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UNITED NATIONS EDUCATIONAL, SCIENTIFIC AND CULTURAL ORGANIZATION



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SMR/302-25

COLLEGE ON NEUROPHYSICS:
"DEVELOPMENT AND ORGANIZATION OF THE BRAIN"
7 November - 2 December 1988

"Modelling the Retina"

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Please note: These are preliminary notes intended for internal distribution only.

MODELLING THE RETINA

3.

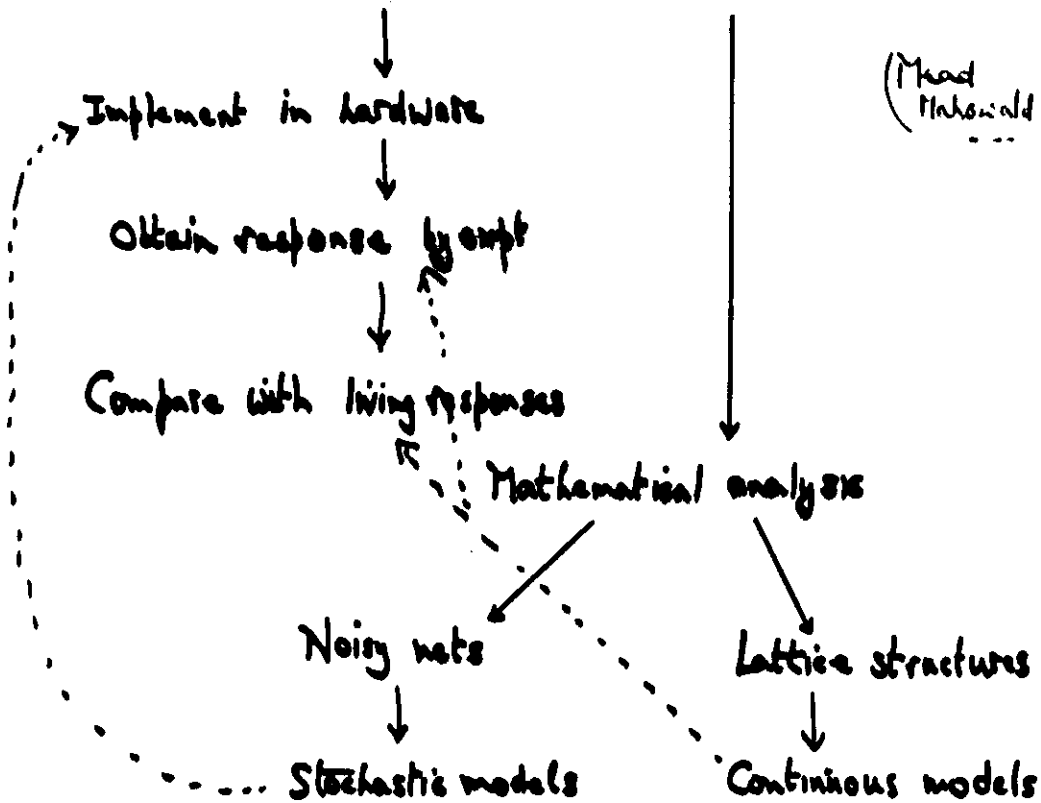
Contents of talk:

2.

- 1) Invertebrate retina (lateral inhibition)
- 2) Math. models of "
- 3) Vertebrate retina
- 4) Math. models of "

WHY → UNDERSTAND LIVING SYSTEM (→ brain?)
→ BUILD HARDWARE REALISATION (→ connectionism)

HOW → CIRCUIT DIAGRAMS OF (L,R,C) CIRCUITS

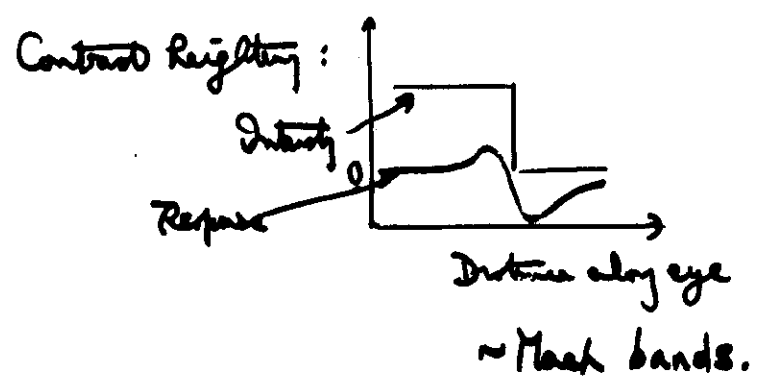
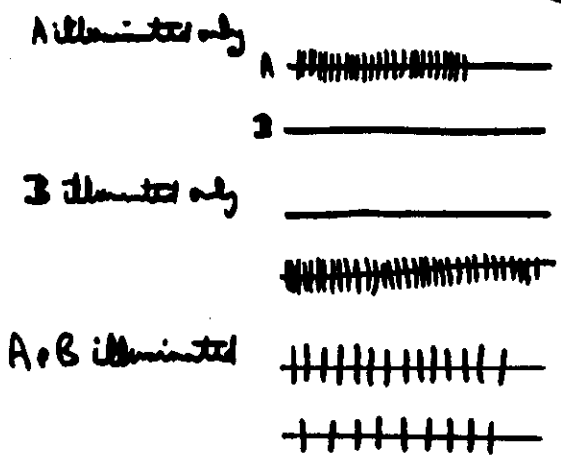
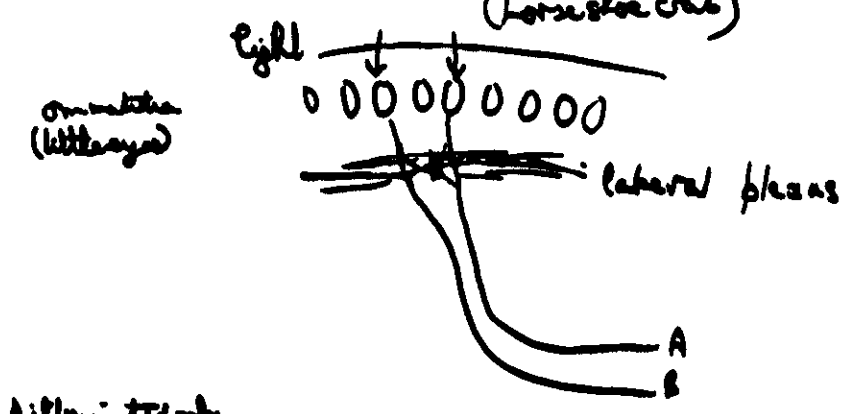


