

SMR 1329 - 1

**COLLEGE ON BIOPHYSICS:
FROM MOLECULAR GENETICS TO STRUCTURAL BIOLOGY**

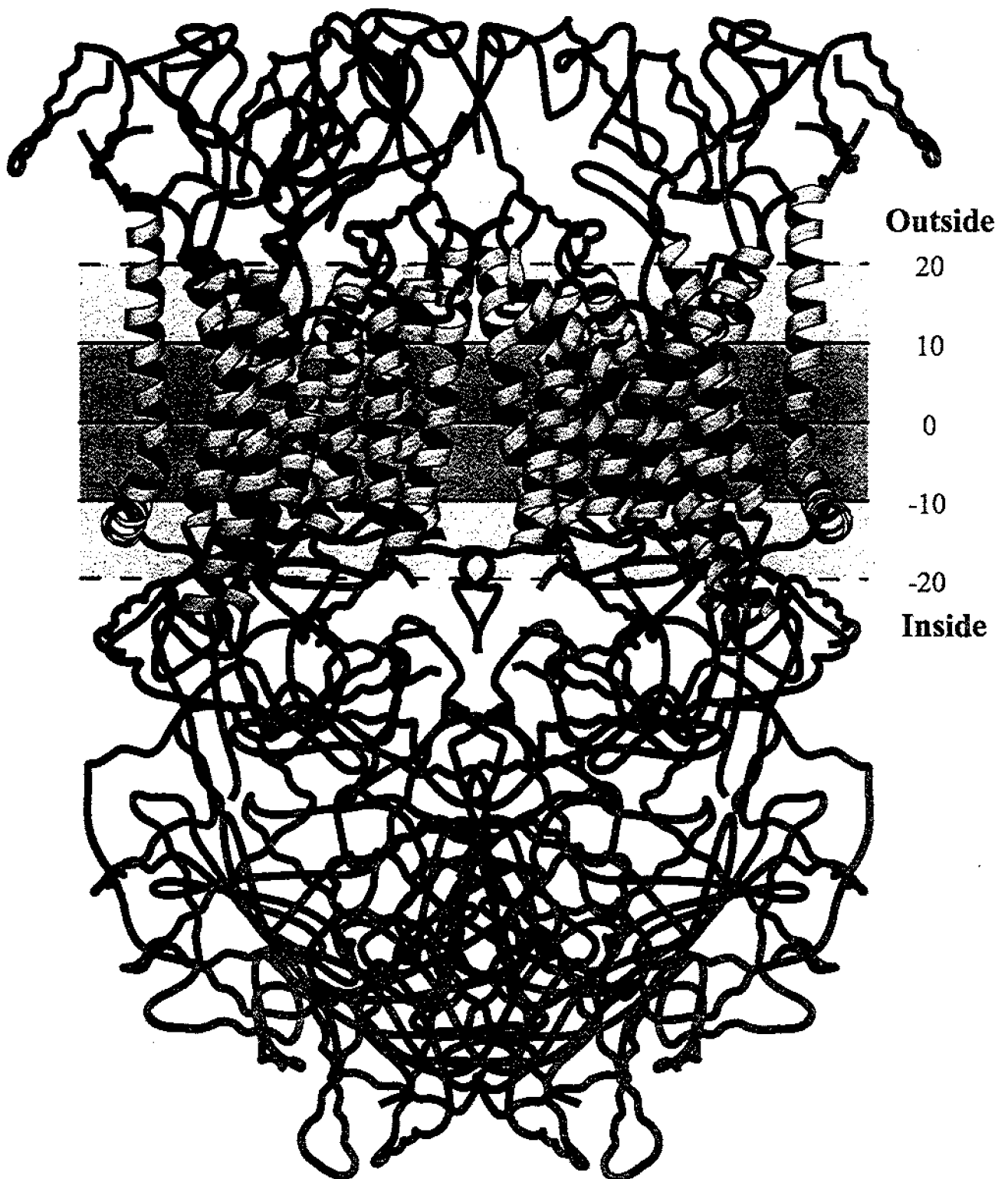
1 - 12 OCTOBER 2001

***PREDICTION OF TRANSMEMBRANE
PREOTEIN TOPOLOGY***

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These are preliminary lecture notes, intended only for distribution to participants.

