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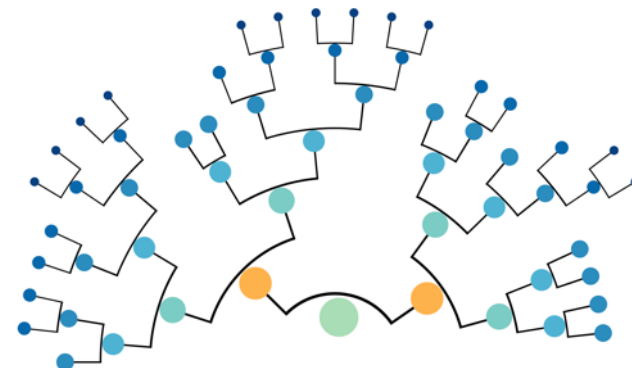
Virtual CECAM-ITCP School 2020

Using Molecular Simulation to Trace the Role of Conformational Dynamics in Enzyme Evolution

Caroline Lynn Kamerlin

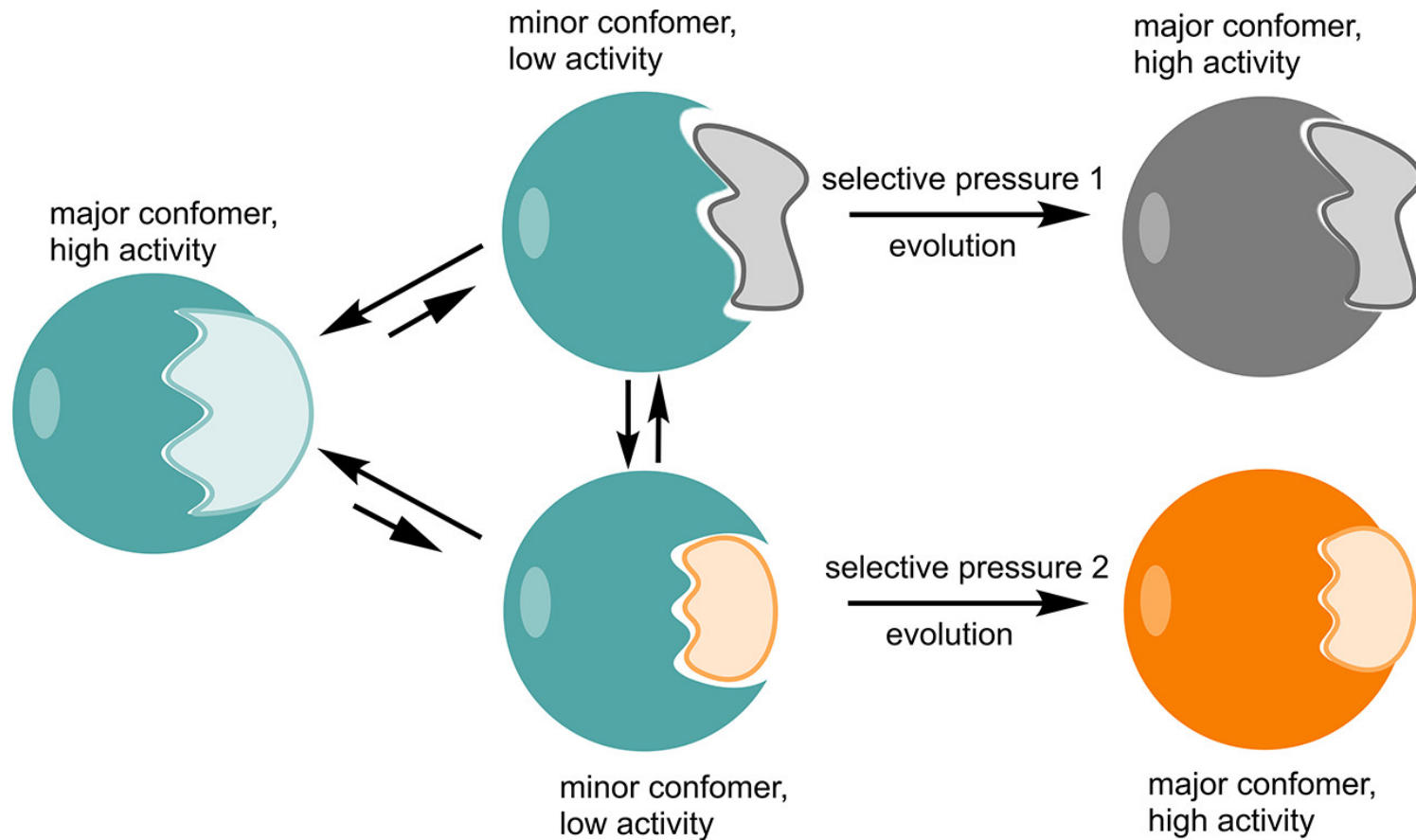
Department of Chemistry - BMC

Uppsala University





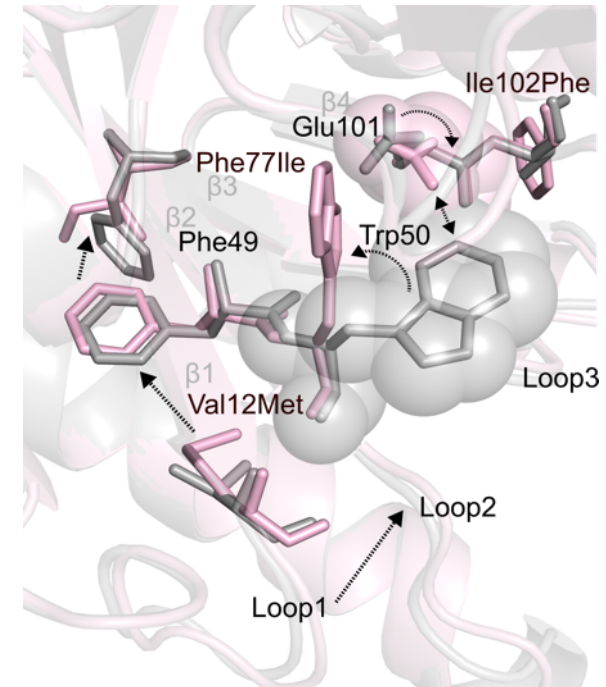
The Role of Conformational Diversity





(Just Some!) Examples of Systems

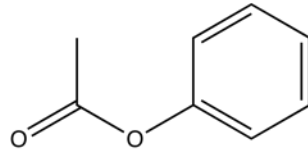
- Electrostatic cooperativity in alkaline phosphatases.
- Loop dynamics and scaffold flexibility controlling the selectivity of organophosphate hydrolases.
- Active site shuffling in a designed Kemp eliminase.
- Substrate and side chain dynamics during the emergence of new functions on non-enzymatic scaffolds, and in *de novo* active sites.



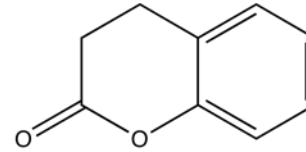
Regulating conformational dynamics appears to be critical for evolvability!



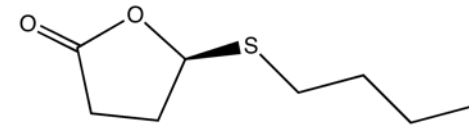
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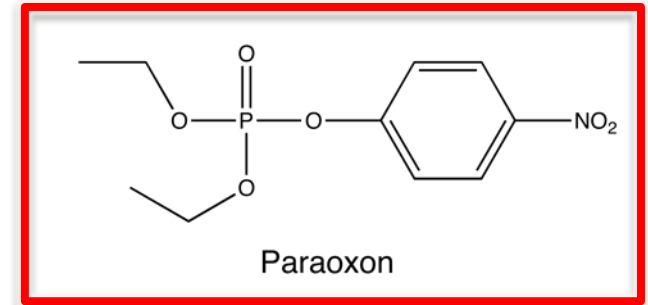
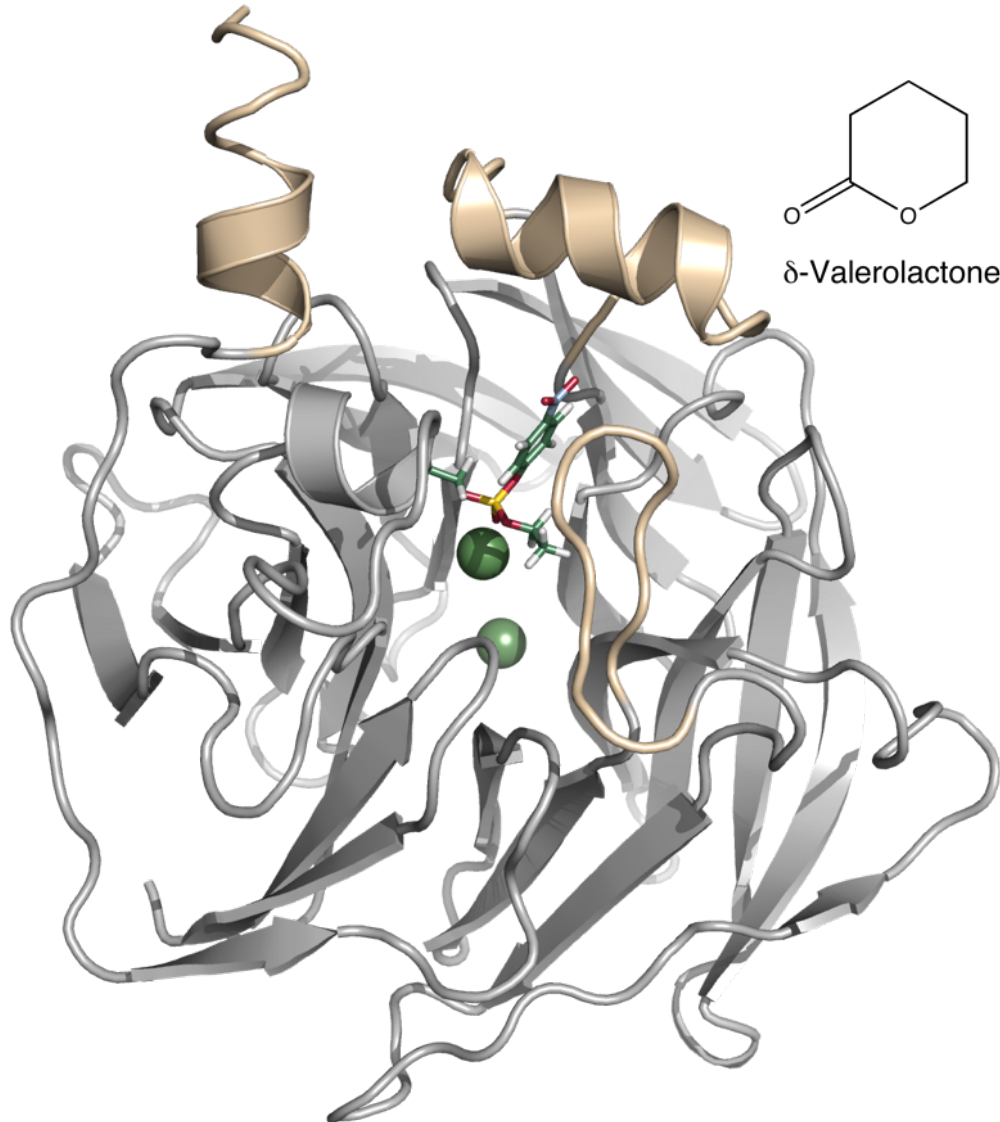
Phenyl Acetate



