



INTERNATIONAL ATOMIC ENERGY AGENCY
UNITED NATIONS EDUCATIONAL, SCIENTIFIC AND CULTURAL ORGANIZATION
INTERNATIONAL CENTRE FOR THEORETICAL PHYSICS
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H4.SMR/775-19

**COLLEGE IN BIOPHYSICS:
EXPERIMENTAL AND THEORETICAL ASPECTS OF
BIOMOLECULES**

26 September - 14 October 1994

Miramare - Trieste, Italy

Protein Dynamics II & III A

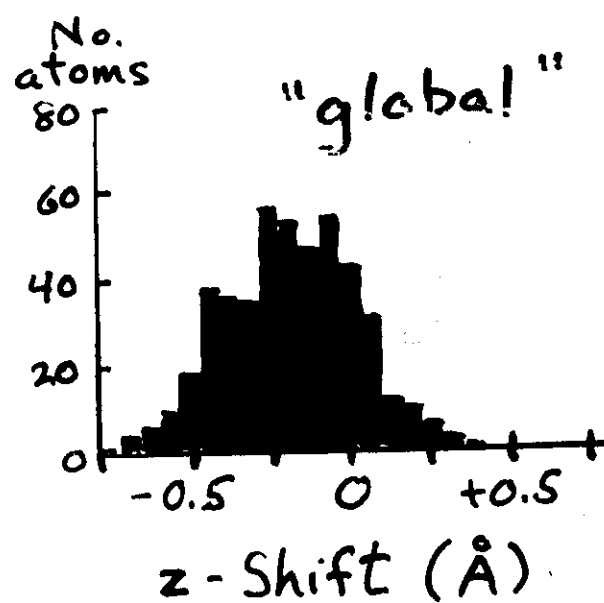
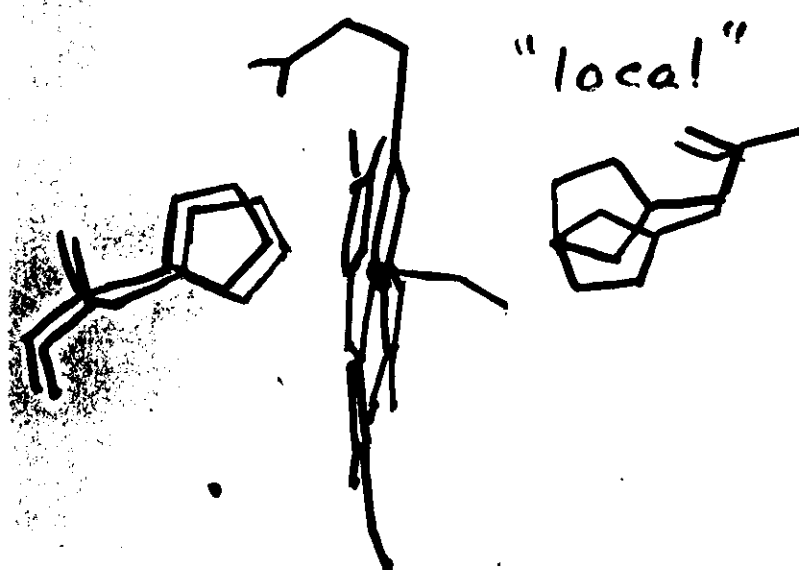
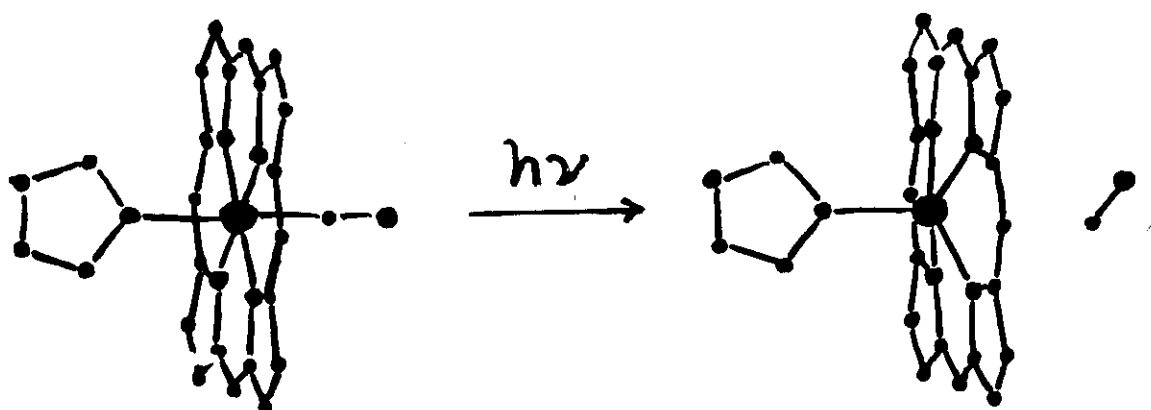
**William A. Eaton
National Institutes of Health
Bethesda, Maryland - USA**