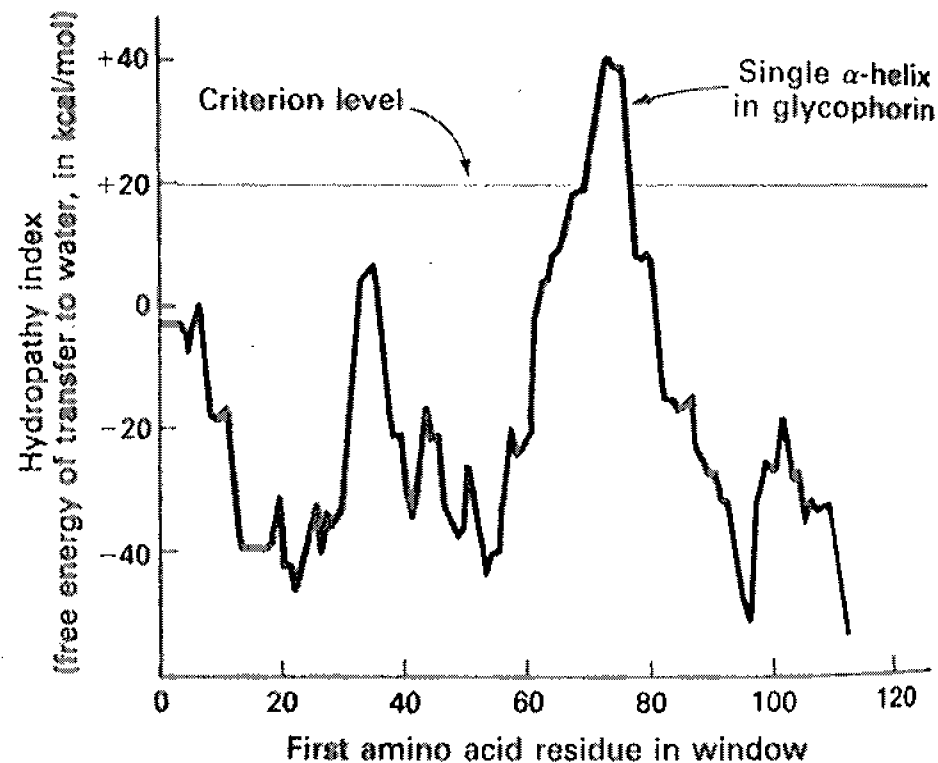
Cytochrome BC1 Complex (1bgy)

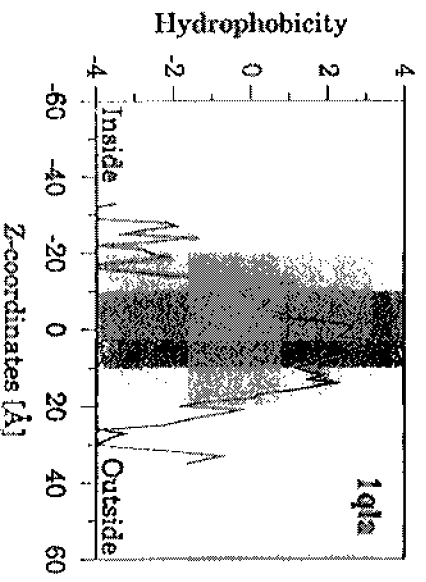
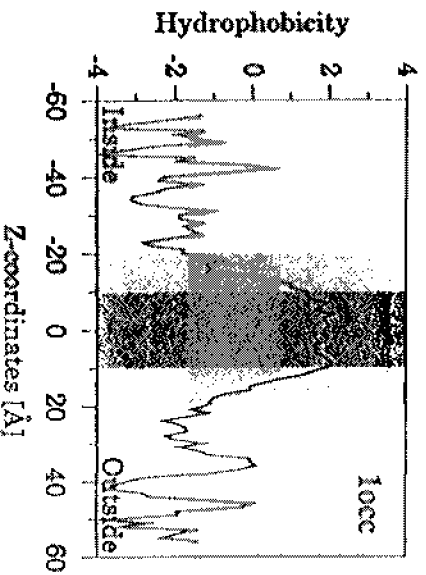
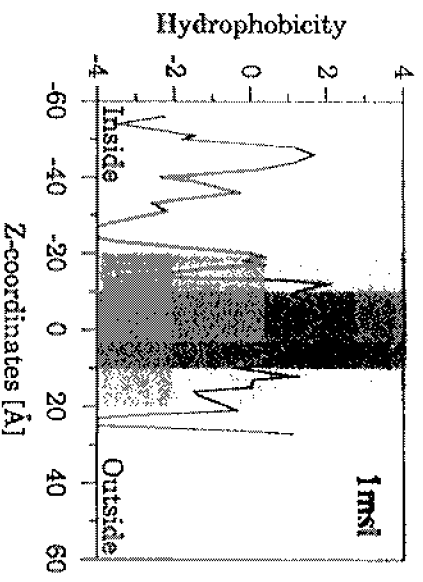