Dihydrocoumarin



L-5-(thiobutyl)-butyrolactone

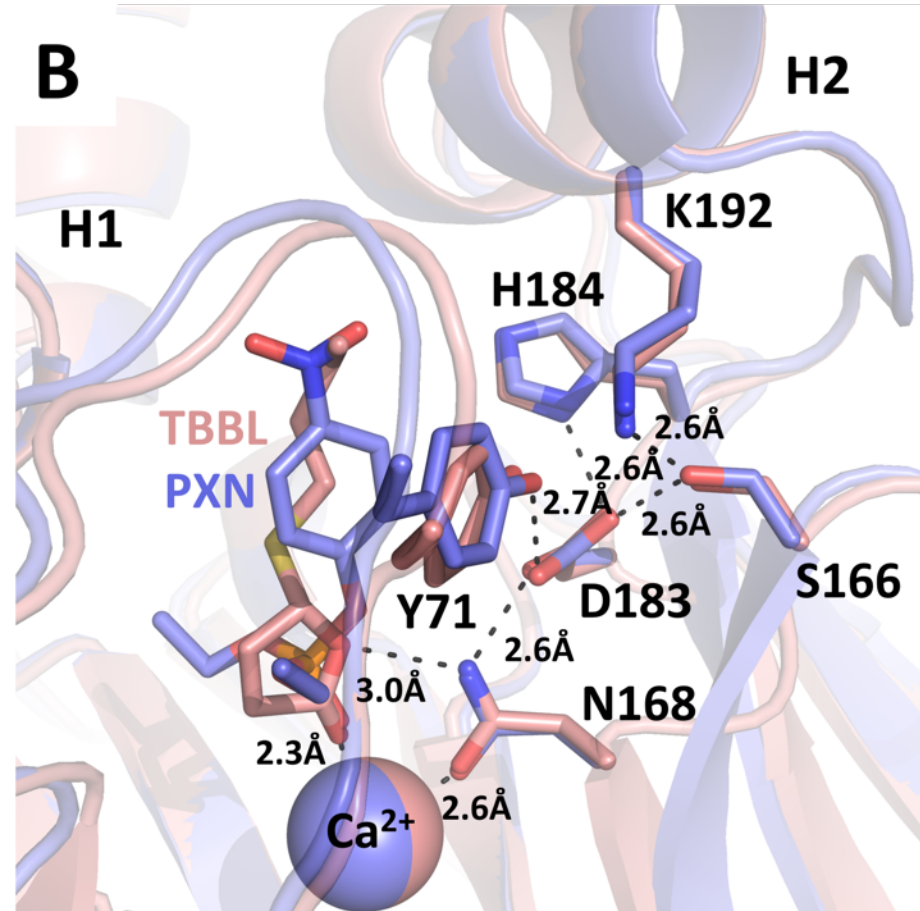
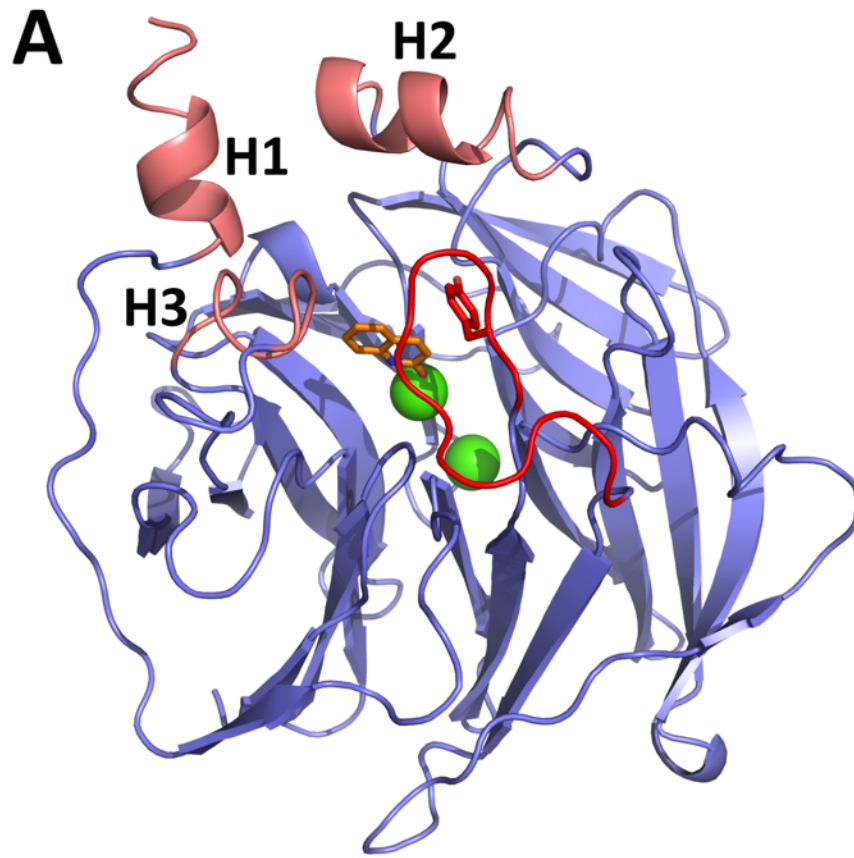


Serum paraoxonase 1 (PON1):

- Anti-atherosclerotic component of high density lipoprotein.
- **Extremely promiscuous** and highly evolvable enzyme.
- Very attractive as a therapeutic agent for treatment of acute organophosphate poisoning.



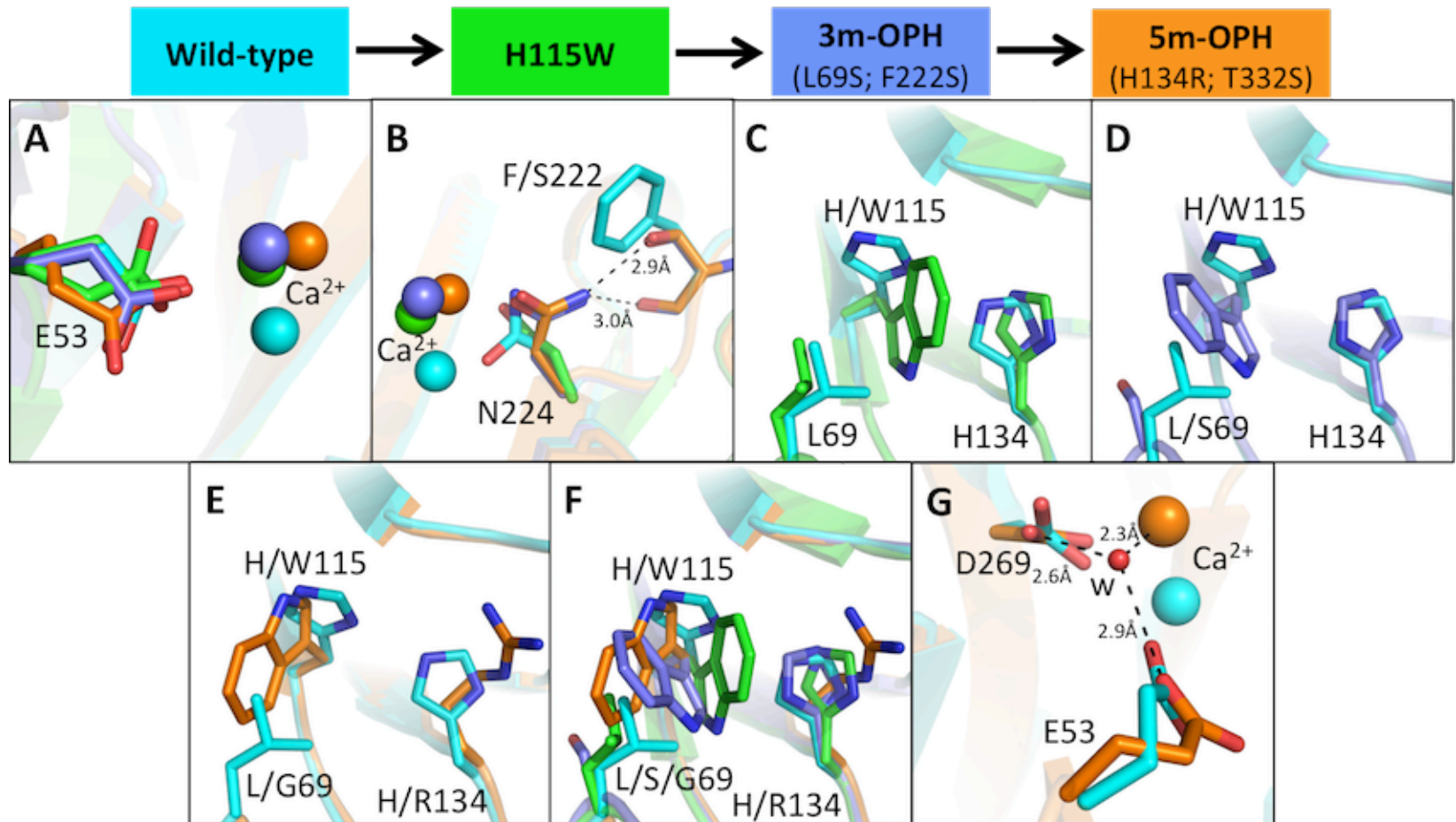
PON1 Active Site Architecture





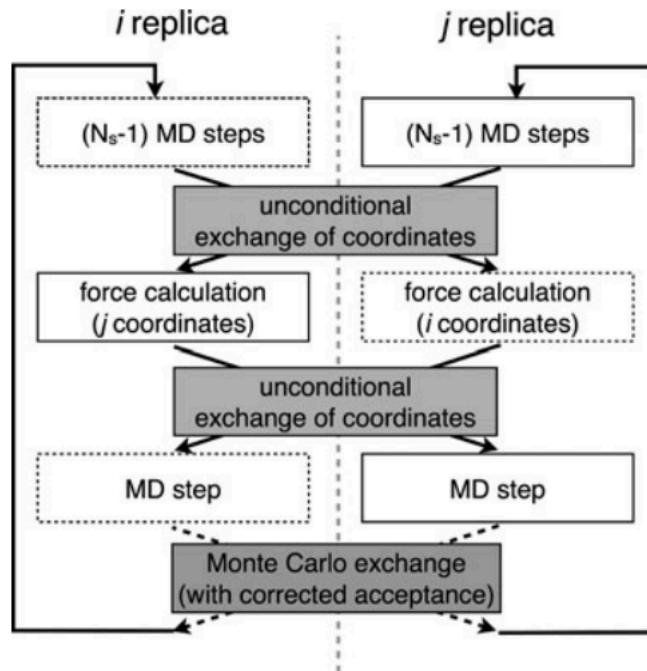
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PON1 Neo- vs. Re-Functionalization





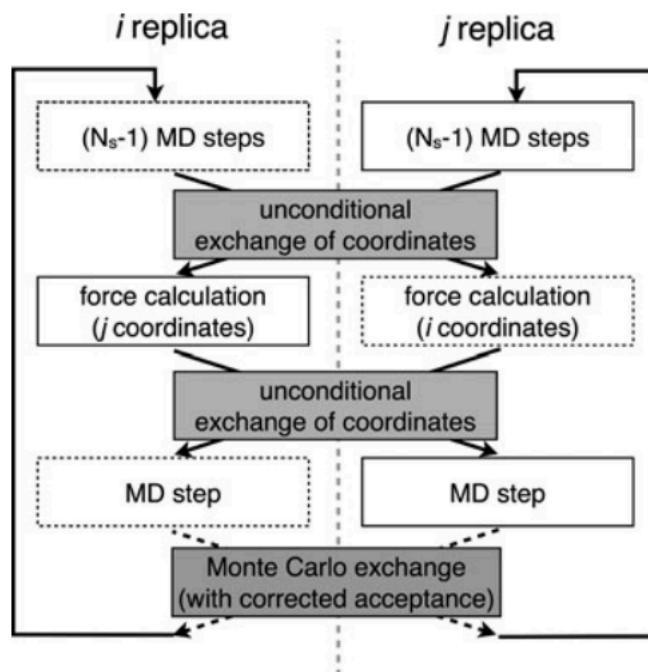
Hamiltonian Replica Exchange



- System has coordinates r + potential $U(r)$.
- Couple to thermal bath, so that probability of exploring a configuration is: $P(r) \propto e^{-\frac{U(r)}{k_B T}}$
- REX samples “cold” replica from which unbiased statistics can be extracted + “hot” replicas used to accelerate sampling.
- Hottest replica samples system fast enough to cross barriers for the process of interest, intermediate replicas smoothing.
- Normally replicas biased by temperature, HREX biased by temperature + potential.