Mathematical analysis

- easier (no hardware to build)

2. Contrast Rejection:

Mutual inhibition in the compound eye of Limulus (horseshoe crab)



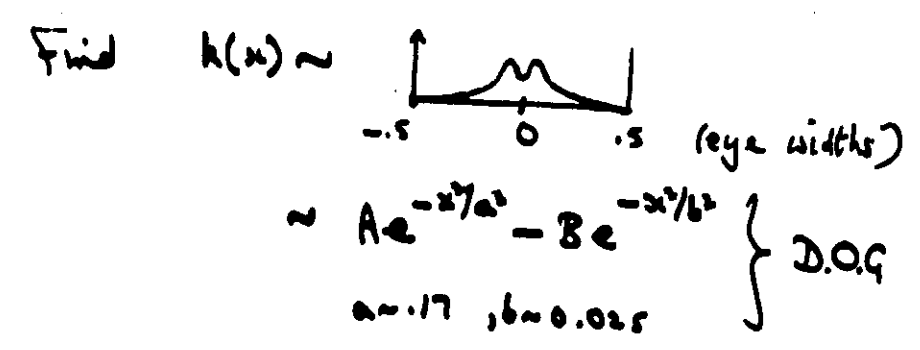
3

Use sine waves
in t and x , so

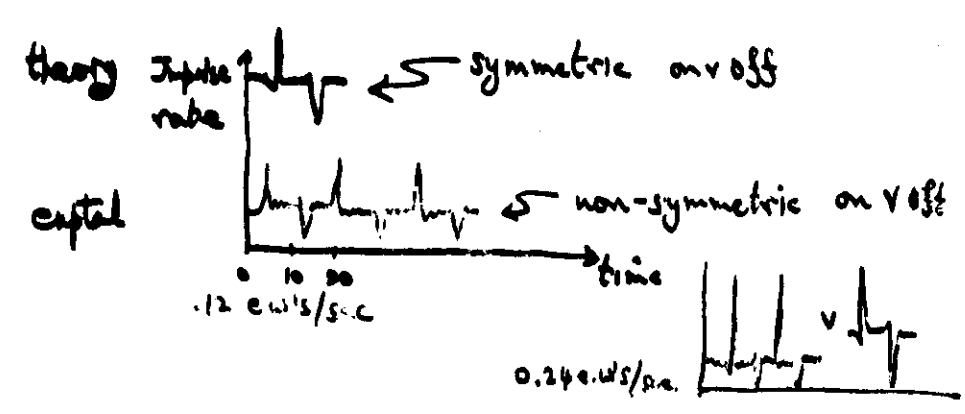
6.
(→9)

$$\tilde{T}(x, \omega) = \tilde{T}(x, \omega) \tilde{I}(x, \omega)$$

$$\tilde{T} = \frac{E(\omega) G(\omega)}{1 + E(\omega) T(\omega) K(x)}$$



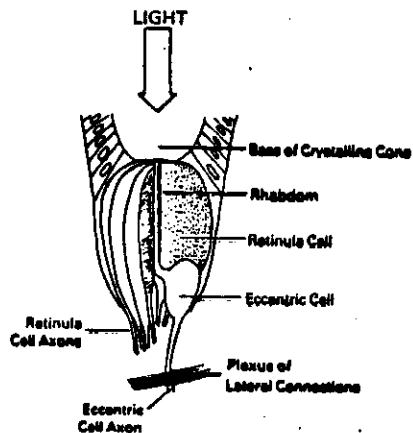
Gives response to moving square-wave stimuli:



A pRAM APPROACH TO VISUAL PROCESSING IN LIMULUS

(Goss and Taylor, 1988)

9.



Hartline-Ratliff equations (in 1-D) (1963):

$$x_i(t) = [e_i(t) - \sum_{j \neq i} K_{ij} x_j(t)] + \text{excitation via log I}$$

inhibition due to jth neighbor

- system characterised by fixed point (solution of n linear simultaneous equations)
- uses step function

(from Y. Bruce & P. Green, 'Visual Perception', Erlbaum (1985))

Brodie et al (1978): 2D, continuous in space/time. Good fit to data, but still uses step function and 'self-inhibition' (not understood)

pRAM model

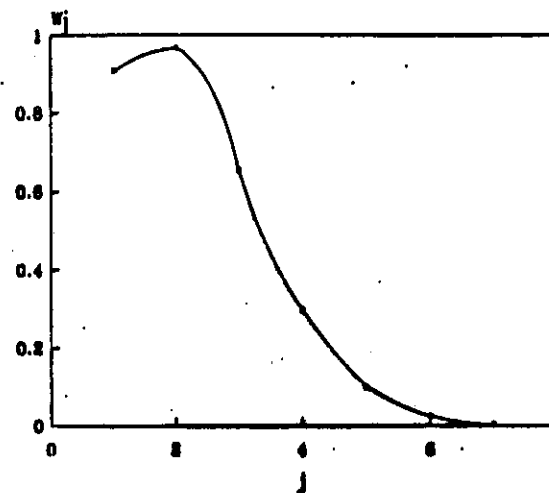
$$x_i(t) = K^{-1} \prod_{s=1}^T (1 + w_s r_i(t-s)) \times \prod_{j=1}^N (1 - W_j x_{i-j}(t-j-d)) (1 - W_j x_{i+j}(t-j-d))$$

1-D array, each unit receives T time-delayed illumination inputs $\{r_i(t-1), \dots, r_i(t-T)\}$ & inhibitory inputs $x_{i \pm j}(t-j-d)$ from its 2N nearest neighbours

$$K = \prod_{s=1}^{d+1} (1 + w_s) \quad \text{normalisation factor (so that } x_i \in [0,1])$$

(d+1) = number of positive w_s
max/min response (d+1) time units after illumination change

10.