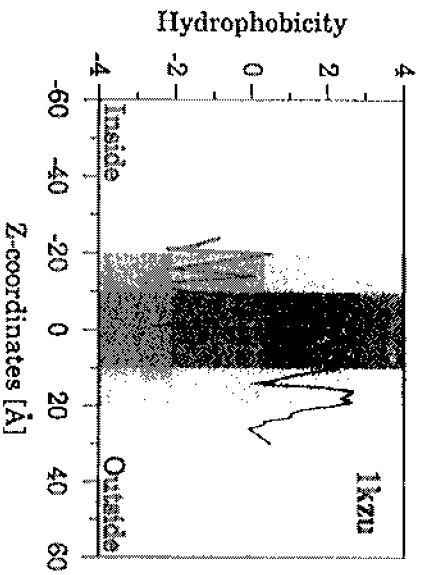
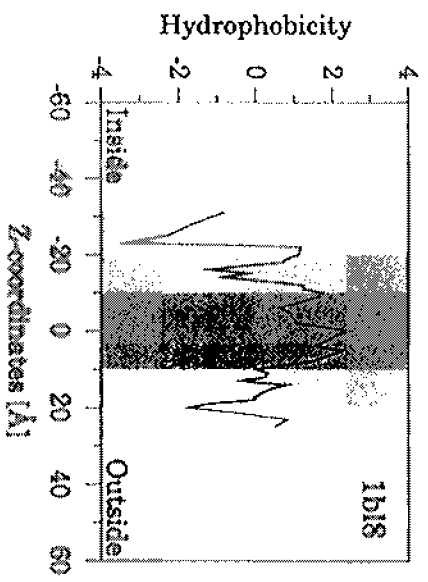
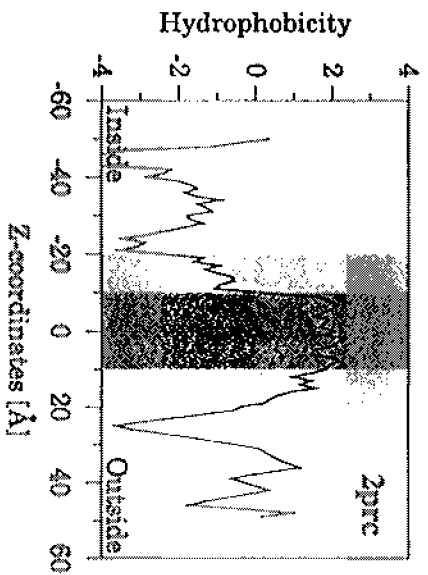
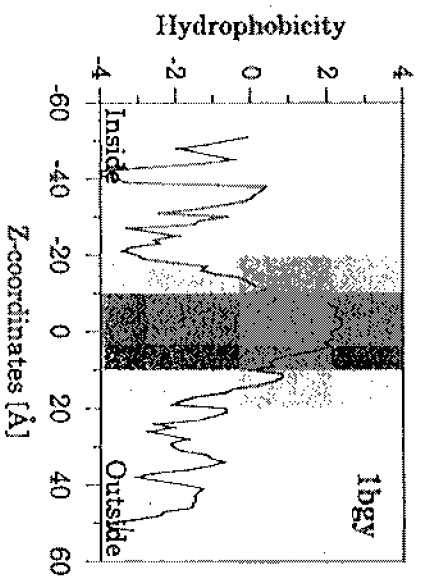
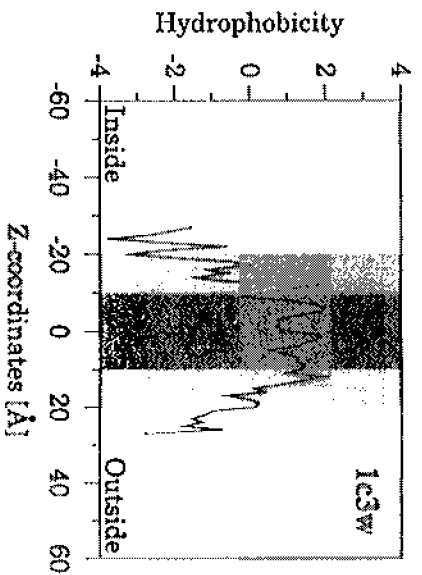