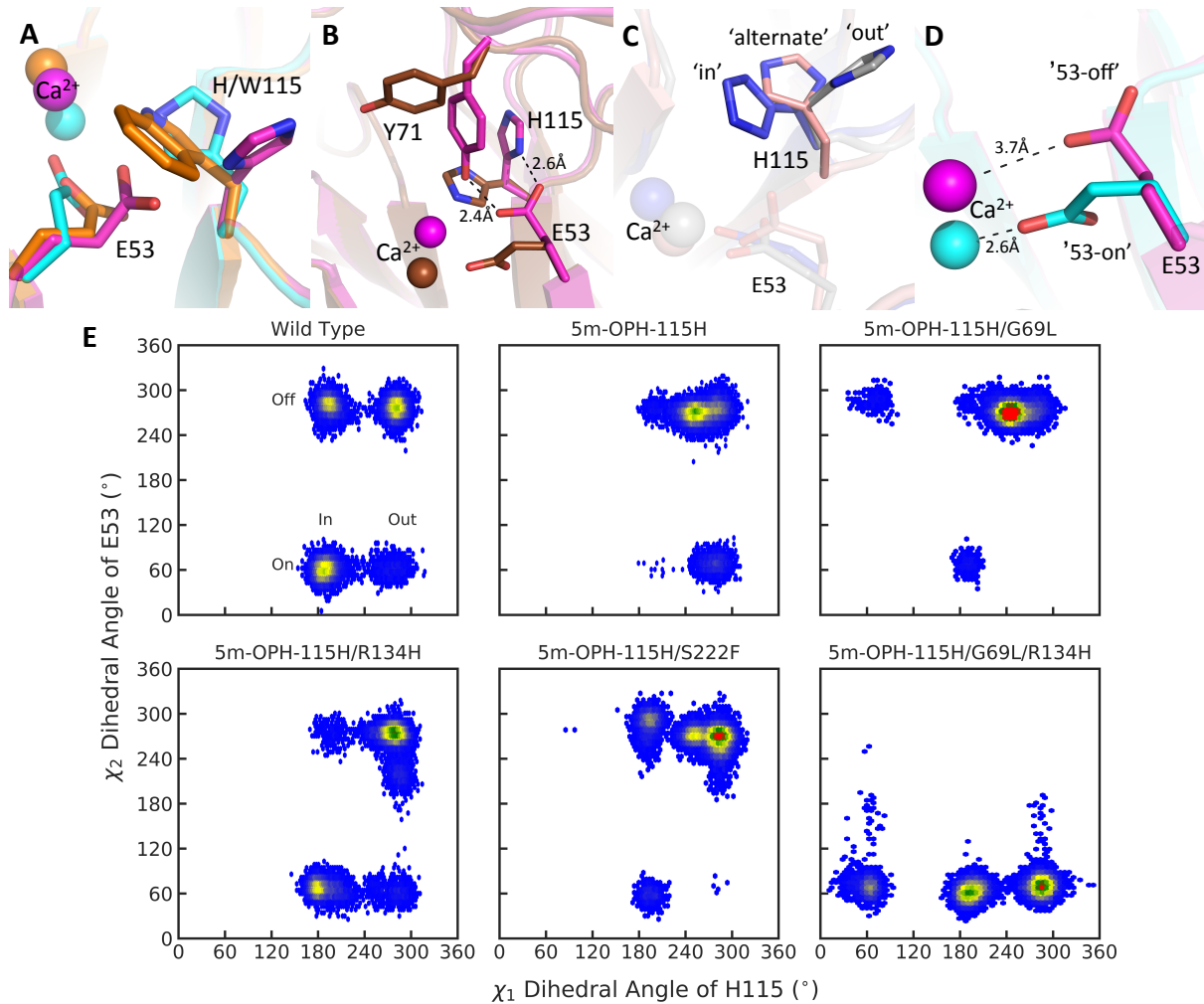
Hamiltonian Replica Exchange



- Simulate each replica at a different temperature with different potential.
- Energy is an extensive property (temperature is intensive) so in HREX can choose specific part of the system to sample (separate “hot”, H , and “cold”, C , regions).
- Charges, Lennard Jones and dihedral parameters of hot region scaled by $\sqrt{\lambda}$, λ , and λ (1^{st} and 4^{th}) or $\sqrt{\lambda}$ (1^{st} or 4^{th}).
- Interactions in hot region kept at $T_{\text{eff}} 1/\lambda$, H and C at $T_{\text{eff}} 1/\sqrt{\lambda}$ and in C at λ .
- λ is chosen to be a real number $0 < \lambda < 1$.

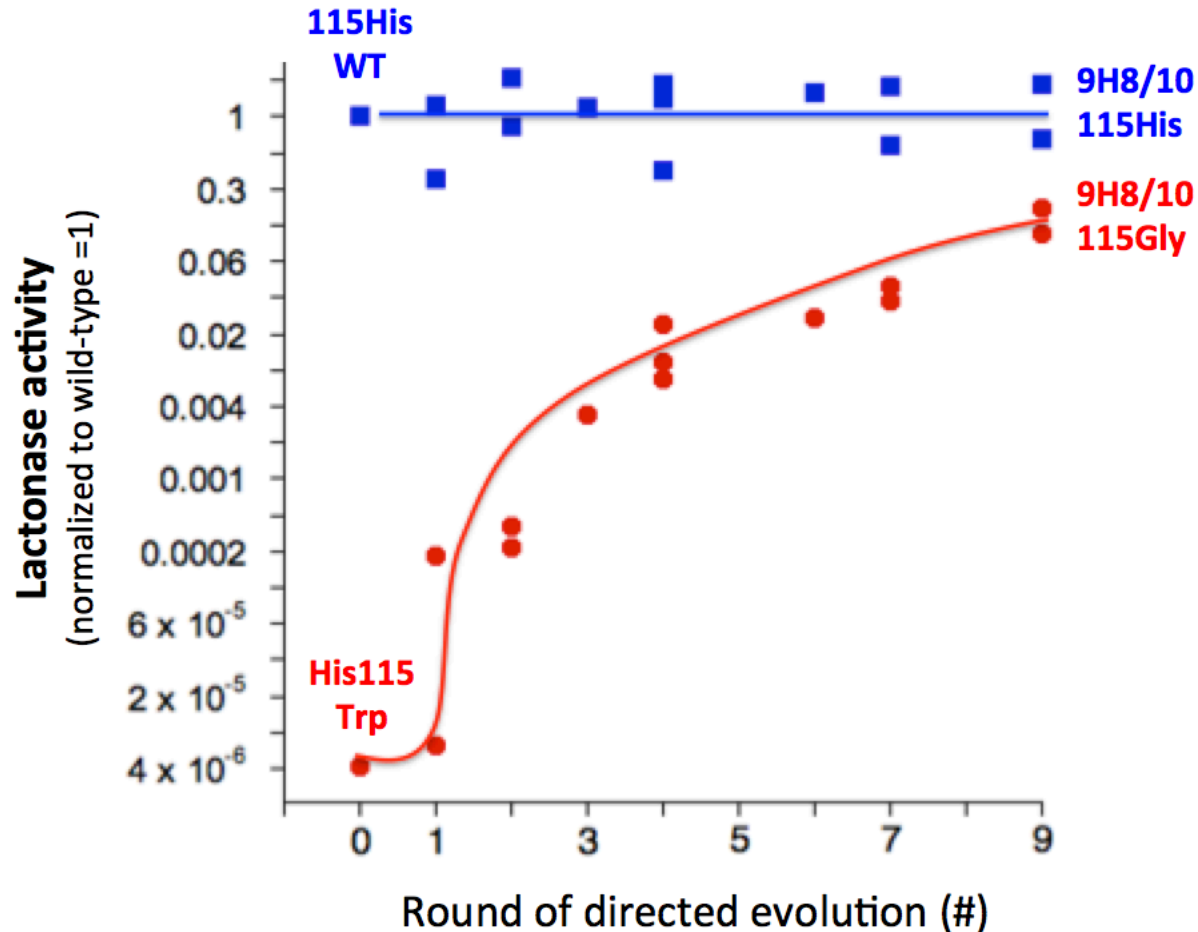


PON1 Neo- vs. Re-Functionalization





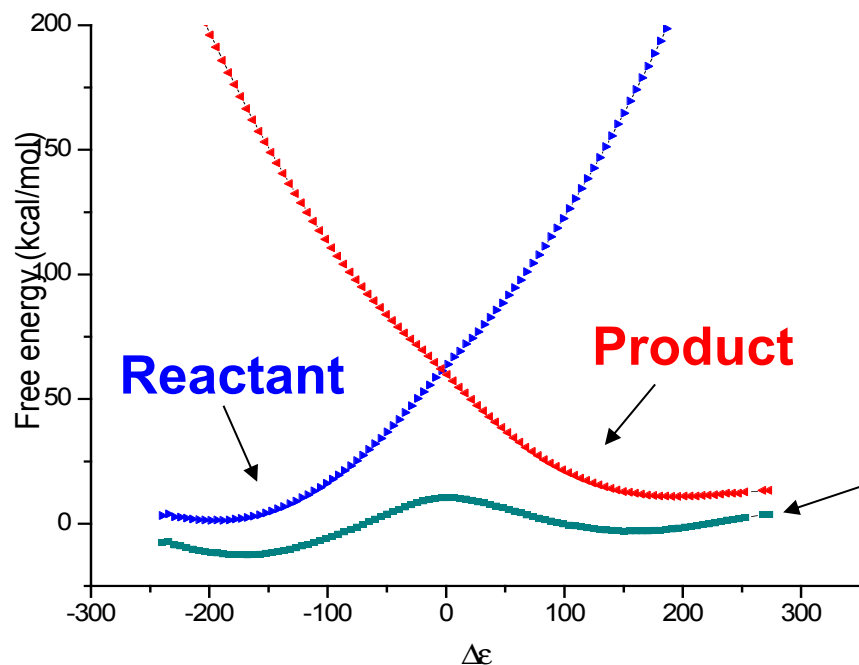
PON1 Neo- vs. Re-Functionalization





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Empirical Valence Bond Approach



Reactant:

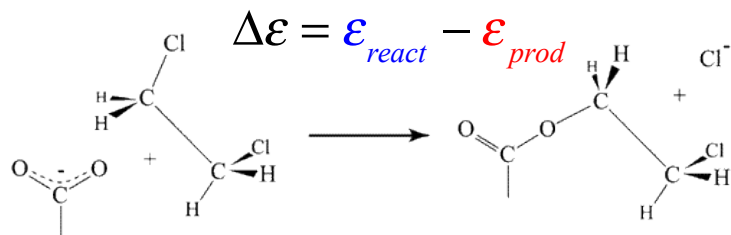
Force field-like functions describing the reactants' bonding pattern

Product:

Force field-like functions describing the products' bonding pattern

Ground State:

Eigenvalue of 2x2 Hamiltonian built from Reactant and Product energies and off-diagonal function (H_{12}).

