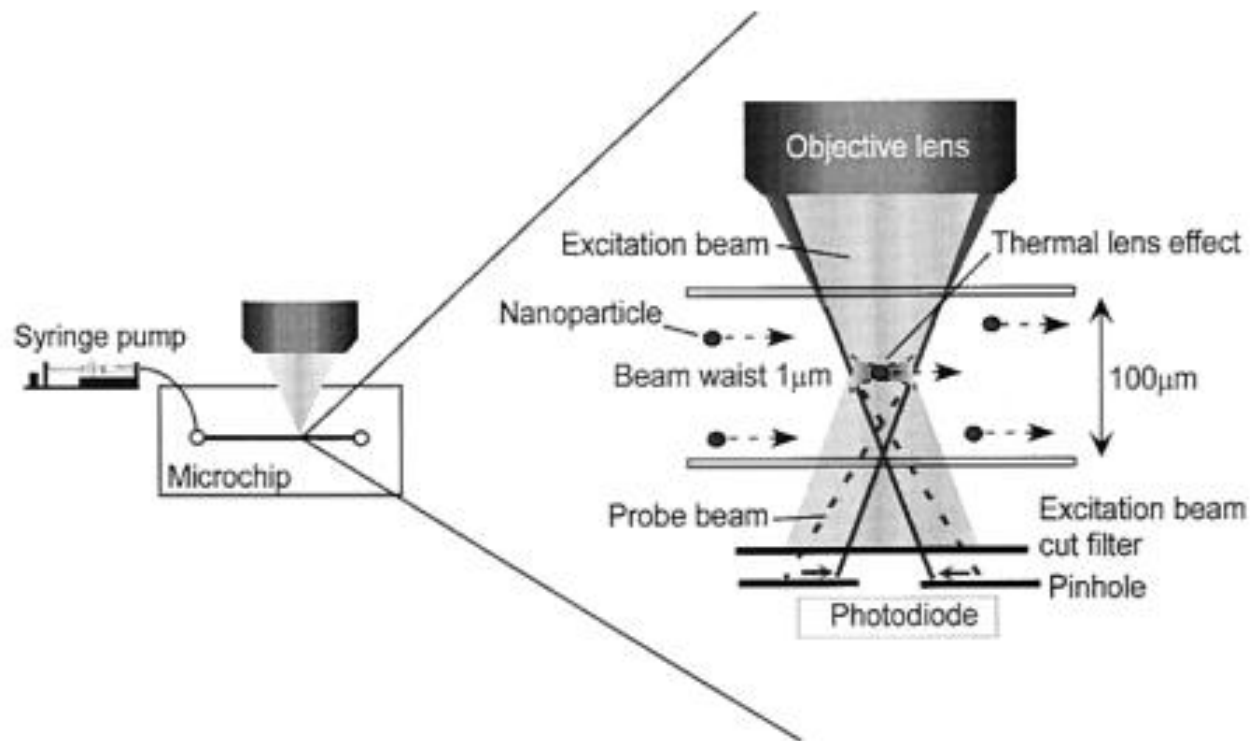
$$\eta = 2/T_M = 15.4$$

# Applications



$$LOD = \frac{3SD_{blank}}{m}$$

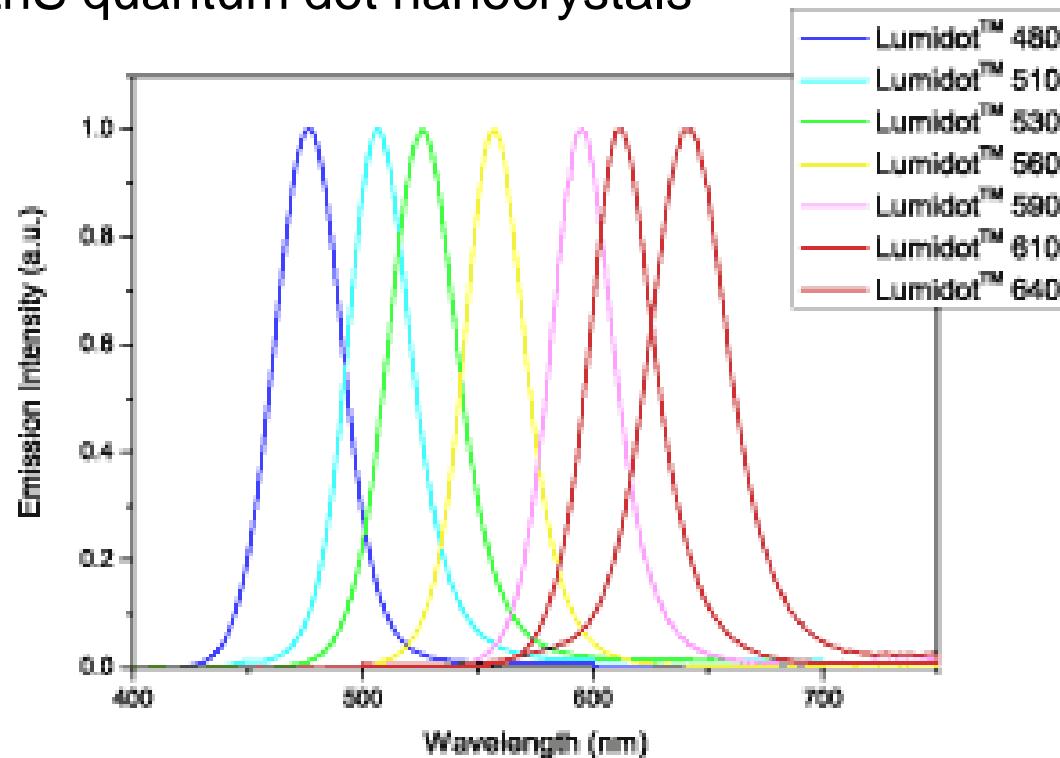
Calibration curves for Fe(II) solutions in 80% water with the