Polarity scale for identifying
transmembrane helices

Amino acid residue	Transfer free energy (kcal/mol)
Phe	3.7
Met	3.4
Ile	3.1
Leu	2.8
Val	2.6
Cys	2.0
Trp	1.9
Ala	1.6
Thr	1.2
Gly	1.0
Ser	0.6
Pro	-0.2
Tyr	-0.7
His	-3.0
Gln	-4.1
Asn	-4.8
Glu	-8.2
Lys	-8.8
Asp	-9.2
Arg	-12.3

Note: The free energies are for the transfer of an amino acid residue in an α helix from the membrane interior (assumed to have a dielectric constant of 2) to water. After D.M. Engelman, T.A. Steitz, and A. Goldman. *Ann. Rev. Biophys. Biophys. Chem.* 15(1986):330.





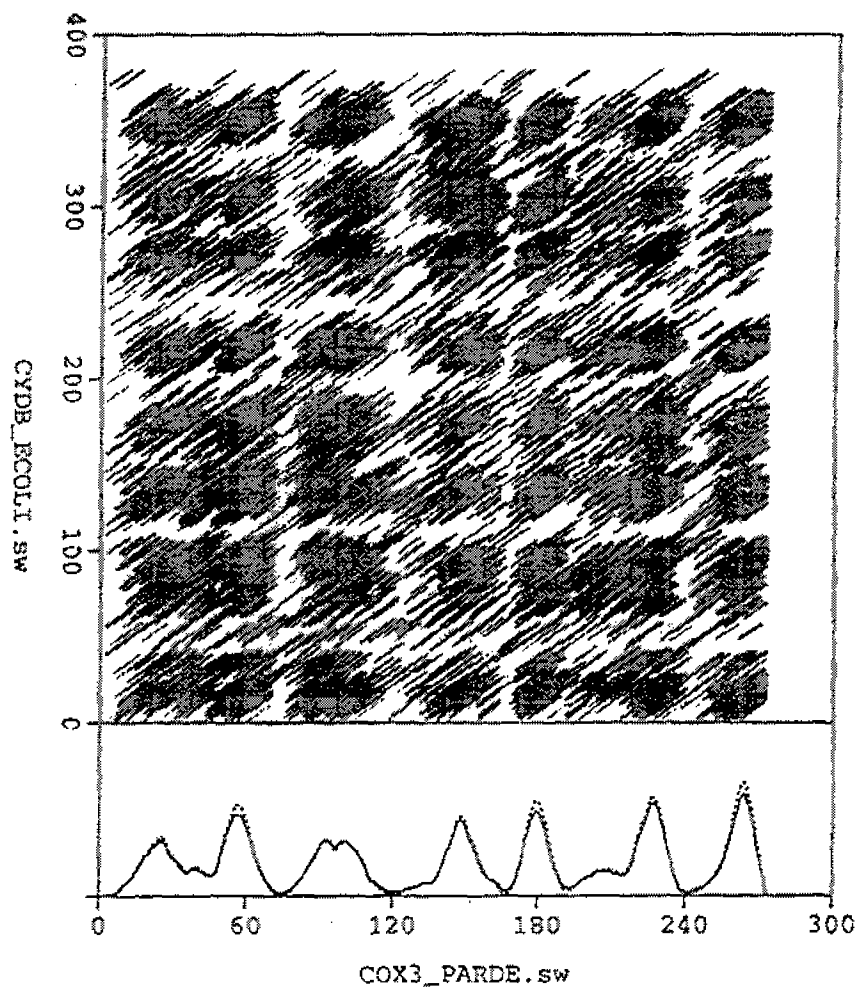
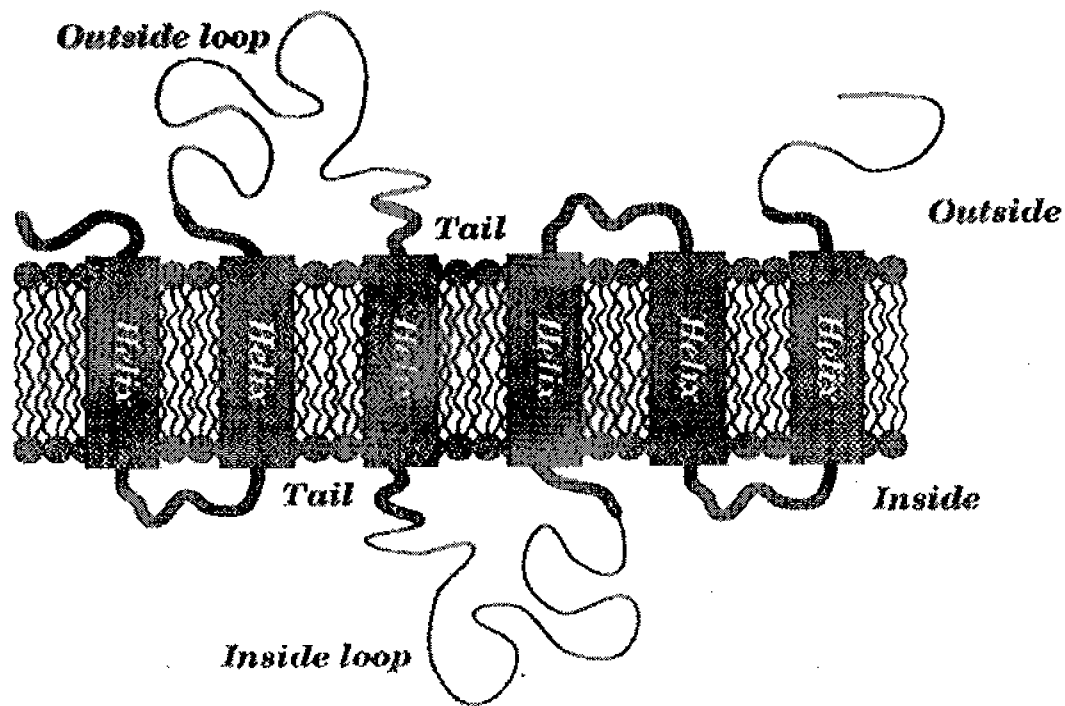


Fig. 1. DAS plot of two arbitrarily chosen proteins (COX3_PARDE versus CYDB_ECOLI). The cross weighted cumulative score profile (dotted line) and the global DAS profile (continuous line) calculated as the average of the cumulative score profiles obtained for comparisons with the other 43 proteins in the test set are also shown for COX3_PARDE. COX3_PARDE has seven and CYDB_ECOLI has eight transmembrane segments.

$$F = \sum_{i=1}^5 \sum_{j=1}^{20} q_{ij} \cdot \log \left(\frac{q_{ij}}{p_j} \right)$$

- where p_j is the frequency of the residue j in the whole protein;
- q_{ij} is the frequency of the residue j in the structural part i of the protein.



Amino acid seq: MGDVCDTEFGILVA...SVALRPRKHGRWIV...FWVDNGTEQ...PEHMTKLIHM...