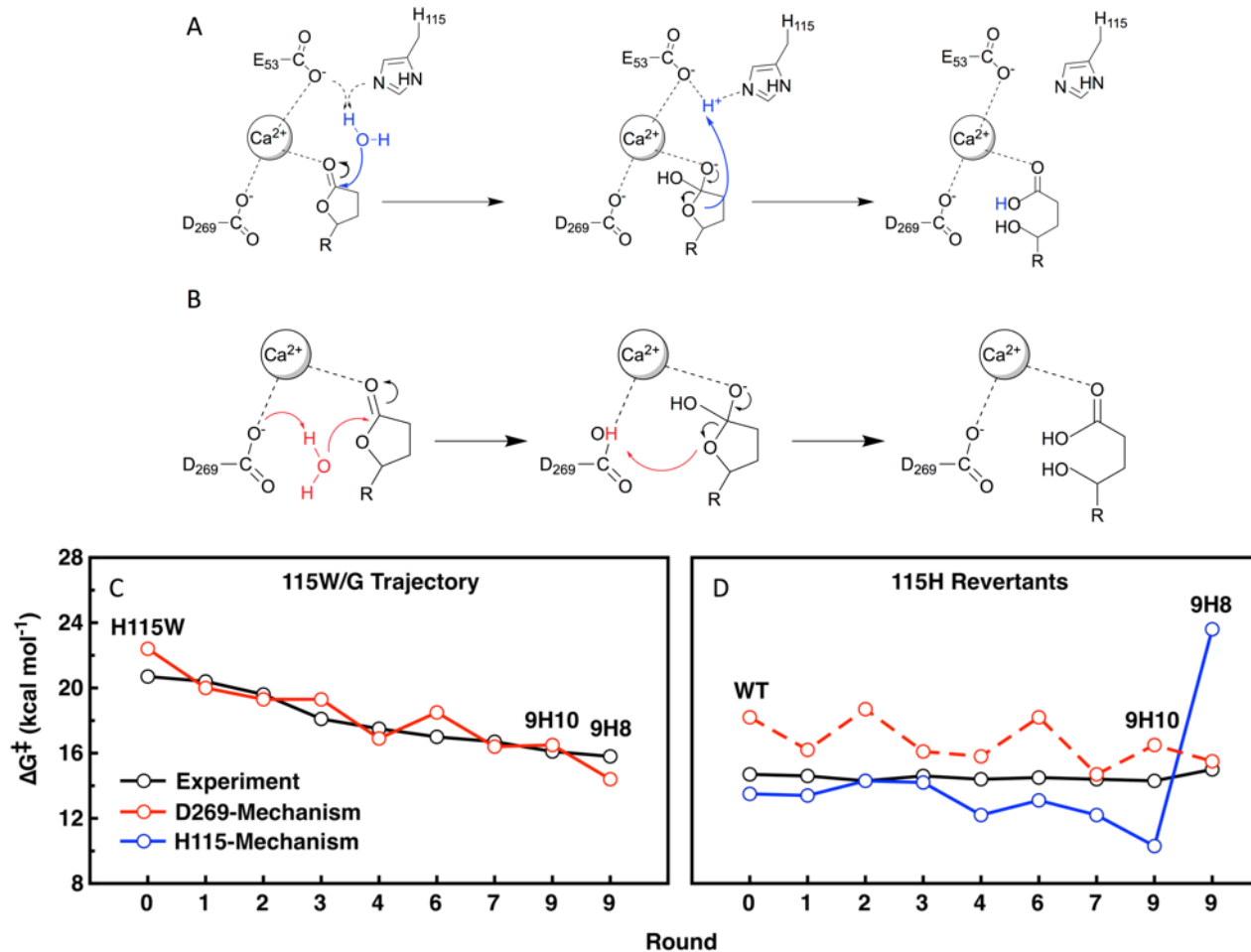


$$H = \begin{pmatrix} \epsilon_{\text{react}} & H_{12} \\ H_{12} & \epsilon_{\text{prod}} \end{pmatrix}$$



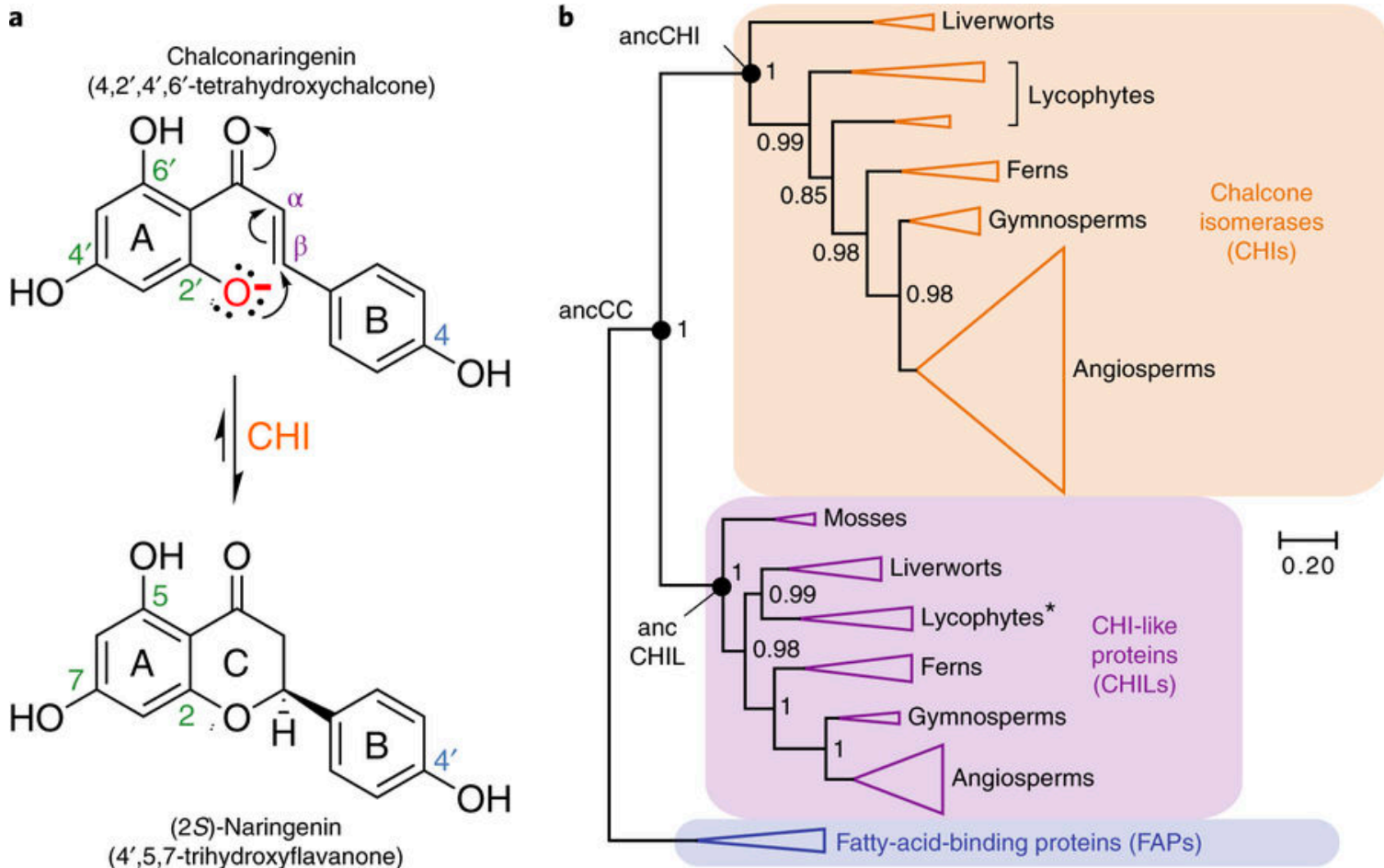
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PON1 Neo- vs. Re-Functionalization



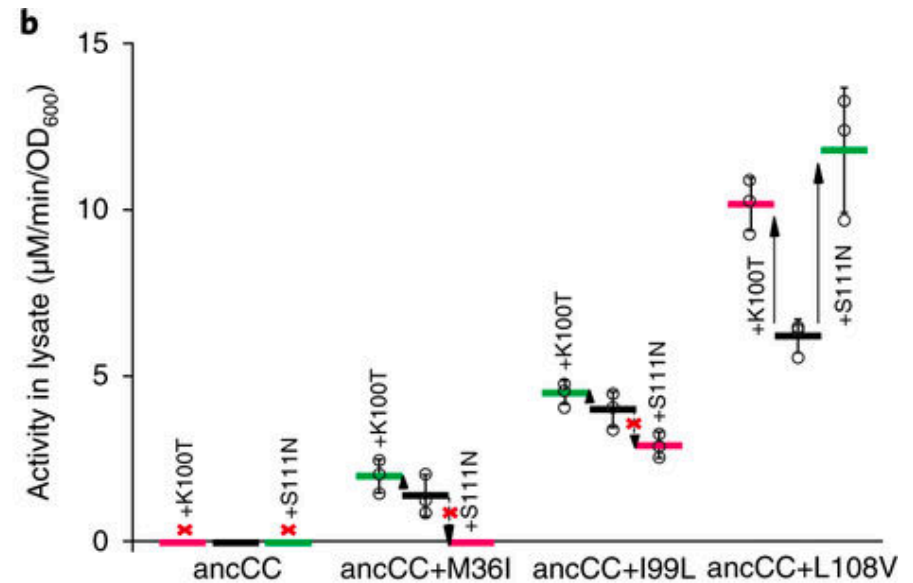
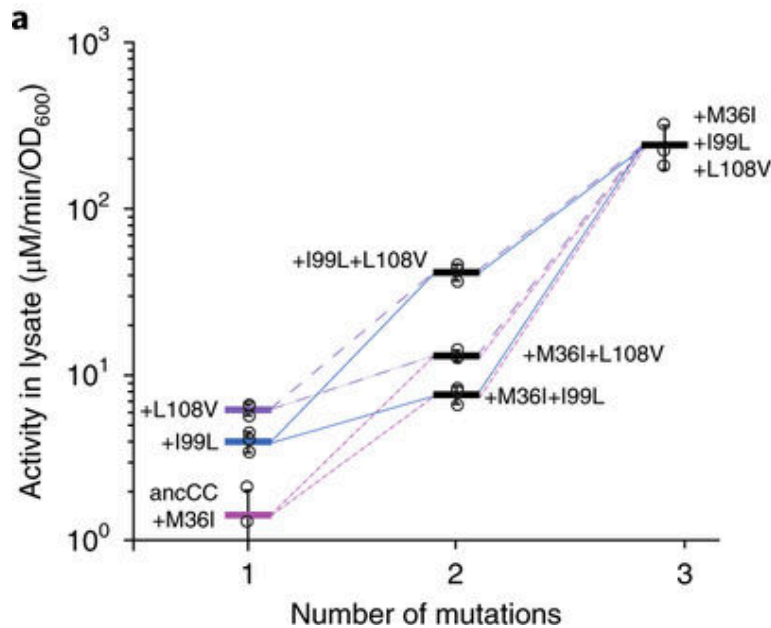


How Do New Enzymes Emerge?