addition of 20% of acetonitrile in 0.2 mm cell at 532 nm for the sample without cavity and inside a cavity



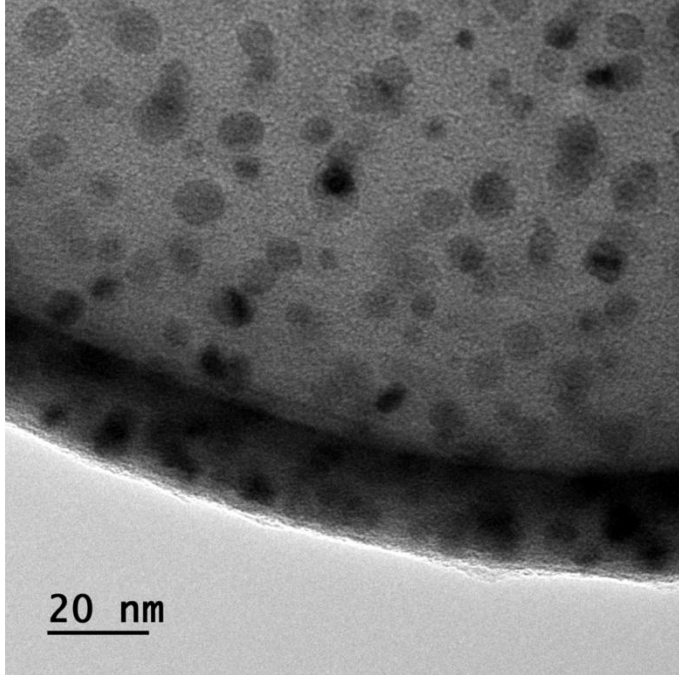
# Encapsulation efficiency of CdSe/ZnS quantum dots by liposomes