State seq: oooooooooohhhhh...hhhhiiiiiihhhh...hhhooooOO...OOooooohhh...



Figure 2. Structural states defined for a typical helical transmembrane protein. The five states are: inside loop (l), inside tail (i), membrane helix (h), outside tail (o) and outside loop (O). Tails (thick lines) are thought to interact with the inside or outside parts of the membrane, while loops (thin lines) do not. Two tails between helices can form a short loop, but longer loops are formed by tail-loop-tail sequences.

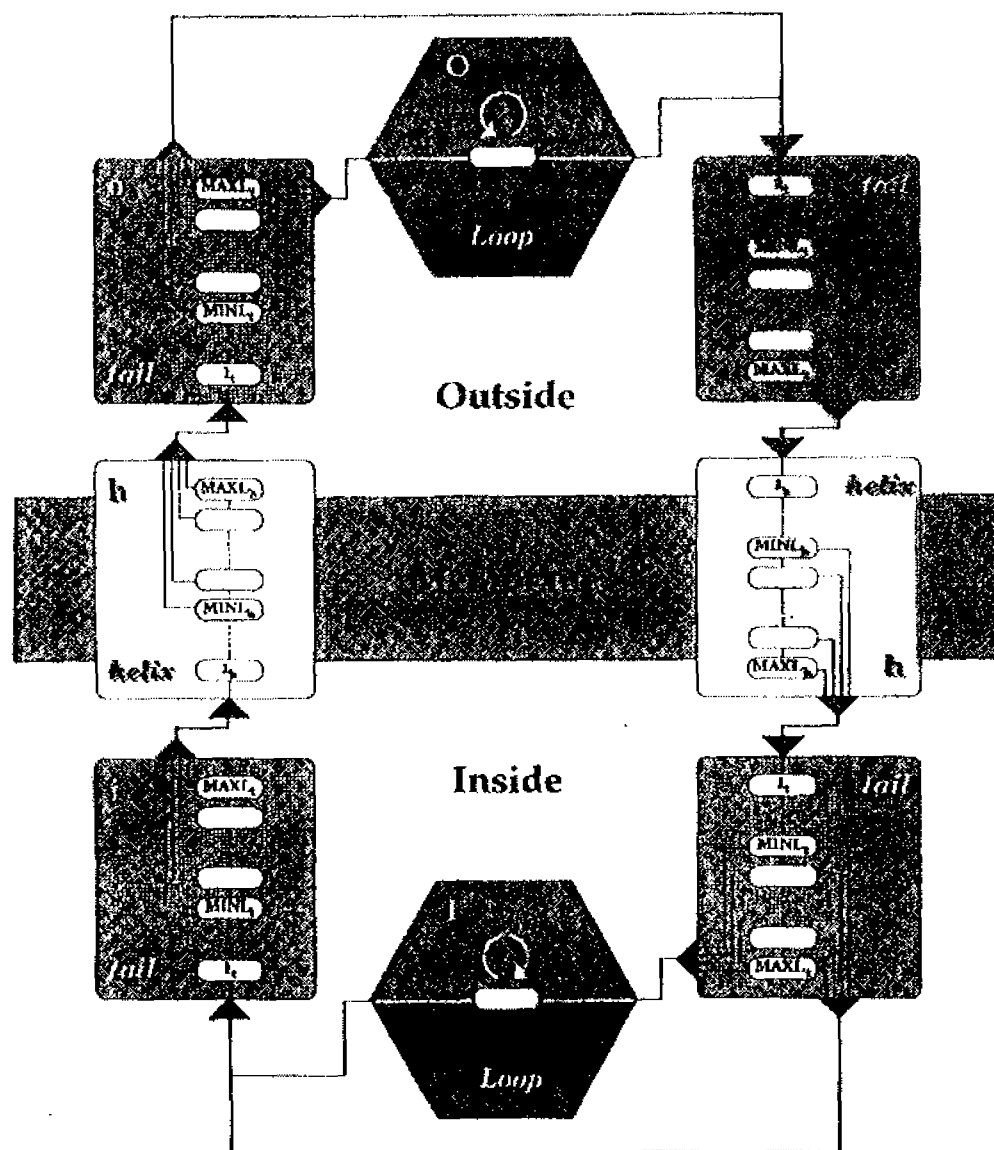


Figure 3. Architecture of HMM used for topology prediction. States with the same transition matrices are colored in the same way: white, helix states; light gray, tail states; dark gray, loop states. Rectangular areas FL type states; hexagonal ones, NFL type states. The observation-symbol probabilities used by states are marked in each state. The structure of substates in the case of the FL type is drawn within states. Lines and arrows show the possible transition between states or sub-states.