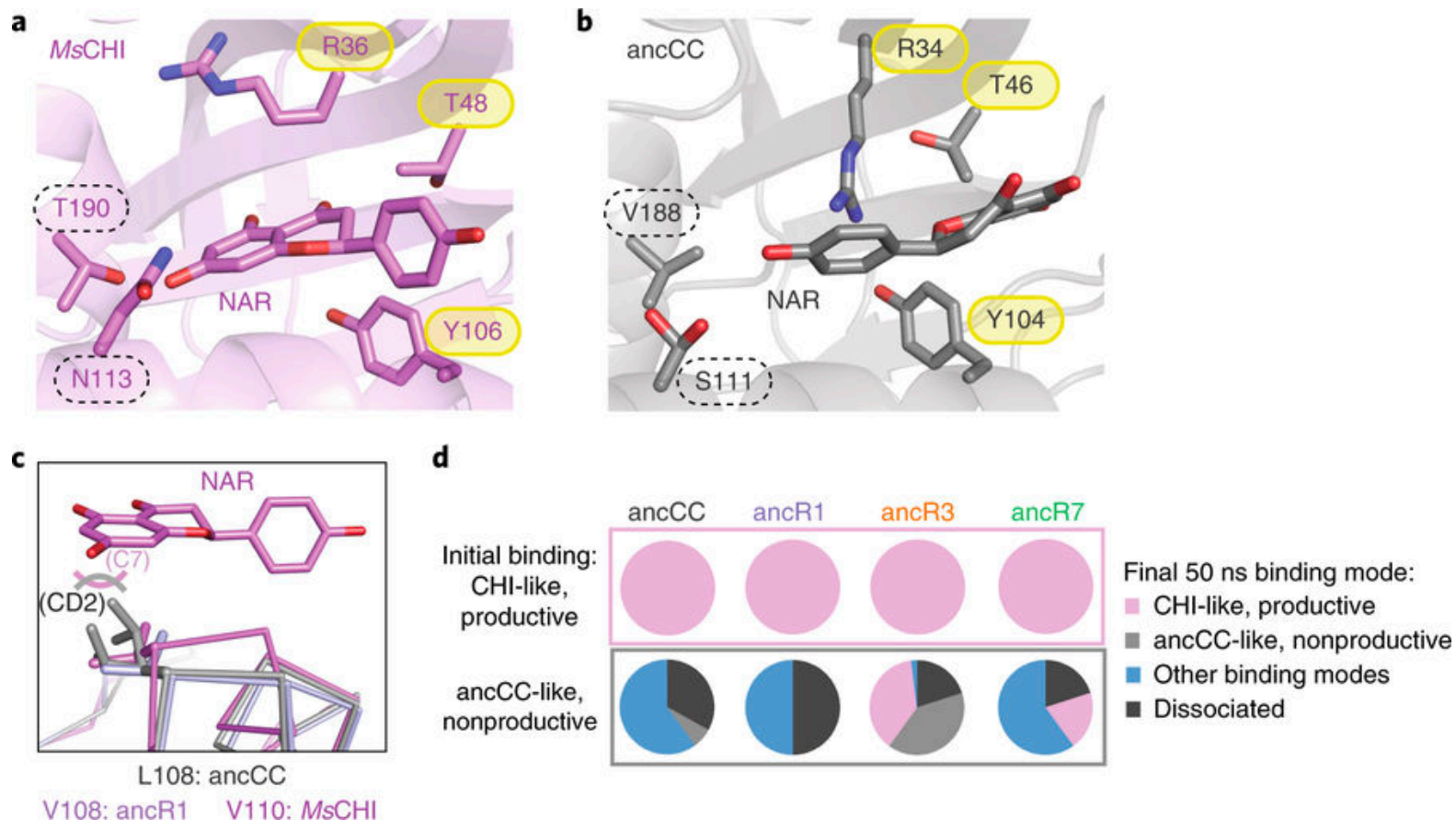


Additivity vs. Epistasis in CHI Evolution





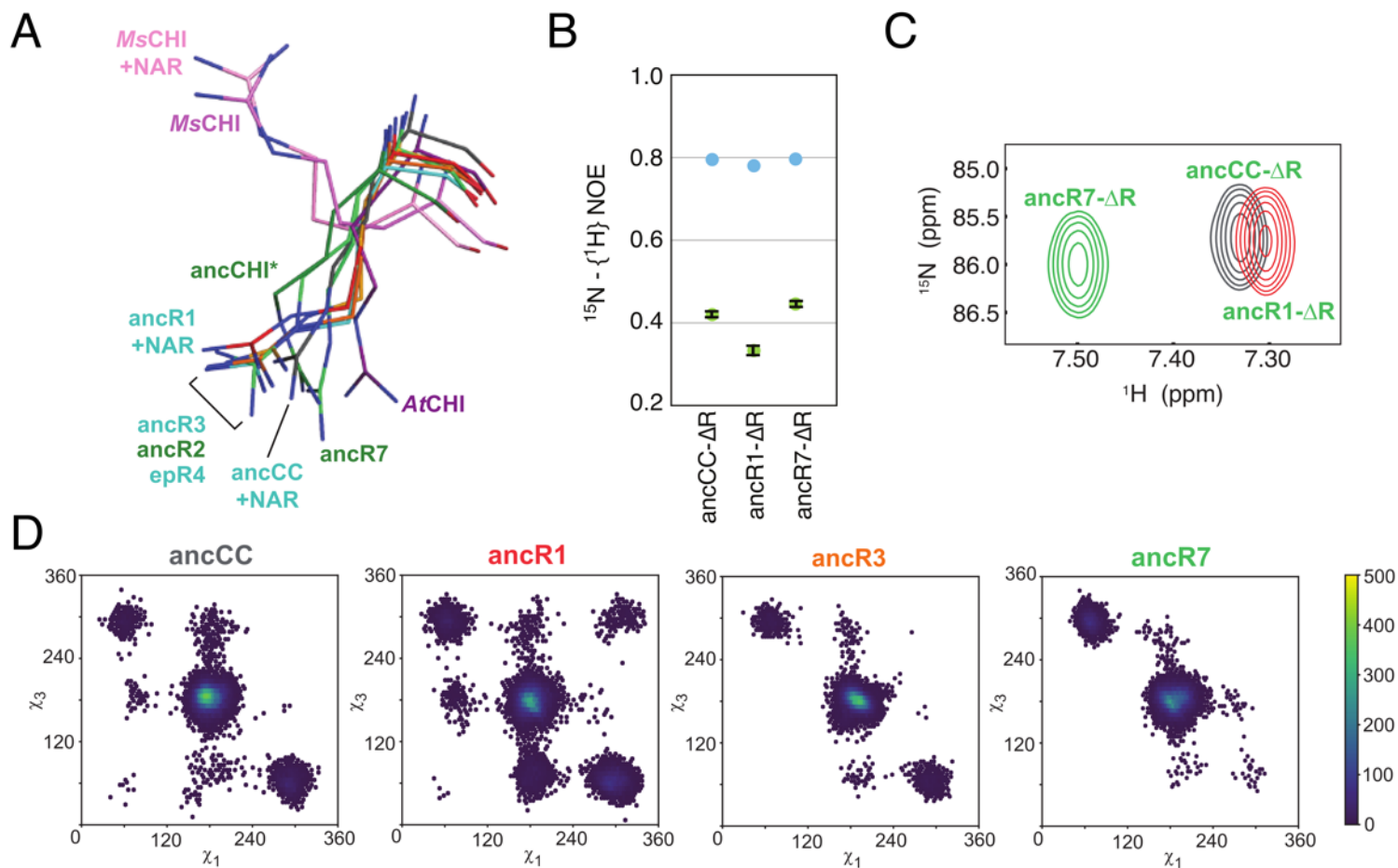
Structural Changes During Evolution





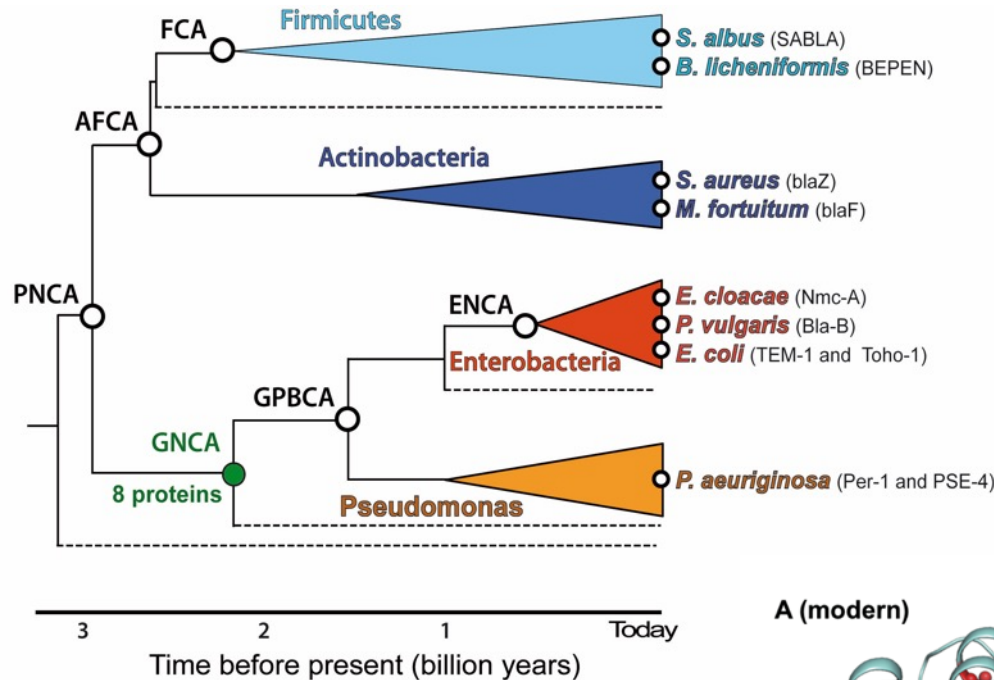
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Evolution Rigidifies a Key Residue



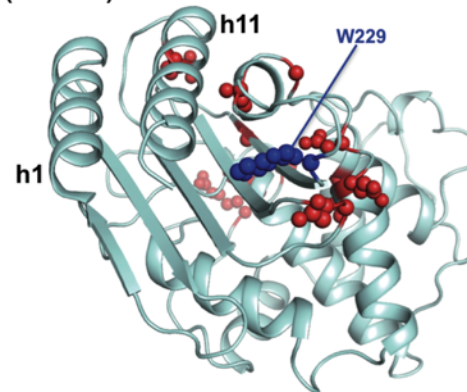


De Novo Active Sites in β -Lactamases

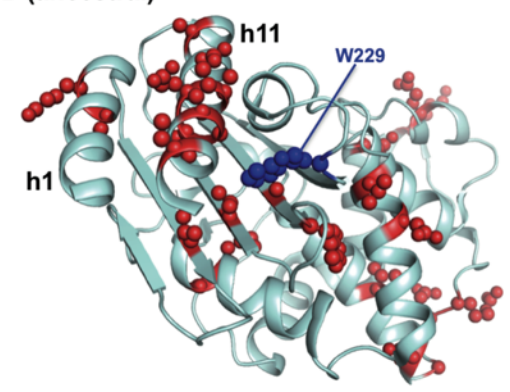


Generating *de novo* active sites, put into resurrected Precambrian β -lactamases, identified through ancestral sequence reconstruction.