Lateral inhibition falls off with distance of inhibiting unit

DOG form (taken from Brodie et al):

$$W_j = \frac{3}{4} [3e^{-j^2/8} - 2e^{-j^2/4}]$$

Effect of illumination initially excitatory (d+1=3 time steps), then inhibitory

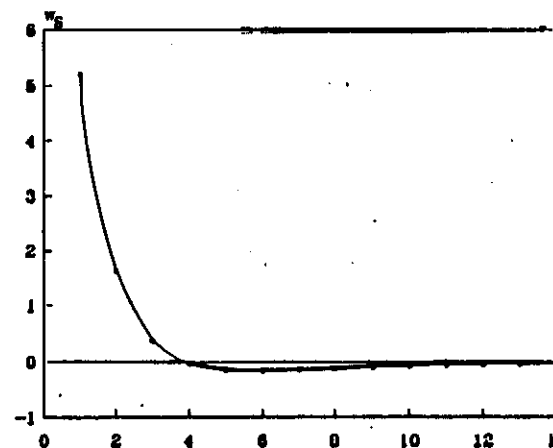


Alternative is self-inhibition (Stephens 1964)

- less pronounced transients
- oscillations

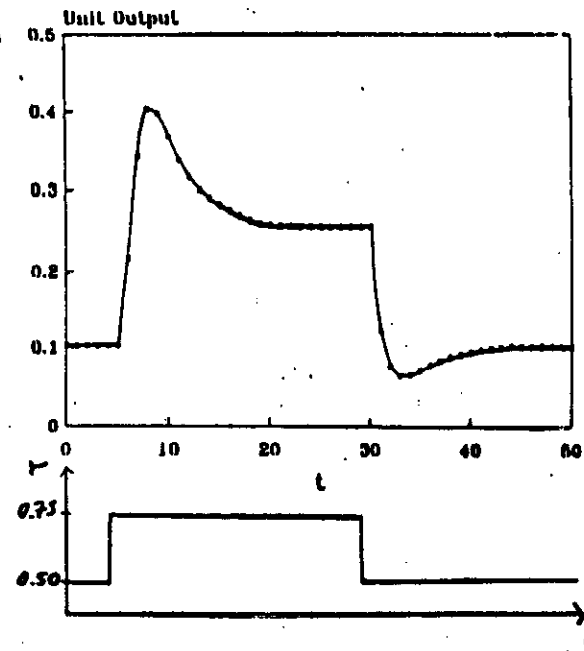


- too many back-connections for hardware implementation
- evidence against (Fahrenbach 1985)

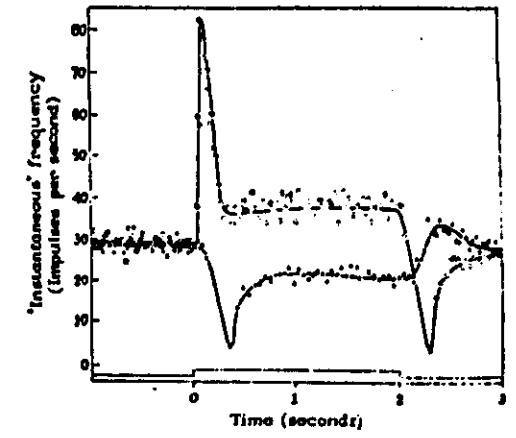
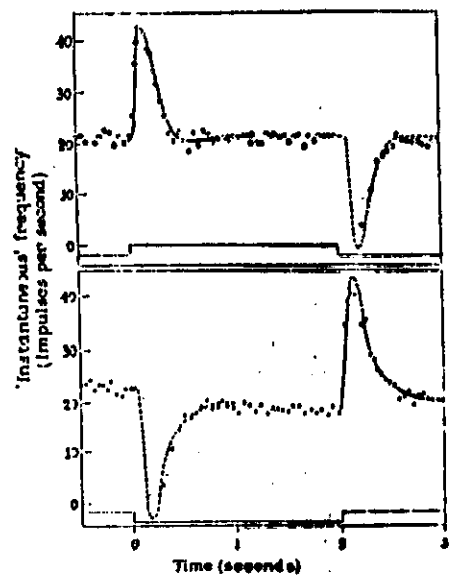
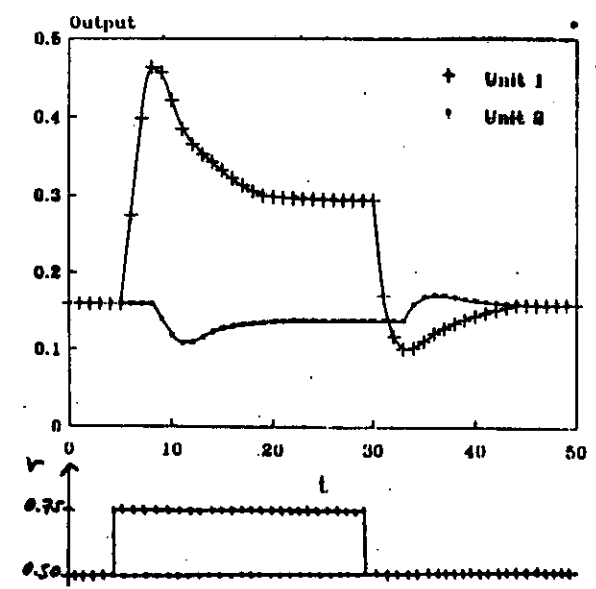