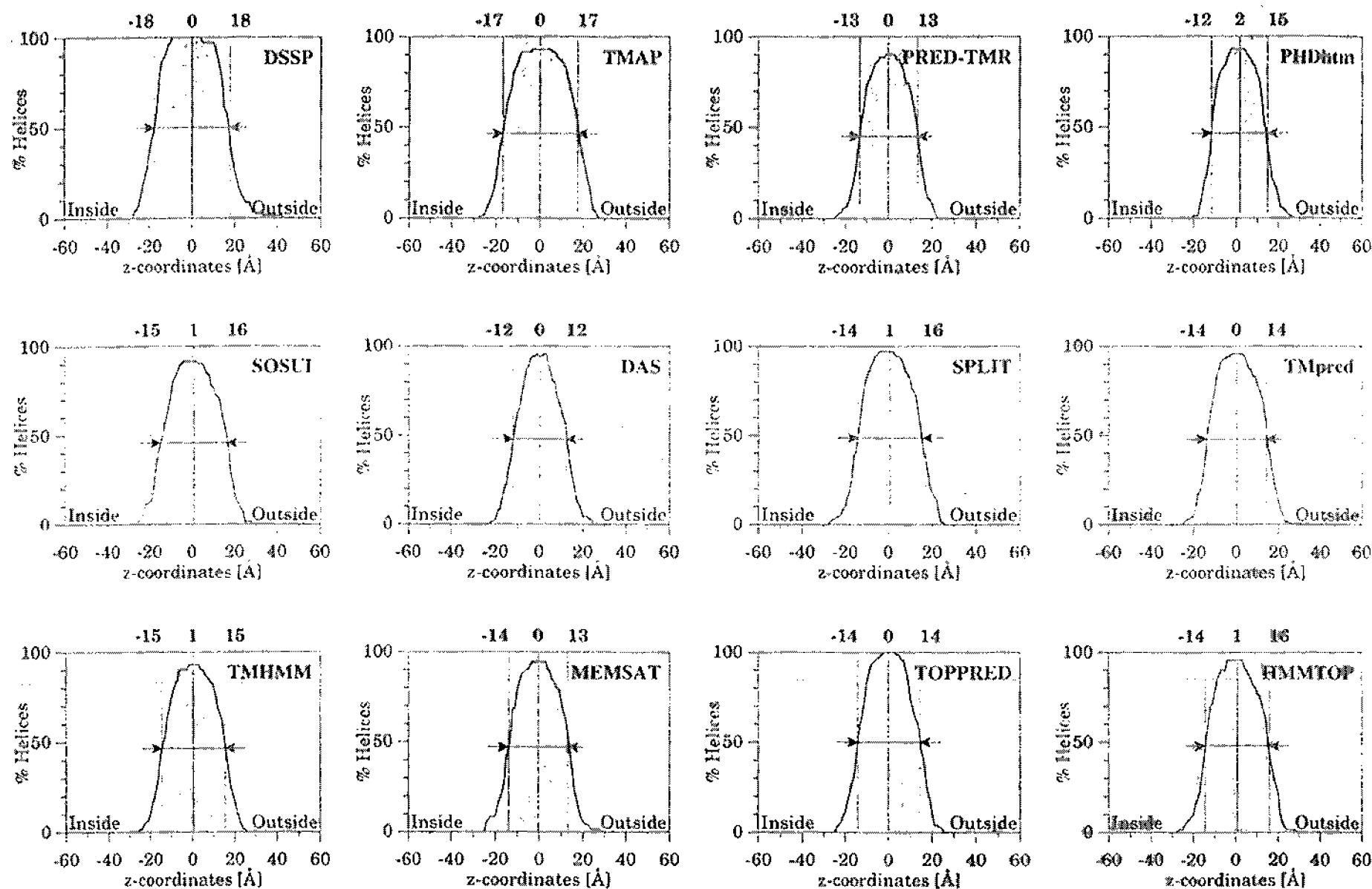
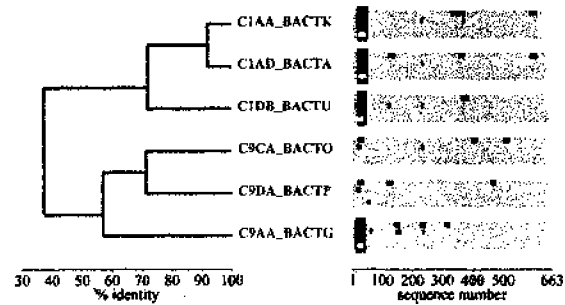


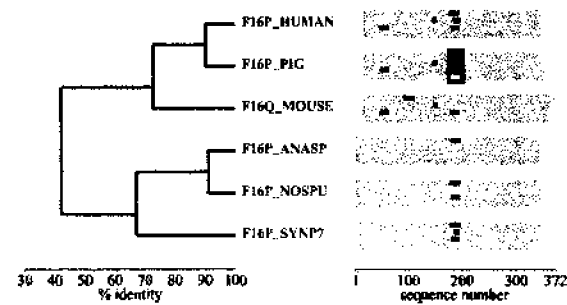
Figure 2. Distribution of transmembrane helices measured parallel to the average direction of the transmembrane helices in 1 Å slices in the eight selected proteins (see the text). Transmembrane helices are defined by the DSSP program and predicted by various transmembrane helix prediction methods. The z-coordinates at the value of 50% and at the mean of the curves are shown above the graphs. The gray areas show the same regions as in Figure 1.