A (modern)

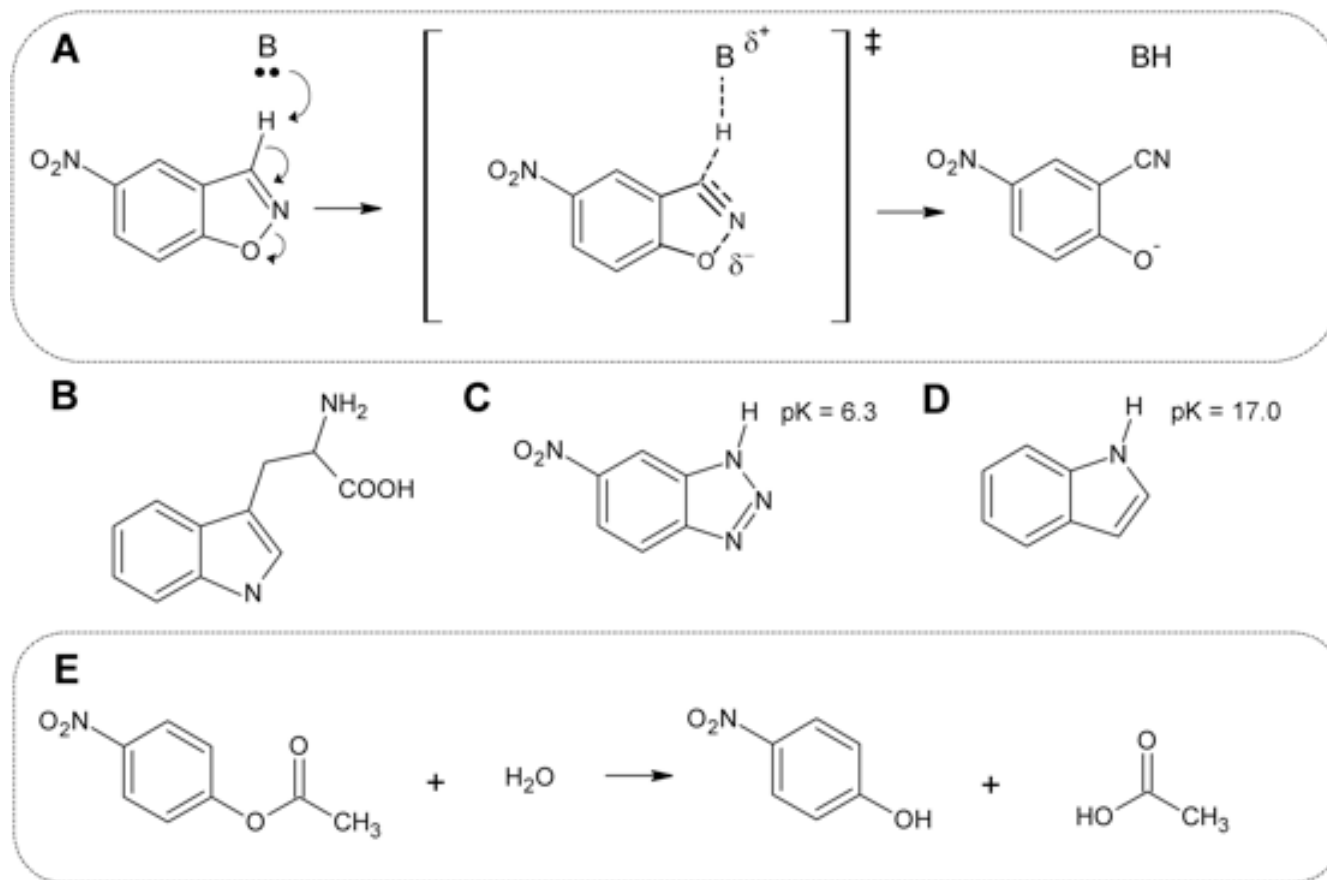


B (ancestral)



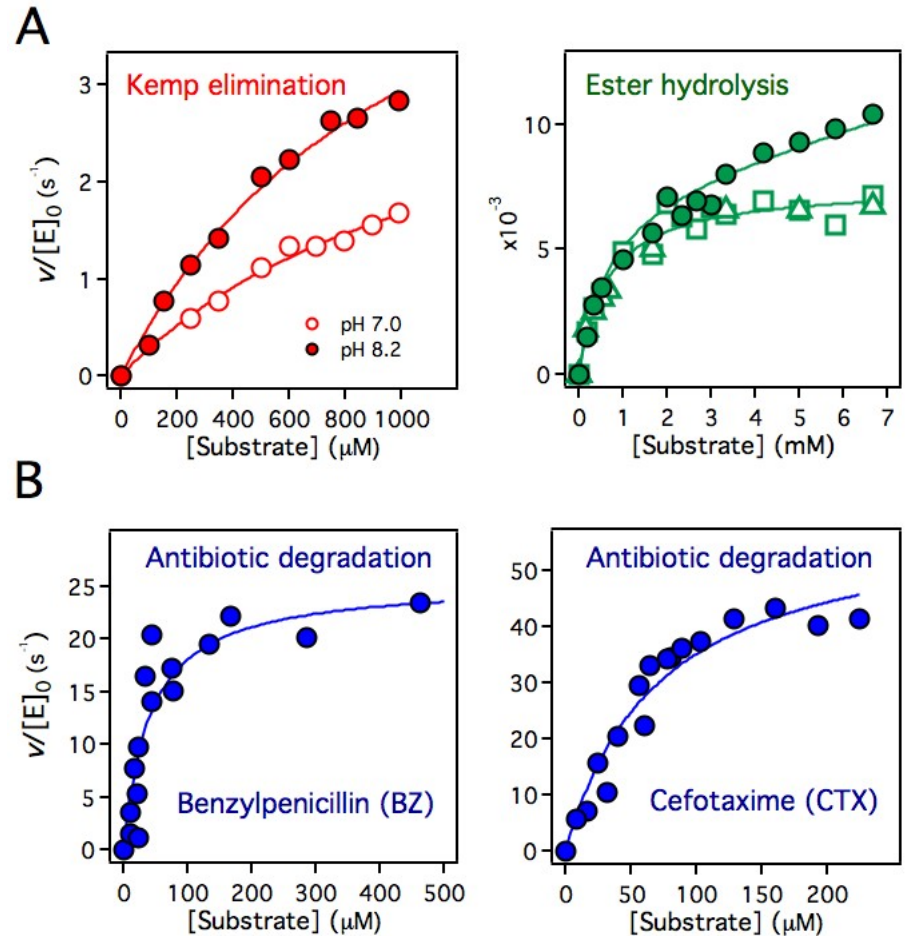
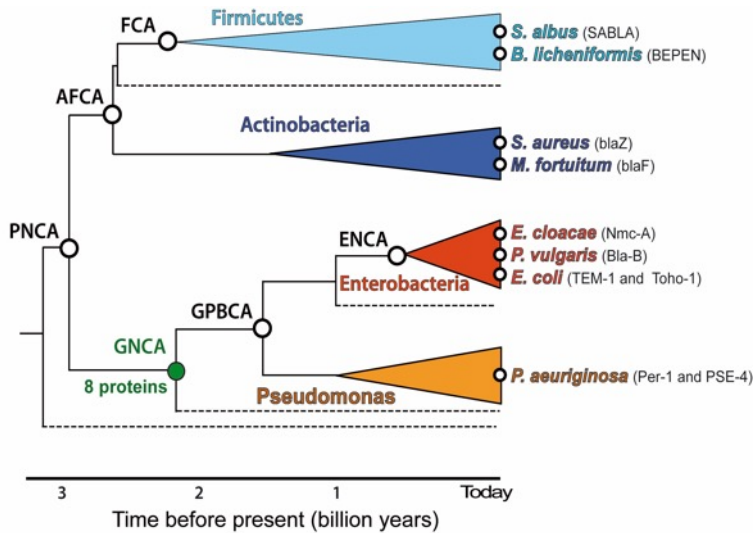