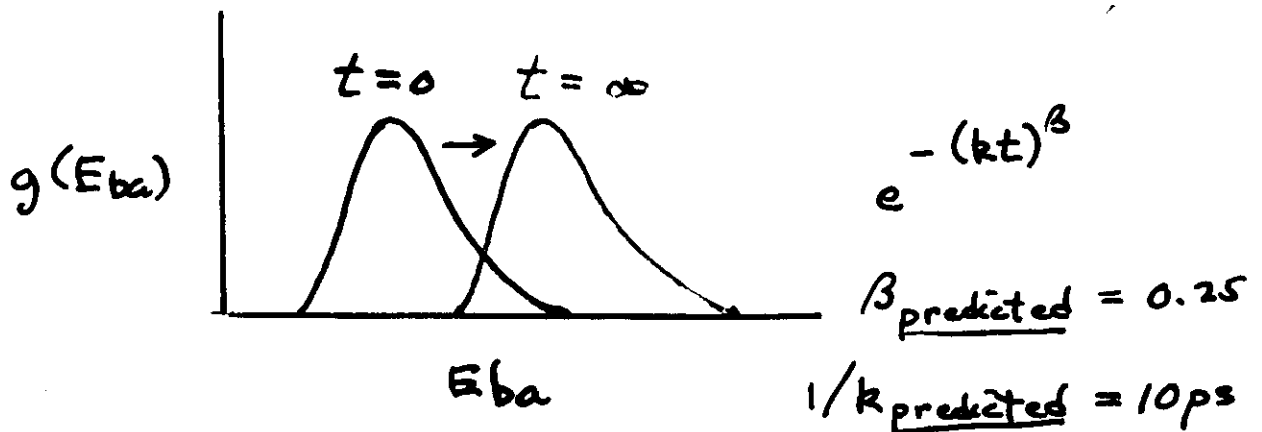
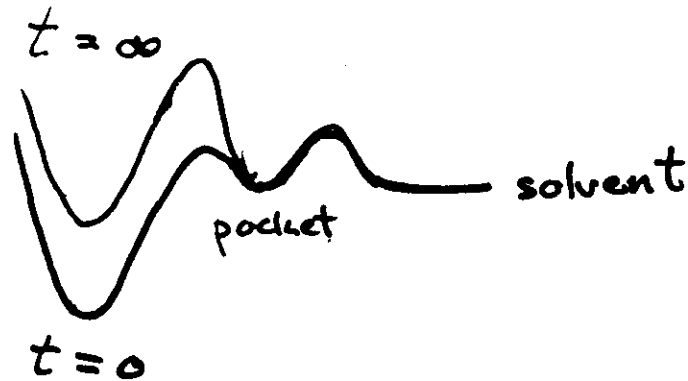
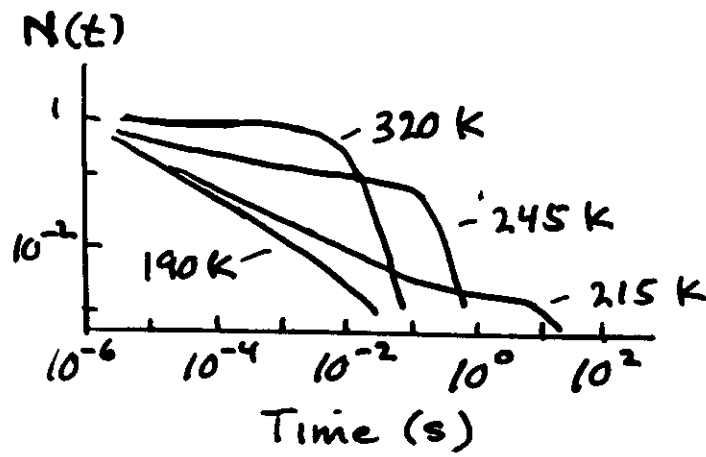
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MICROPROCESSOR LAB.	VIA BEIRUT, 31	TEL. 224471	TELEFAX 224163	TELEX 460392	GALILEO GUEST HOUSE	VIA BEIRUT, 7	TEL. 22401	TELEFAX 224559	TELEX 460392

Eaton

Lecture II

Two "Fairytale"





An alternative view (Agmon & Hopfield, 1983) - protein relaxation shifts barrier height distribution.