Globular proteins

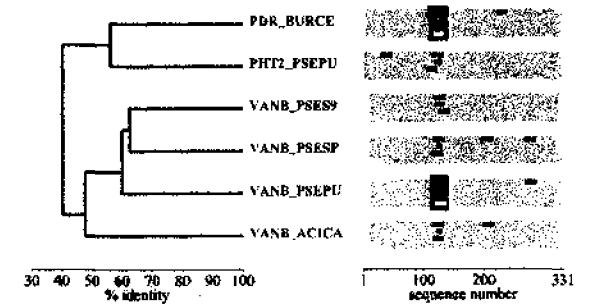
Insecticidal toxin (1c1y)



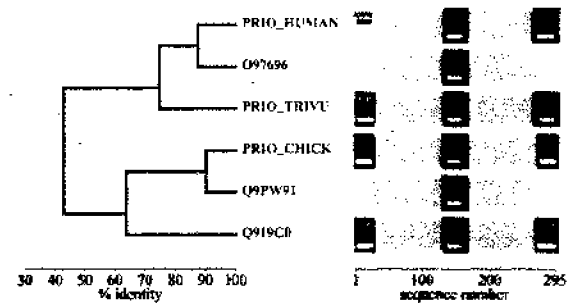
Fructose-1,6-bisphosphatase (1f1p)



Phthalate dioxygenase reductase (2p1a)