De Novo Active Sites in β -Lactamases



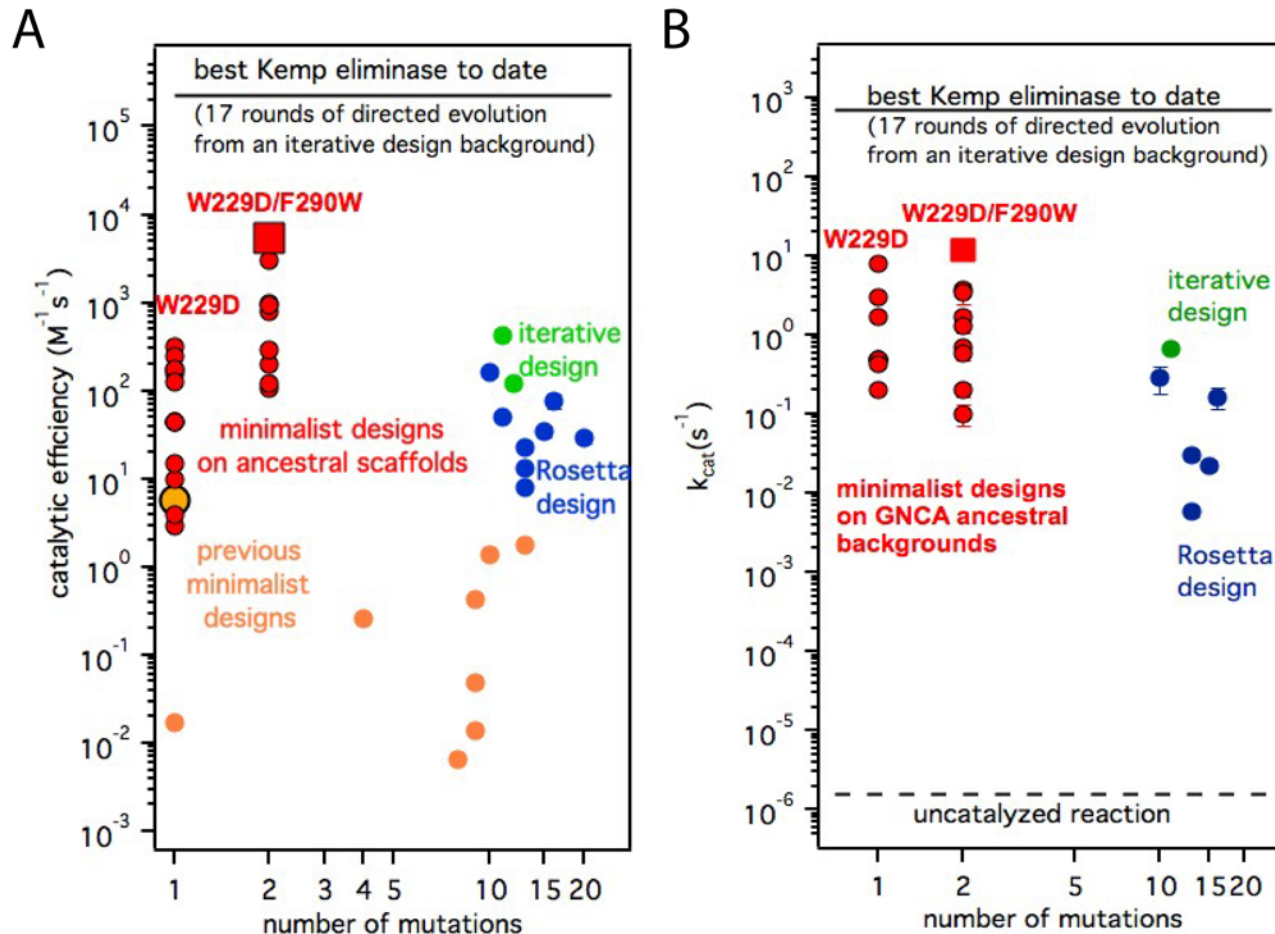


De Novo Active Sites in β -Lactamases





De Novo Active Sites in β -Lactamases





Random Library Screening

Clone	$k_{\text{cat}} / K_{\text{M}}$ ($\text{M}^{-1} \text{s}^{-1}$)	T_{M} ($^{\circ}\text{C}$)
GNCA4-WT	3047±282	80
3C11	608±68	77
4B4	1770±126	81
8F11	5980±117	80
6D5	2476±420	81
7C1	600±56	72
8E12	2222±167	70
6A12	1036±159	79
7D1	1880±155	67
2H4	2280±146	ND
5H8	2066±67	64

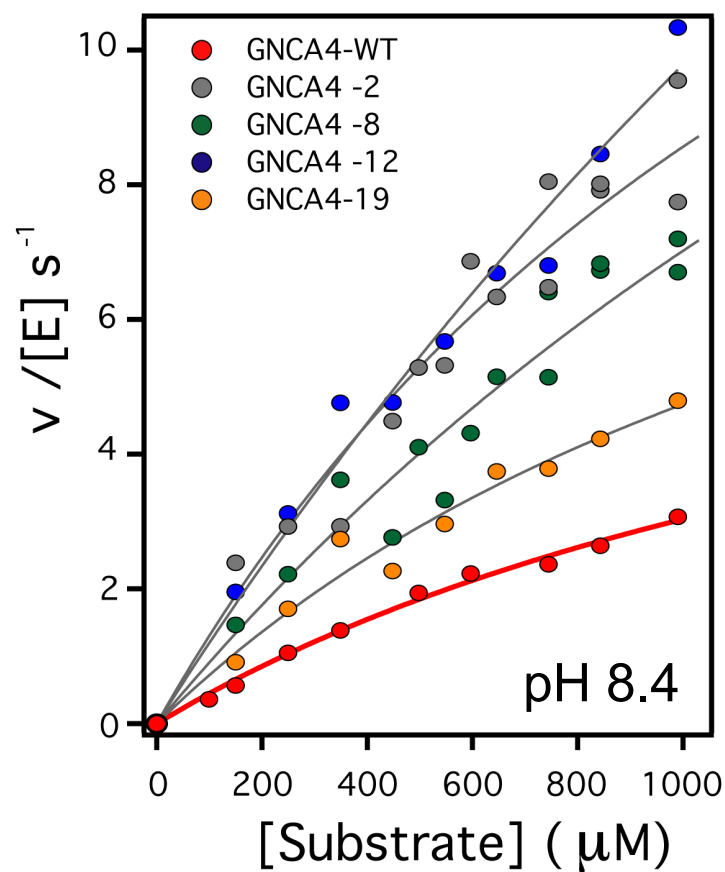
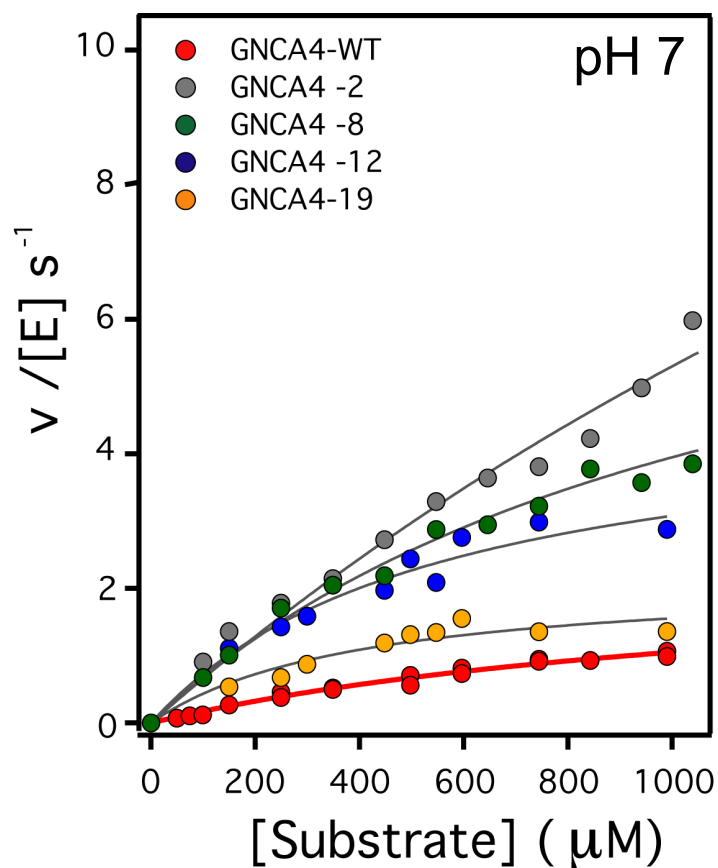
Library of variants with random mutations / average mutational load of 3-5 mutations:

- 522 tested, 300 with greatly diminished activity
- Best variant carried 6 mutations, only 2-fold more active than wild-type



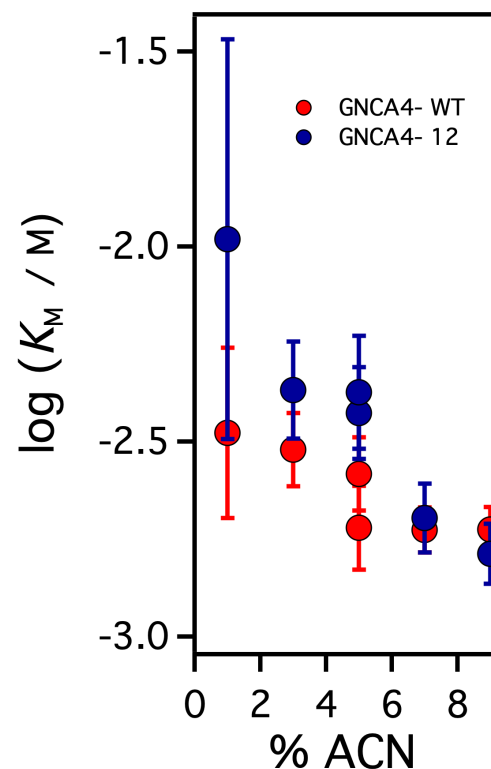
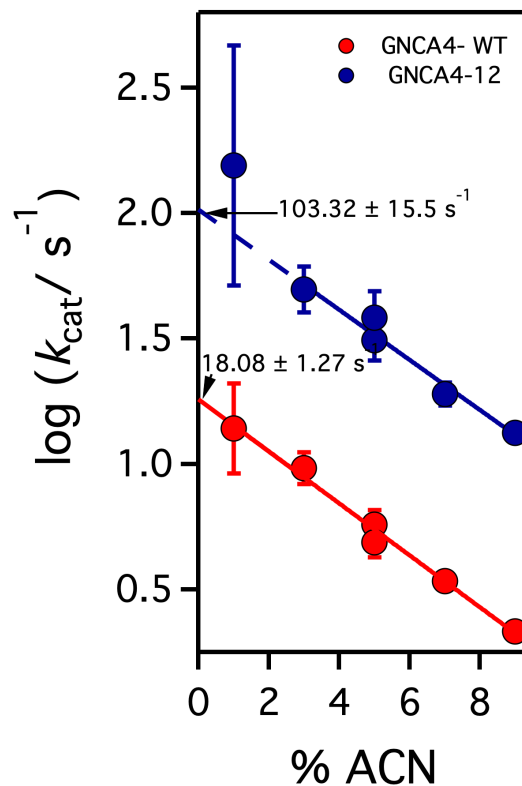
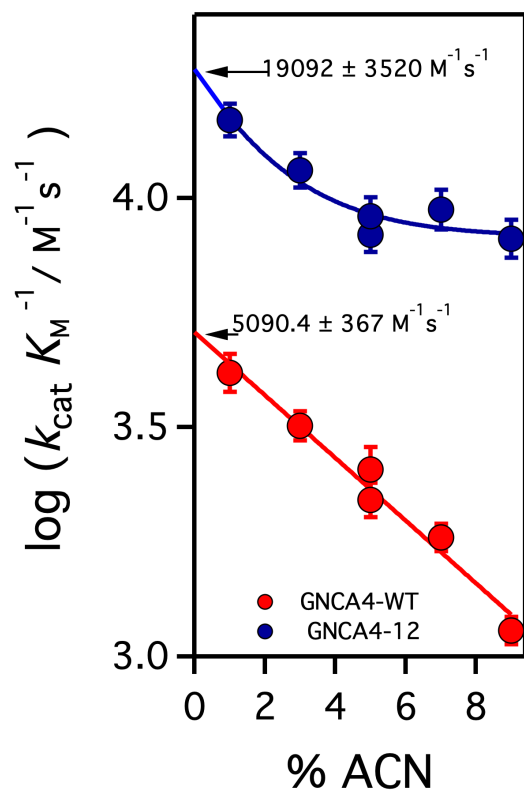
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Activity Enhancement with FuncLib