## Lumidot<sup>™</sup>: Quantum Dot Nanocrystals

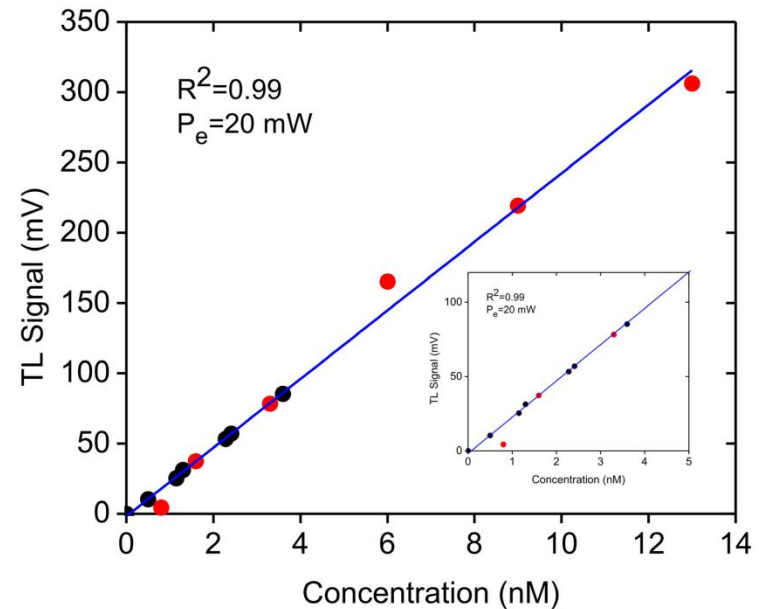
CdSe/ZnS quantum dot nanocrystals



Emission spectra of Lumidot<sup>™</sup> CdSe/ZnS nanocrystals



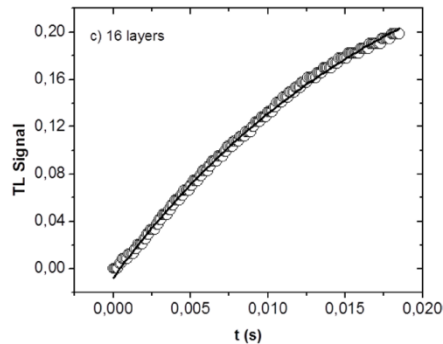
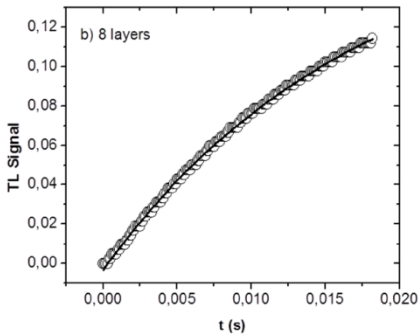
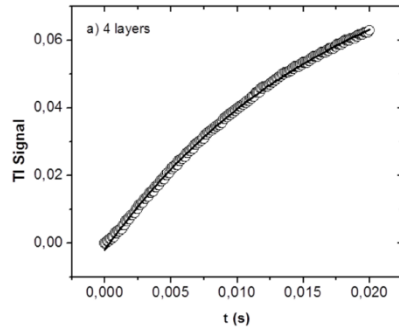
HR-TEM micrographs of the CdSe/ZnSe QDs encapsulated by liposome complexes (lipodots).



Calibration curve describing the TL intensity as a function of QDs concentration in aqueous solution 0 to 13 nM. The solid line represents a result of a best least squares linear fit with  $R^2 = 0.99$ . Red circles correspond to measurements for QDs in water (without liposomes) and the black circles correspond to measurements for lipodots (QDs inside liposomes).

Jessica Batalla, Humberto Cabrera, Eduardo San Martín-Martínez, Dorota Korte, Antonio Calderón and Ernesto Marín, “Encapsulation efficiency of CdSe/ZnS quantum dots by liposomes determined by thermal lens microscopy”, *Biomedical Optics Express* **6(10)**, 3898-3906 (2015).

# Thermal diffusivity of few-layers graphene



Typical time evolution of the TL signal for four (a), eight (b) and sixteen (c) graphene layers. Solid curves are the results of the best least squares fits to equation  
150 nm each layer

$$S(z, t) = \Phi_0 \arctan \left\{ \frac{4m(z)v(z)t / t_c(z)}{v^2(z) + [1 + 2m(z)]^2 + [1 + 2m(z) + v^2(z)]2t / t_c(z)} \right\}$$

$$D = (6.5 \pm 0.09) \times 10^{-4} \text{ m}^2/\text{s} \quad D = (1.9 \pm 0.07) \times 10^{-4} \text{ m}^2/\text{s}$$

$$D = (1.9^3 \pm 0.05) \times 10^{-4} \text{ m}^2/\text{s}$$

for four, eight and sixteen graphene layers respectively

Humberto Cabrera, Doroteo Mendoza, Jose Luis Benitez, Claudia Bautista, Salvador Alvarado Ramírez, Ernesto Marin, “Thermal diffusivity of few-layers graphene measured by an all-optical method”, Journal of Physics D: Applied Physics **48**, 465501 (2015).



For 5 persons



For 10 persons



*Thanks for your attention!*

For 2 persons (side by side)



For 3 persons



For 2 persons (face to face)



Basic concepts of microscopy