$$w_j = 16e^{-j^2/8} - \frac{3}{8}e^{-j^2/4}$$

convergence if $|x_i(t) - x_i(t-j)| < 0.0001$ $i=1..64$
 $j=1..8$

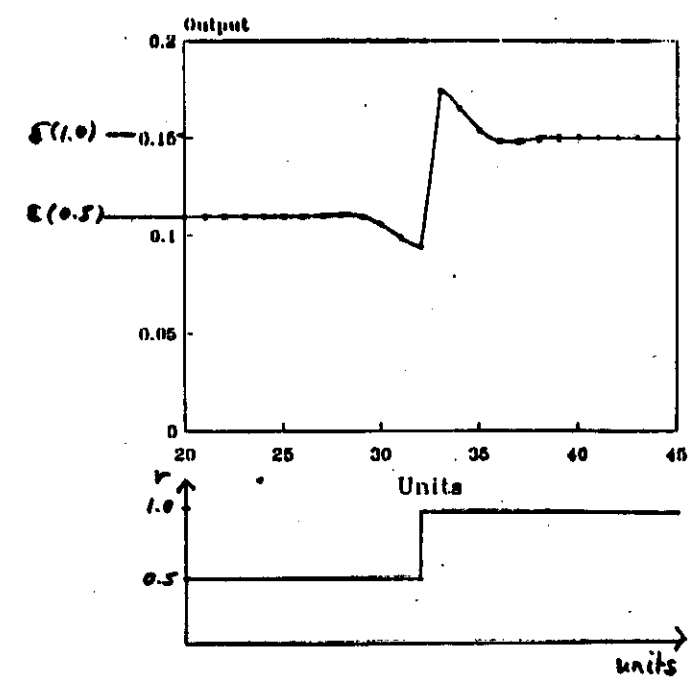


Temporal response of model

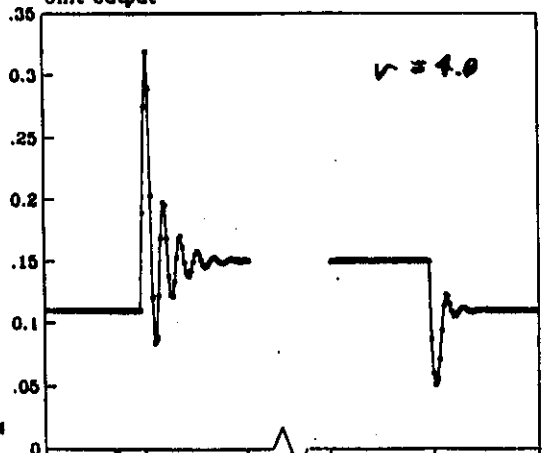
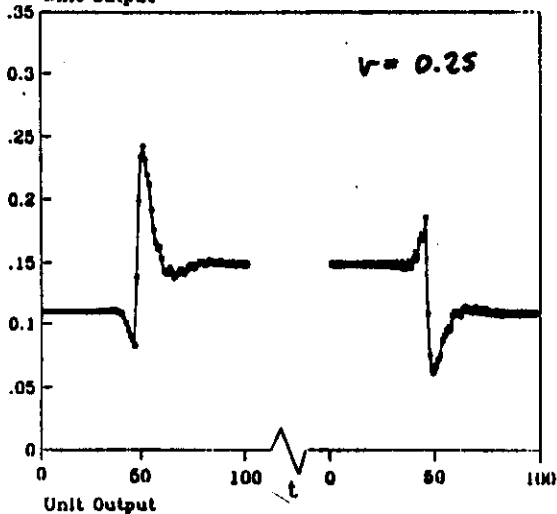
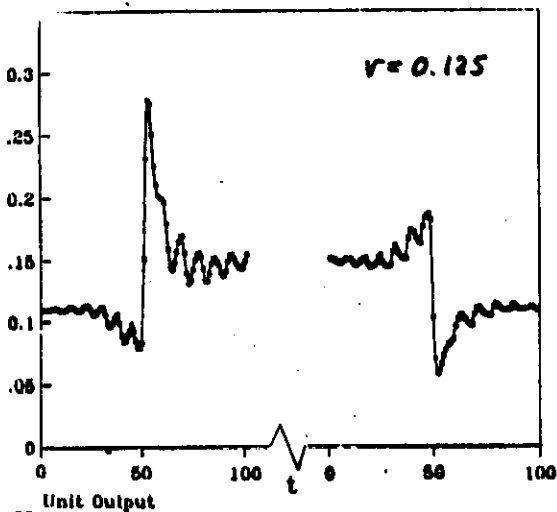


F. Ratcliff, H.K. Hartline &
 W.H. Miller, J. Opt. Soc. Am. 53 (1)
 (1963), 110

Spatial response (Mach bands)



Enhancement ~ 50% of $E(1.0) - E(0.5)$



Spatiotemporal behaviour 19.

Response of central unit as illumination step $r = 0.5 \rightarrow 1.0$ passes over it

Low velocity (retinal units/time step)

Anticipatory Mach bands contain oscillations due to neighbour-neighbour (second order) interactions

Intermediate velocity:

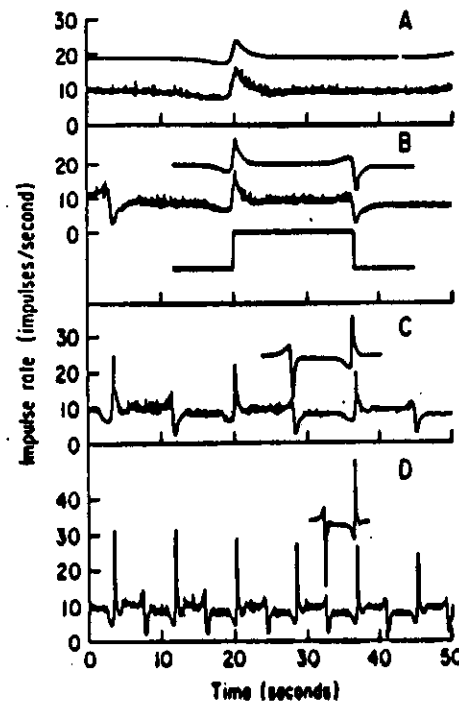
Inhibition from neighbours, but more distant ones now have no time to contribute inhibitory surges

High velocity ($v > 1$):

No time for anticipatory inhibition to be felt before maximum associated with the arrival of the step

Damped oscillations arise from time-delayed lateral inhibition from excited neighbours

Data from S.E. Brodie, B.W. Knight & F. Ratliff, J. Gen. Physiol. 72 (1978), 129



velocities in eyewndths per second

$u = 0.03$

$u = 0.06$

$u = 0.12$

$u = 0.24$

Data comparable with PRAM-net results for intermediate velocities (Brodie model predicts on- and off-transients of equal magnitude)