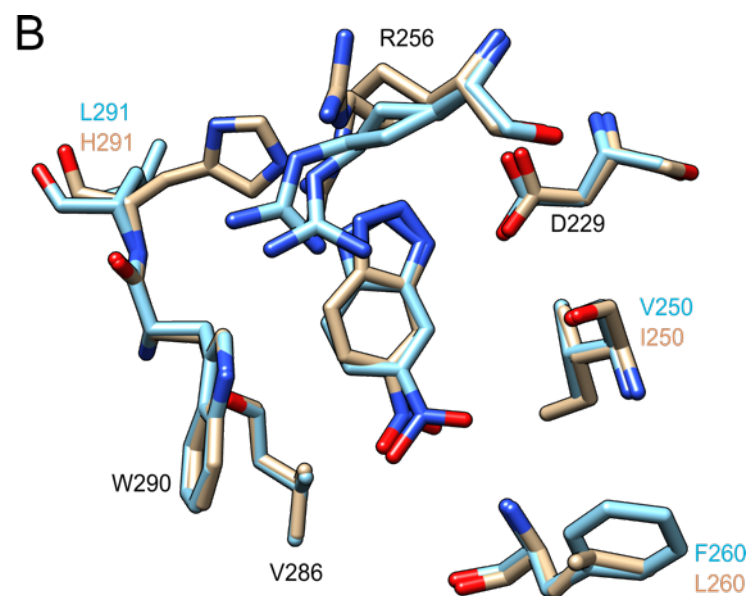
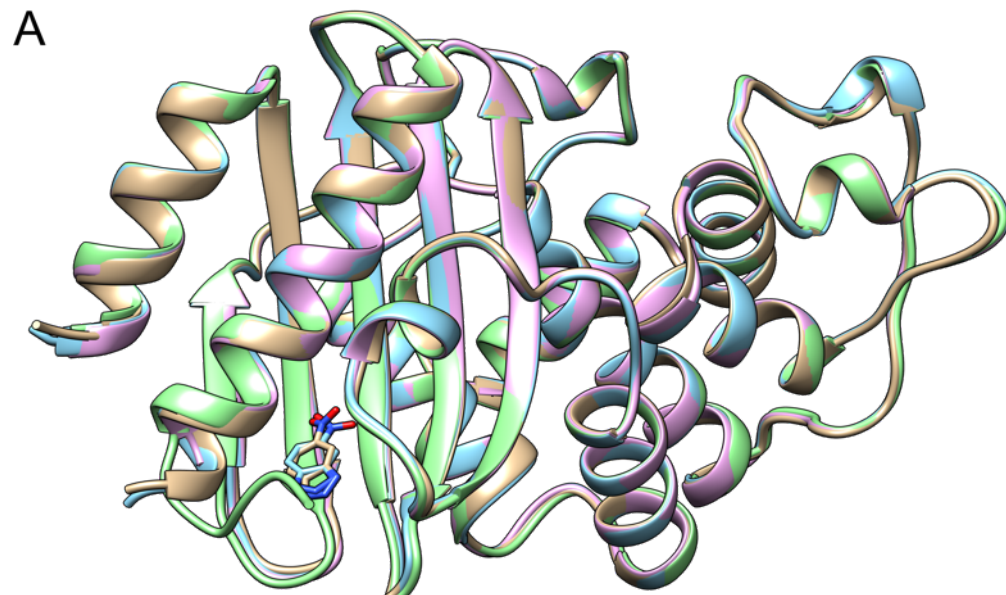
Activity Enhancement with FuncLib





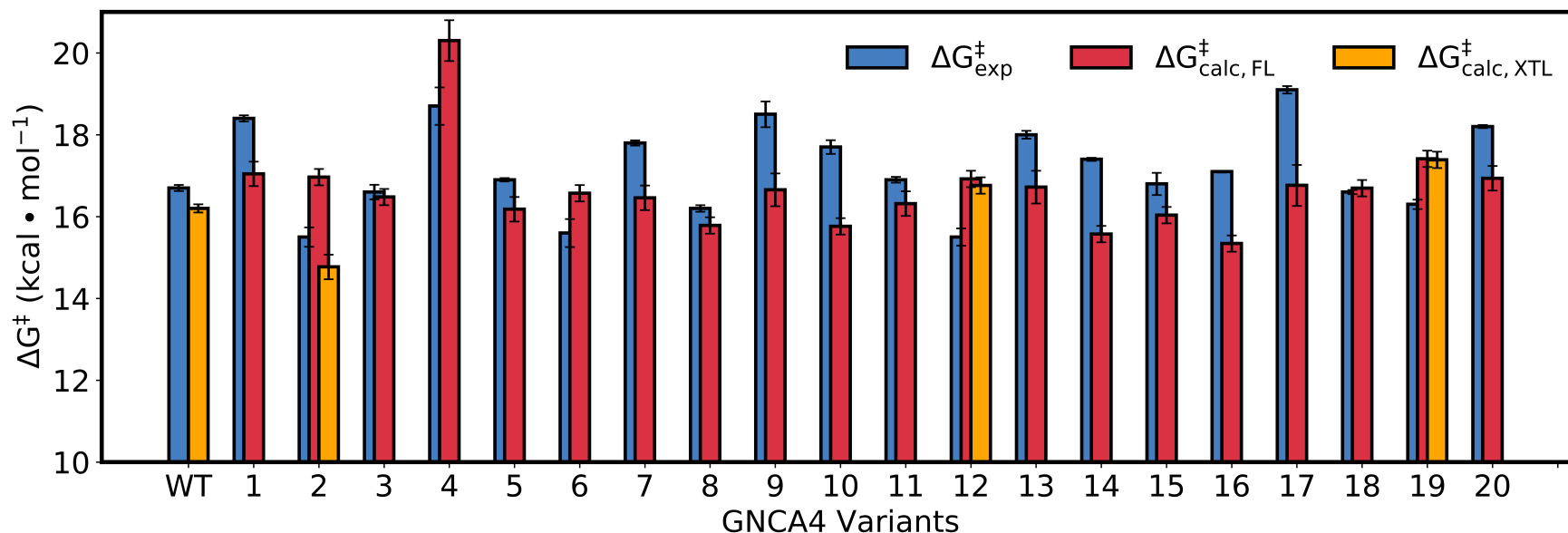
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Minimal Structural Changes





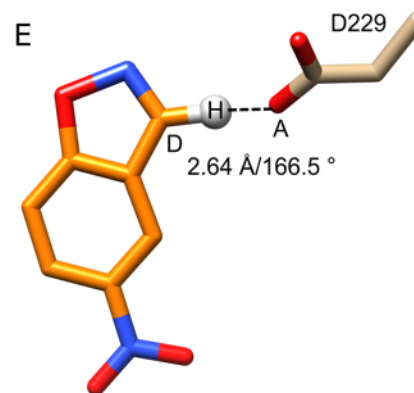
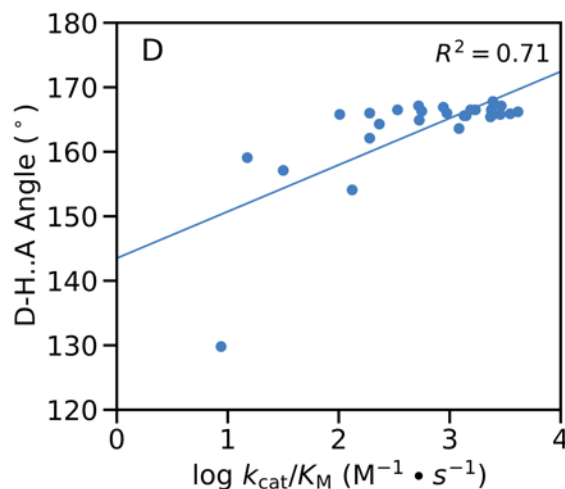
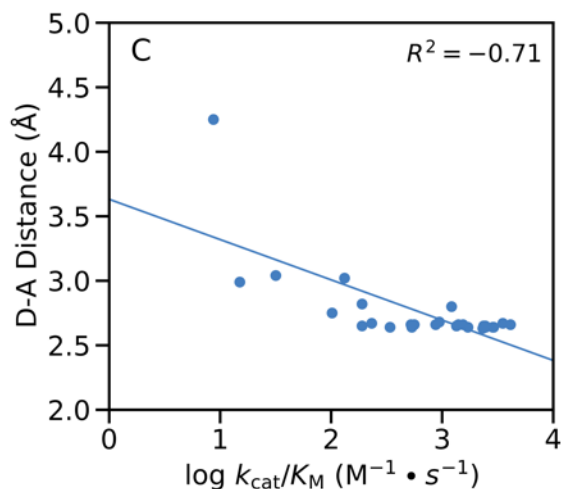
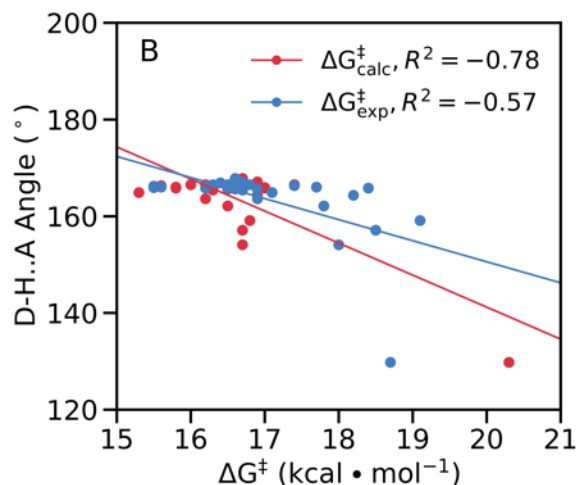
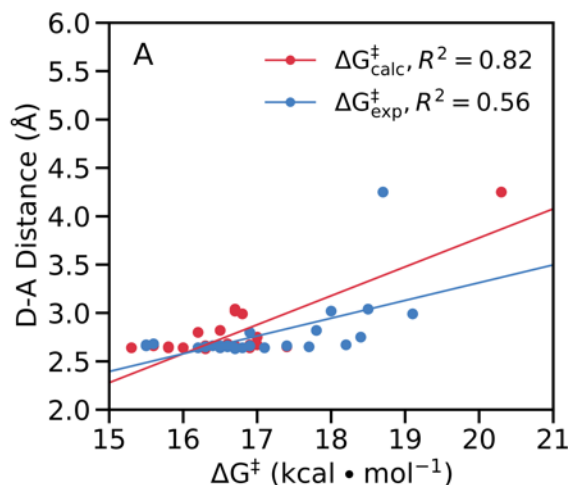
Can EVB Further Refine the Ranking?





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Geometric Preorganization and Activity





So What Drives Enzyme Evolution?

- Comparison of several enzymes shows strong correlation between the structural and electrostatic features of their active sites and variations in substrate selectivity.
- These enzymes don't know in advance what substrate will bind, but exploit conformational dynamics to adjust their active site environment to a given substrate after the binding step.
 - *Just having key catalytic residues in place is not enough!*
- Regulating both local and global conformational dynamics appears to be an important factor in allowing for the emergence of new enzyme activities.

Conformational dynamics needs to be accounted for in both experimental and computational protein engineering studies!



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Further Reading if Interested

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Viewpoint

Manipulating Conformational Dynamics To Repurpose Ancient Proteins for Modern Catalytic Functions

Jasmine M. Gardner, Michal Biler, Valeria A. Risso, Jose M. Sanchez-Ruiz,* and Shina C. L. Kamerlin*



Cite This: *ACS Catal.* 2020, 10, 4863–4870



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Perspective

Harnessing Conformational Plasticity to Generate Designer Enzymes

Rory M. Crean, Jasmine M. Gardner, and Shina C. L. Kamerlin*



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