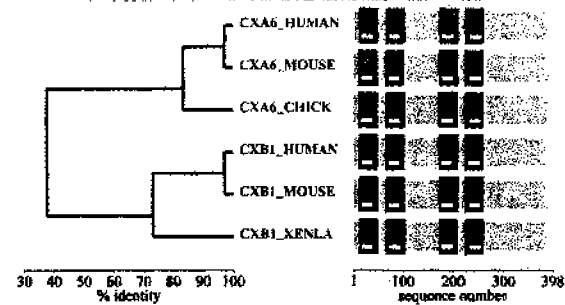


Prion proteins

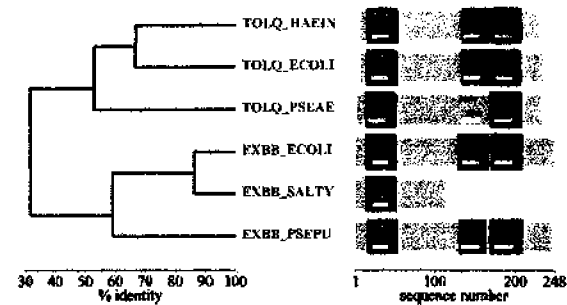


Transmembrane proteins

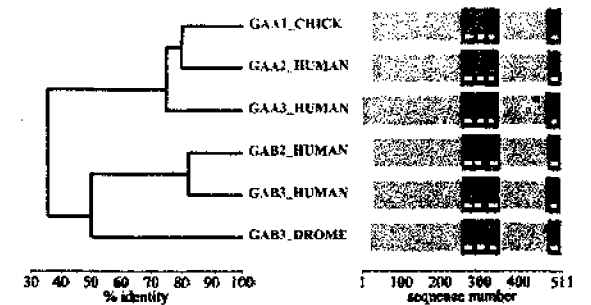
Gap junction β -1 protein

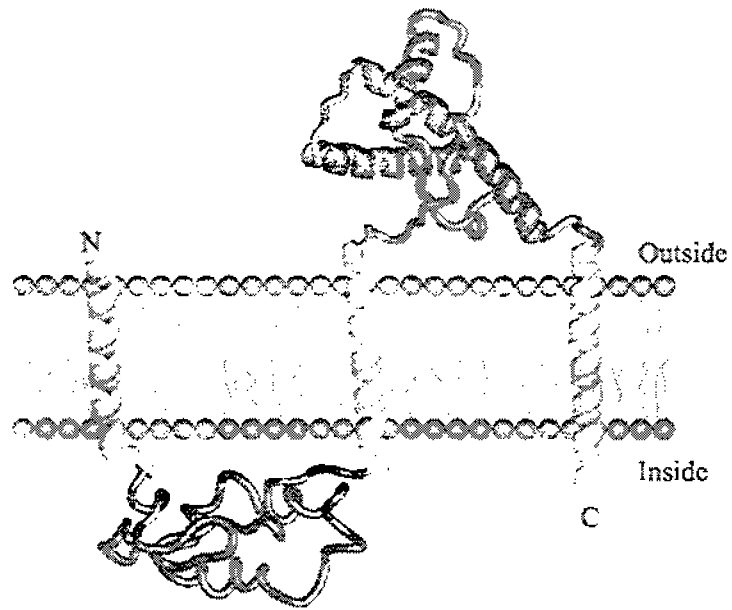


Biopolymer transport protein

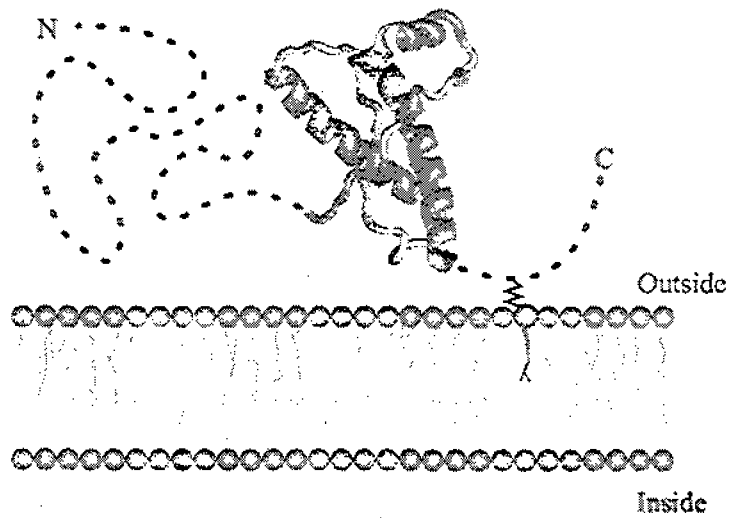
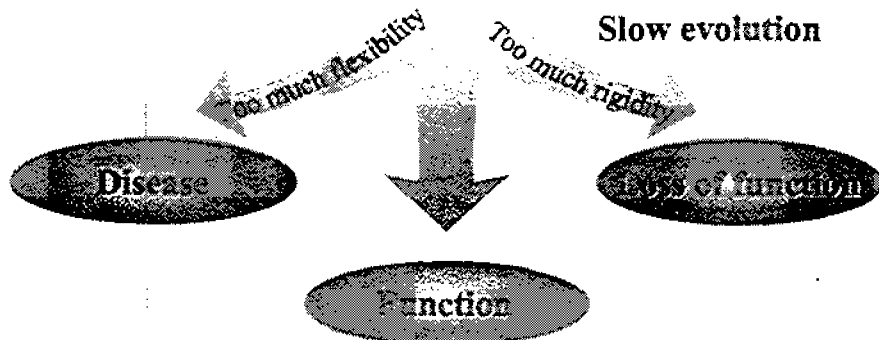


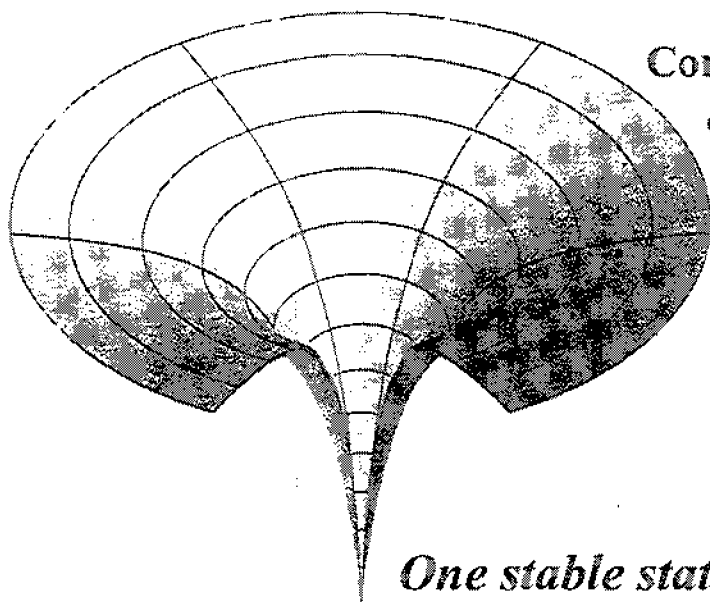
Gamma-aminobutyric-acid receptor



A

Mutation and
rapid evolution

B**C**



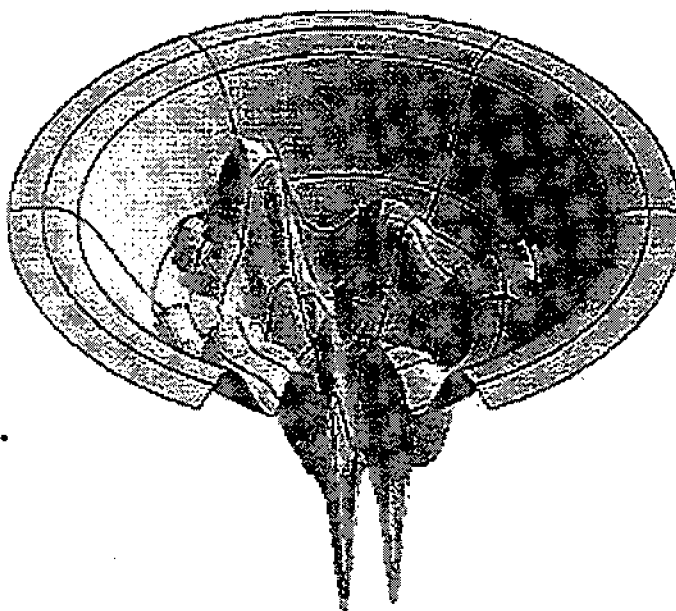
Common energy landscape
of a globular or a trans-
membrane protein.

One stable state



Cellular relocalization

Energy landscape
corresponding to
the transient state
from transmembrane
protein to globular one.



Two (or more) stable state