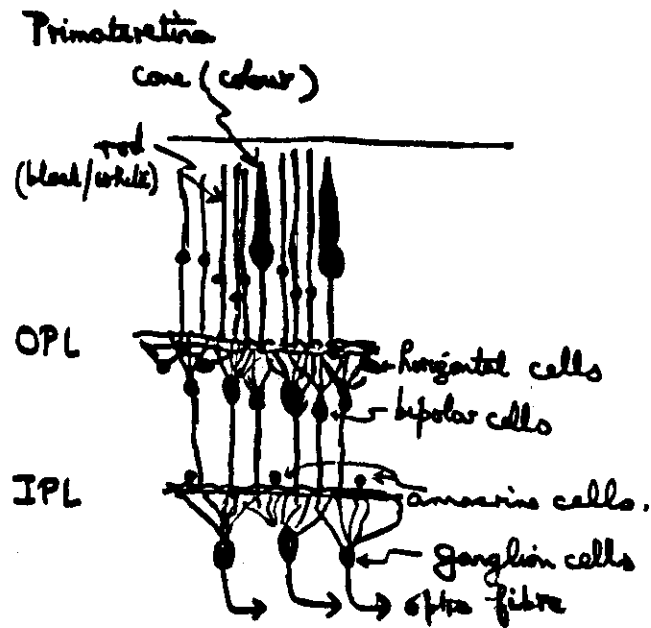
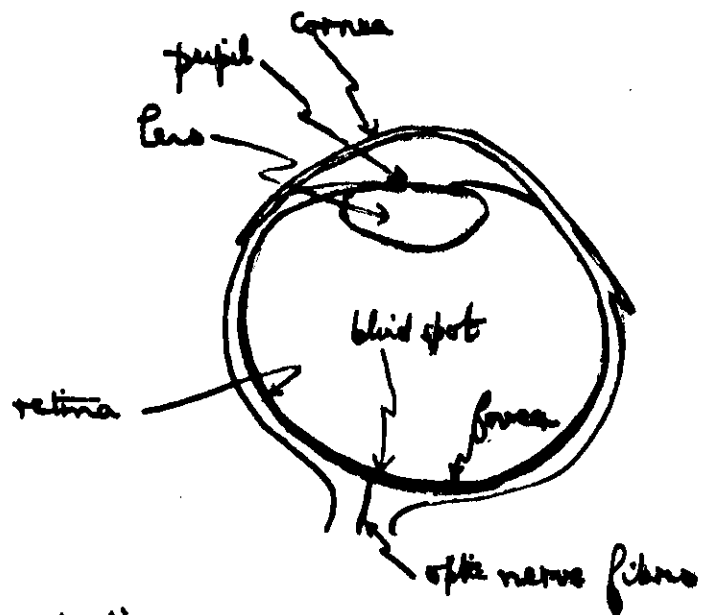
Low velocities: discrepancy could be due to the small no. of units used in the simulation

High velocities: finite speed of information transmission ($v=1$) - anticipatory Mach bands impossible for $v > 1$

$v=1$ vs $u = 2-20$

(78x faster than max velocity so far used)

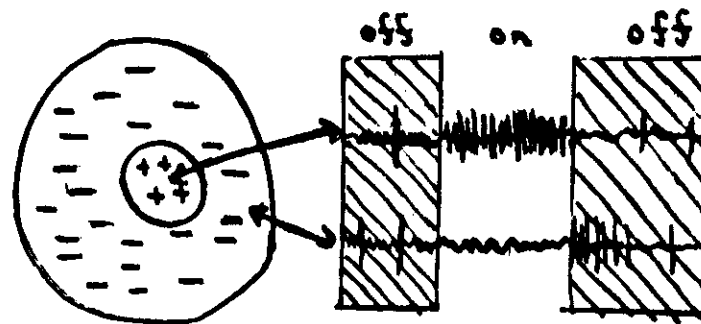
3 Vertebrate Retina.



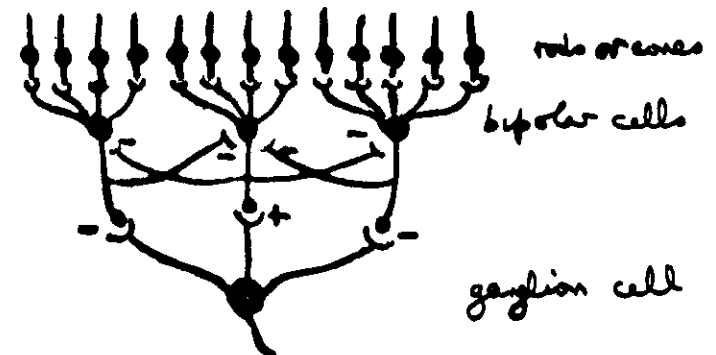
↑ incident light

16

Receptive field of ganglion cell:



On-centre off-surround
Also have off-centre on-surround
Neuronal connections to achieve this:

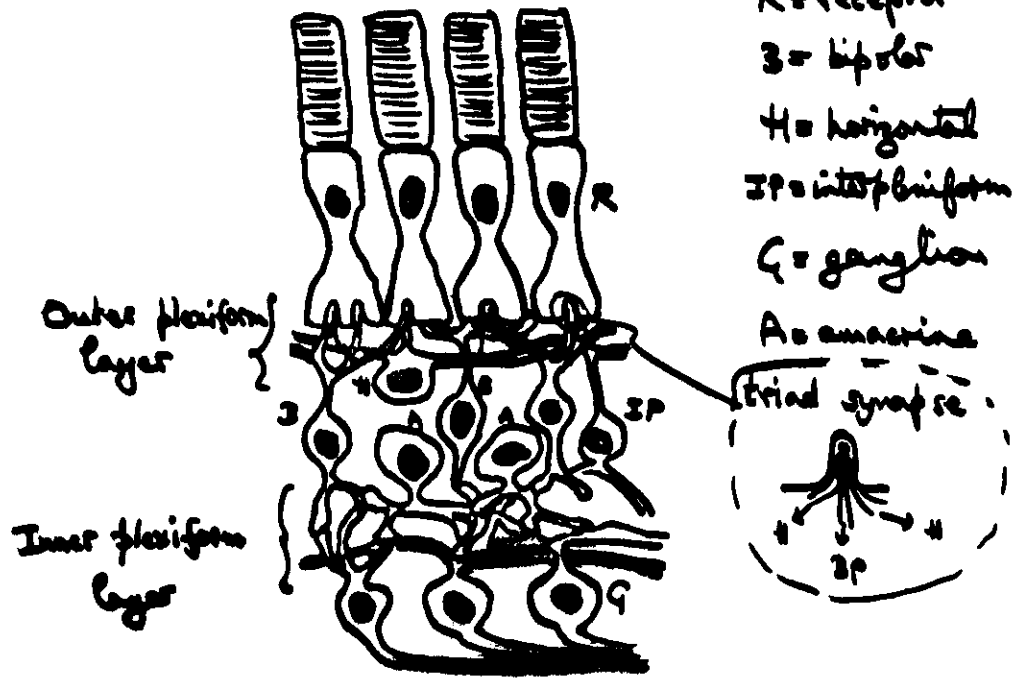


But (a) what are horizontal cells + amacrine cells for?

(b) can have more complicated ganglion cell responses - 'moving spot' detectors
(c) need to describe quantitatively.

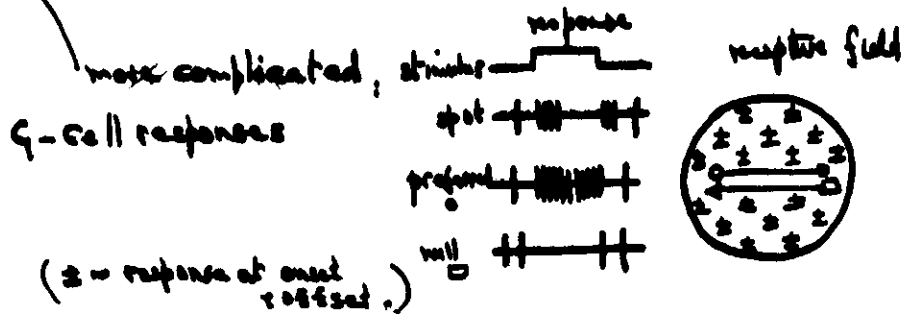
17

More detailed structure of retina:



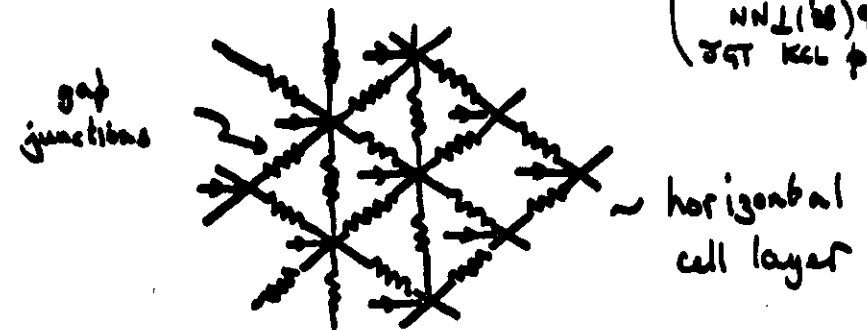
Frog, pigeons : Bipolar → Amacrine → Ganglion

Perinates : Bipolar → Ganglion

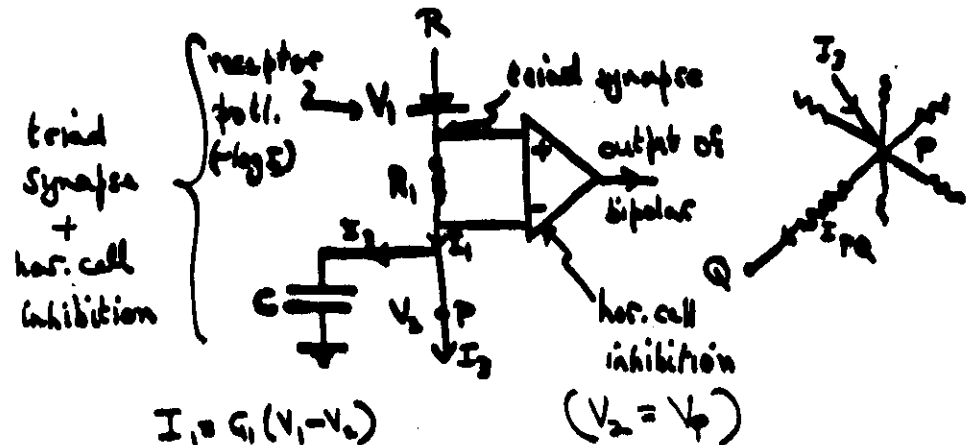


18. 9. Mathematical Modelling Attempt to model

hor. cell layer by
resistive network (are connected electrically)



(Mandelstam & Pashenvald
NNL (1988) 91
JGT KCL proposed)

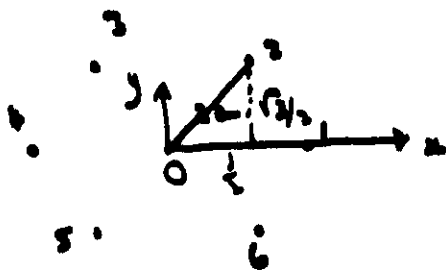


$$I_1 = G_1(V_1 - V_0)$$

$$I_2 = C \dot{V}_2$$

$$I_3 = \sum_{(Q,P)} I_{PQ} = \sum_{(Q,P)} G_2(V_P - V_Q) = I_1 - I_2$$

$$\Rightarrow G_1[V_1(P) - V(P)] - C \dot{V}(P) = G_2[NV(P) - \sum_{(Q,P)} V(Q)]$$



For a small, $V(1) - V(0) = aV_x + \frac{1}{2}a^2V_{xx}$

$V(2) - V(0) = \frac{1}{2}aV_x + \frac{\sqrt{3}}{2}aV_y + \frac{1}{2}a^2\left(\frac{1}{4}V_{xx} + \frac{\sqrt{3}}{4}V_{xy} + \frac{3}{4}V_{yy}\right)$

$V(3) - V(0) = - + -$

$V(4) - V(0) = -aV_x + \frac{1}{2}a^2V_{xx}$

$V(5) - V(0) = - - - (+)$

$V(6) - V(0) = + - - (-)$

$$\sum V(0) - 6V(0) = 2\pi^2(V_{xx} + V_{yy}) + O(a^4)$$

$$\Rightarrow \dot{V} + (G_1/C)V - (3G_1 a^2/2C)\nabla^2 V = (G_1/C)V_1$$

Use F.T. $\tilde{V} = \int dt d^2x e^{-i(\omega t + \mathbf{k} \cdot \mathbf{x})} V(\mathbf{x}, t)$

$$\Rightarrow \tilde{V} = [(i\omega C/G_1) + 1 + k^2(G_1 a^2/G_1)]^{-1} \tilde{V}_1$$

$$\Rightarrow V(\mathbf{x}, t) = (2\pi)^{-2} \int d^2k d\omega [(i\omega C/G_1) + 1 + k^2\lambda^2]^{-1} \tilde{V}_1(\mathbf{k}, \omega) e^{i(\mathbf{k} \cdot \mathbf{x} + \omega t)}$$

with $\lambda^2 = (G_1 a^2/G_1)$

Take small spots: $\tilde{V}_1 = \xi(\omega) e^{-k^2 d^2}$

$V_1(\mathbf{x}) \sim e^{-r^2/d^2}$ size $\sim d$.

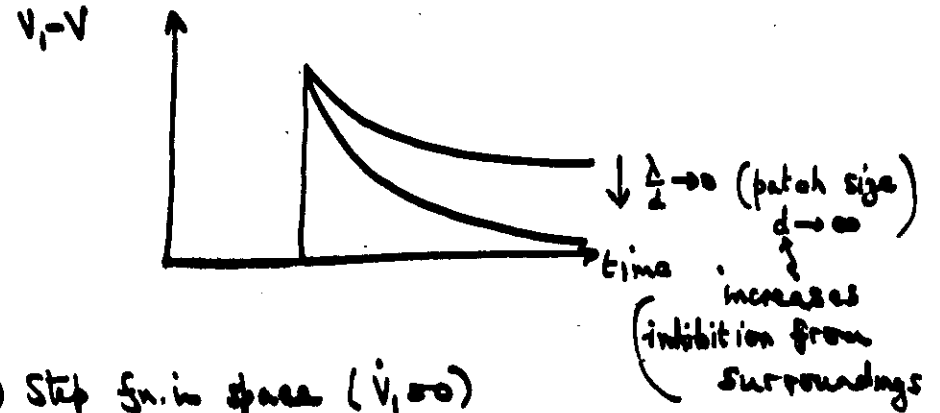
1) Step fn. in time

$$V_1(t) = a + b\theta(t)$$

$$\tilde{V}(\omega) = \int_0^\infty e^{-i\omega t} dt = \frac{1}{i(\omega - i\epsilon)}$$

Can show

$$V_1(\mathbf{x}, t) - V(\mathbf{x}, t) = \frac{1}{2\pi} b\theta(t) e^{-tG_1/C} + V_1 \frac{\lambda^2}{d^2} + O(\frac{\lambda^4}{d^4})$$



2) Step fn. in space ($\dot{V}_1 = 0$)

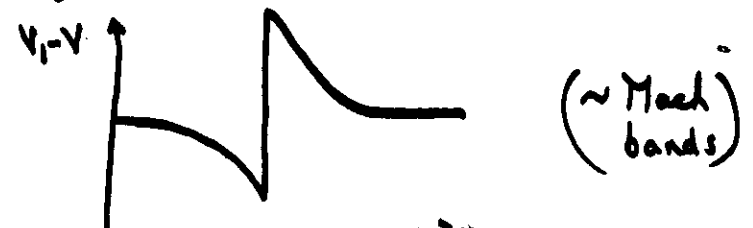
$$V - \lambda^2 \nabla^2 V = V_1(\mathbf{x})$$

In 1D: $\dots \dots \dots \tilde{V} \dots \dots \dots V_1$

$$V = b - \frac{1}{2} b e^{-x/\lambda} \quad x > 0$$

$$= \frac{1}{2} b e^{x/\lambda} \quad x < 0$$

$$\Rightarrow V_1 - V = \frac{1}{2} b [e^{-x/\lambda} \theta(x) - e^{x/\lambda} \theta(-x)]$$



2/a

3) Moving edge

$$V_1 = a + f \Theta(x-vt)$$

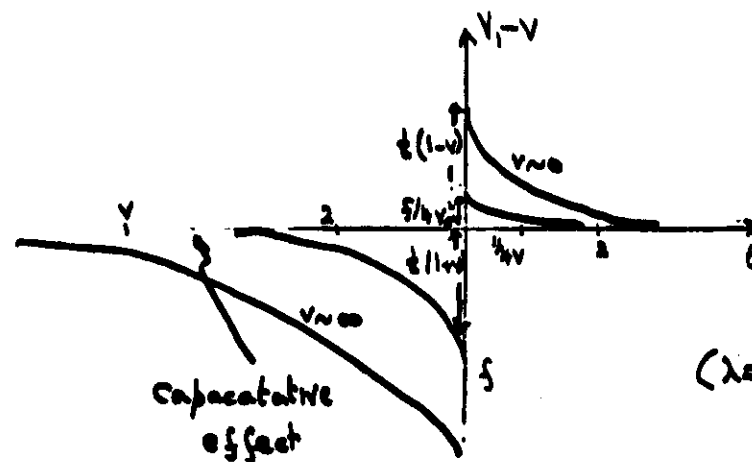
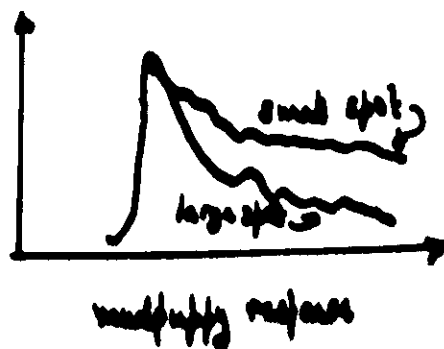
$$\Rightarrow \tilde{V}_1 = (2\pi)^2 a \delta^2(k) \delta(\omega) + (2\pi)^2 f \delta(k_y) \left(\frac{1}{ik_x} \right) \delta(\omega + k_x v)$$

$$\Rightarrow V_1 - V = \frac{f}{\sqrt{k\lambda^2 + \frac{v^2}{c_1^2}}} \left[\frac{e^{+k \cdot (vt-x)}}{k_-} \Theta(vt-x) + \frac{e^{-k \cdot (x-vt)}}{k_+} \Theta(x-vt) \right]$$

$$k_{\pm} = \frac{1}{2\lambda} \left[\left(\frac{v}{c_1} \right) \pm \sqrt{k\lambda^2 + \left(\frac{v}{c_1} \right)^2} \right]$$

$$v \sim 0: V_1 - V = \frac{f}{2} \left[-\frac{1}{\lambda} e^{-(vt-x)\lambda} \Theta(vt-x) + \frac{(1-\frac{v}{c_1})}{\lambda} e^{-(x-vt)\lambda} \Theta(x-vt) \right]$$

$$v \sim \infty: V_1 - V = \frac{f}{2} \left[-e^{-q_1(vt-x)\lambda/c_1} \Theta(vt-x) + \frac{q_1 \lambda}{v c_1} e^{-\frac{v}{c_1} \lambda (x-vt)} \Theta(x-vt) \right]$$



$$(\lambda = \frac{1}{2}, c = q_1 = 1)$$

2/

So, v is important for large v.

2.3. Central Pattern Generators (CPG's)

(Loech: Science 200, 1348, 78
lobster: TINS April 82, 120)

- Small #s of neurons controlling

rythmic muscular contractions:

in walking, flying, swimming

stomach muscles

insects

leech

lobster

- Circadian rhythm generators

crab eye
response

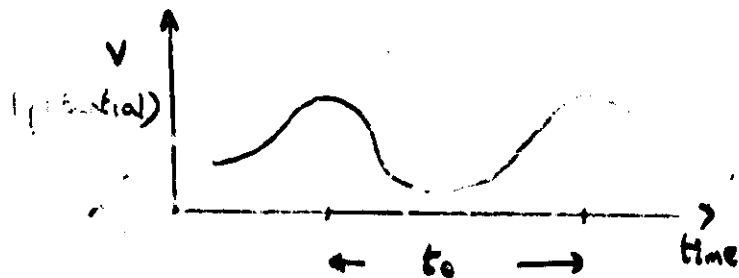
response of eye

to light

flash

Noon 6pm night 6am Noon

- cockroach limb movement:



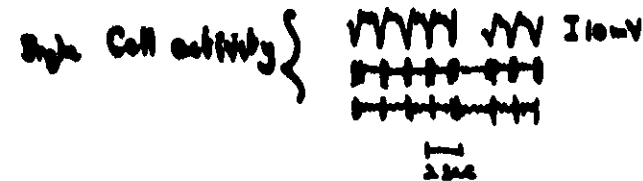
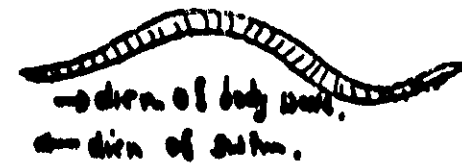
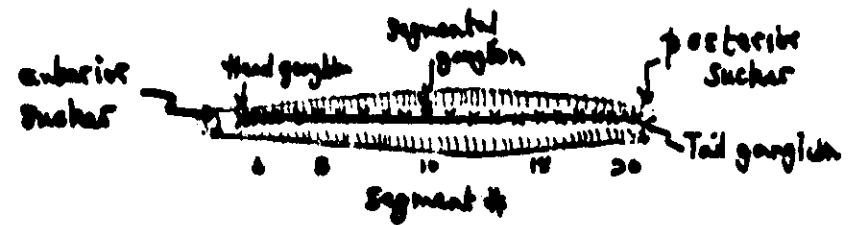
non-shifting interneurons v. L. Hartman

May be due to either

(a) endogenous oscillator

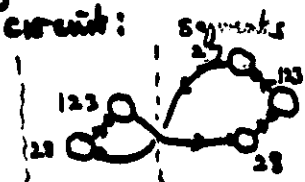
(b) oscillatory network

e.g. (b): leech swimming movement



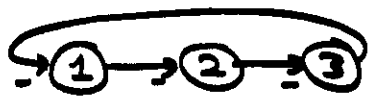
Caused by

5-neuron circuit:

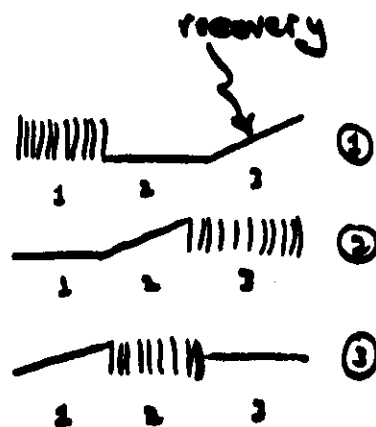


all synapses inhibitory

eg. 3 cell network:



phase 1	1	0	0 _r
2	0	0 _r	1
3	0 _r	1	0



odd # of recurrently inhibitory cells

eg. 5:

	a28	an2	p28	p128	p27	
phase 1	1	0	1	0	0 _r	
2	0	0 _r	1	0	1	
3	0	1	0	0 _r	1	
4	0 _r	1	0	1	0	
5	1	0	0 _r	1	0	

(need cell in recovery phase for (0)'ⁿ)

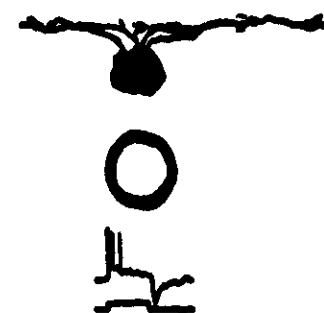
($\Rightarrow$) "The Crustacean Stomatodentate System"
ed. Selverston & Moulins, Springer '87

Inner Plexiform Layer:

Ganglion cells: on p



on d



small convergence

16 cones $\rightarrow$ 4 CBP's $\rightarrow$ 1 on p

Sterling et al
TINS 2 (1986)

high convergence

200 CBP's $\rightarrow$ 1 on d

gives transient + steady

continuous limb:

$$\int (V_i - V)(x) dx = 0$$

so only transient.

Rods: 1500 rods $\rightarrow$ 1 on p
(thru amacrine)

(no hor. effects: only on on d)

Conclusions:

- can model invertebrate retina by
noisy neurons, or by linear model (to within
self-inhibition?)
- can model vertebrate IPL by R-C network
- can obtain analytic solutions in continuum
limit ([2] - field theory)
- can model on or OFF α - and β -ganglion cells
(without details of mechanisms, no motion detection)