Stenbach, Frauenfelder et al. 1991 - predicted kinetics of protein relaxation

$$e^{-(kt)^\beta}$$

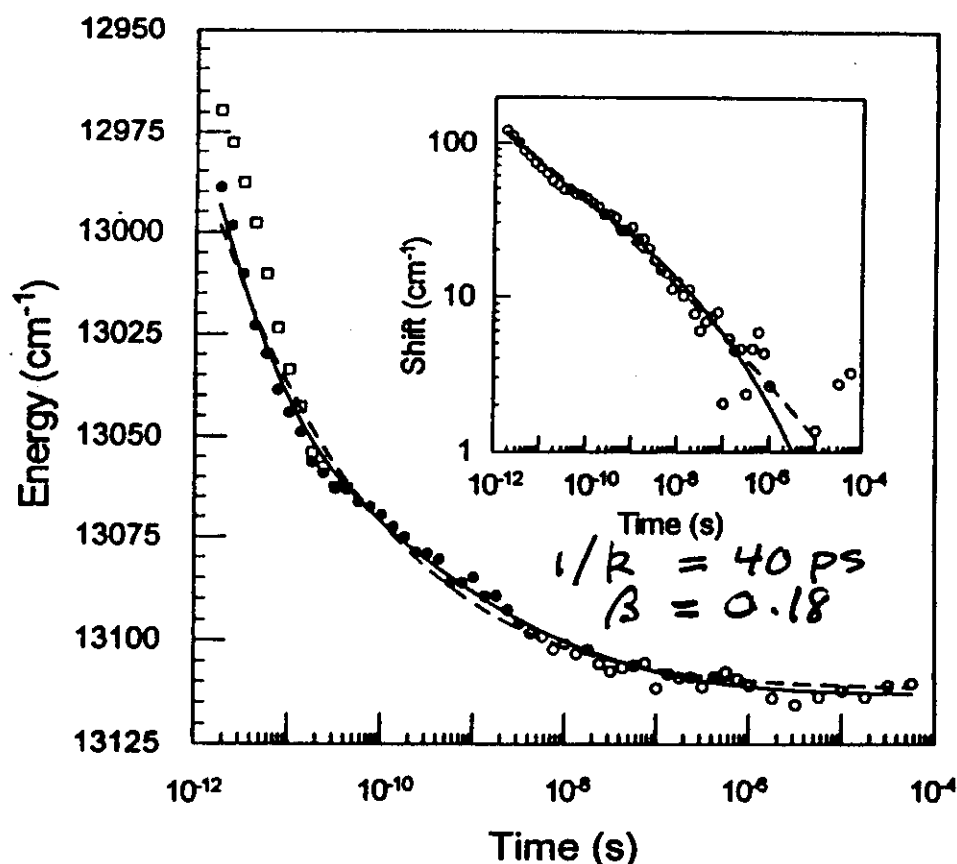
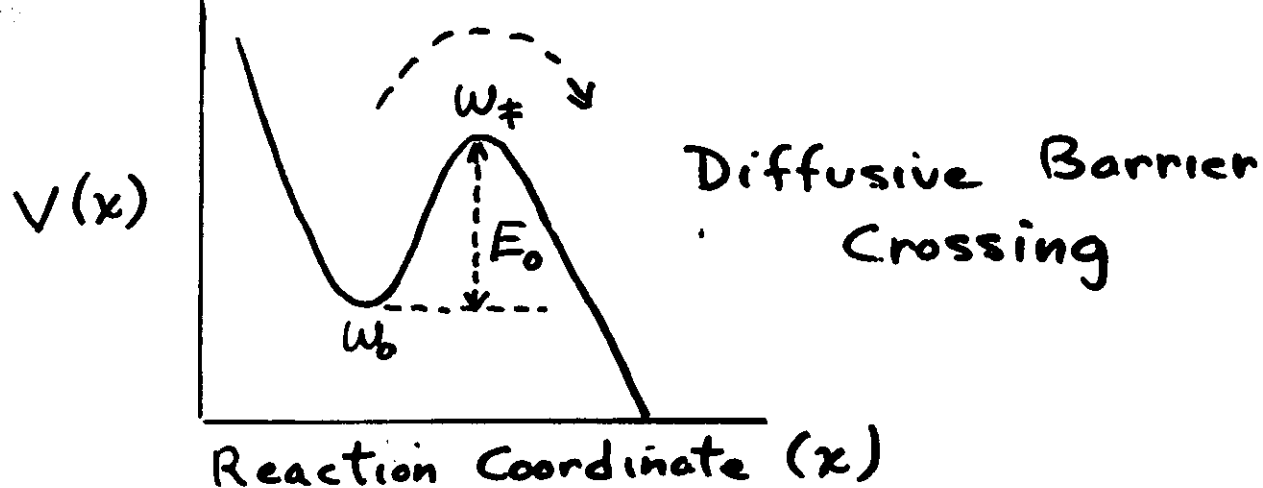


Fig. 2. Time-dependent position of band III with and without compensation for thermal relaxation. The filled and open circles represent data collected with femtosecond and nanosecond pump pulses, respectively. The open squares show data uncorrected for thermal relaxation. Note that the thermal correction is significant only for times shorter than ~ 20 ps. The data were modeled with a modified stretched exponential function (solid line), and a conventional stretched exponential function (dashed line) over the entire range of times (see parameters in table 1). The inset figure is a log-log plot of the thermally compensated shift of band III relative to equilibrium Mb.



Dynamics described by Langevin equation:

$$M \frac{d^2 x}{dt^2} = -\frac{dV}{dx} - \zeta \frac{dx}{dt} + R(t)$$

which, in high friction limit, yields Kramers' equation:

$$k = \frac{M \omega_0 \omega^\ddagger}{2\pi\zeta} \exp\left(-\frac{E_0}{k_B T}\right)$$

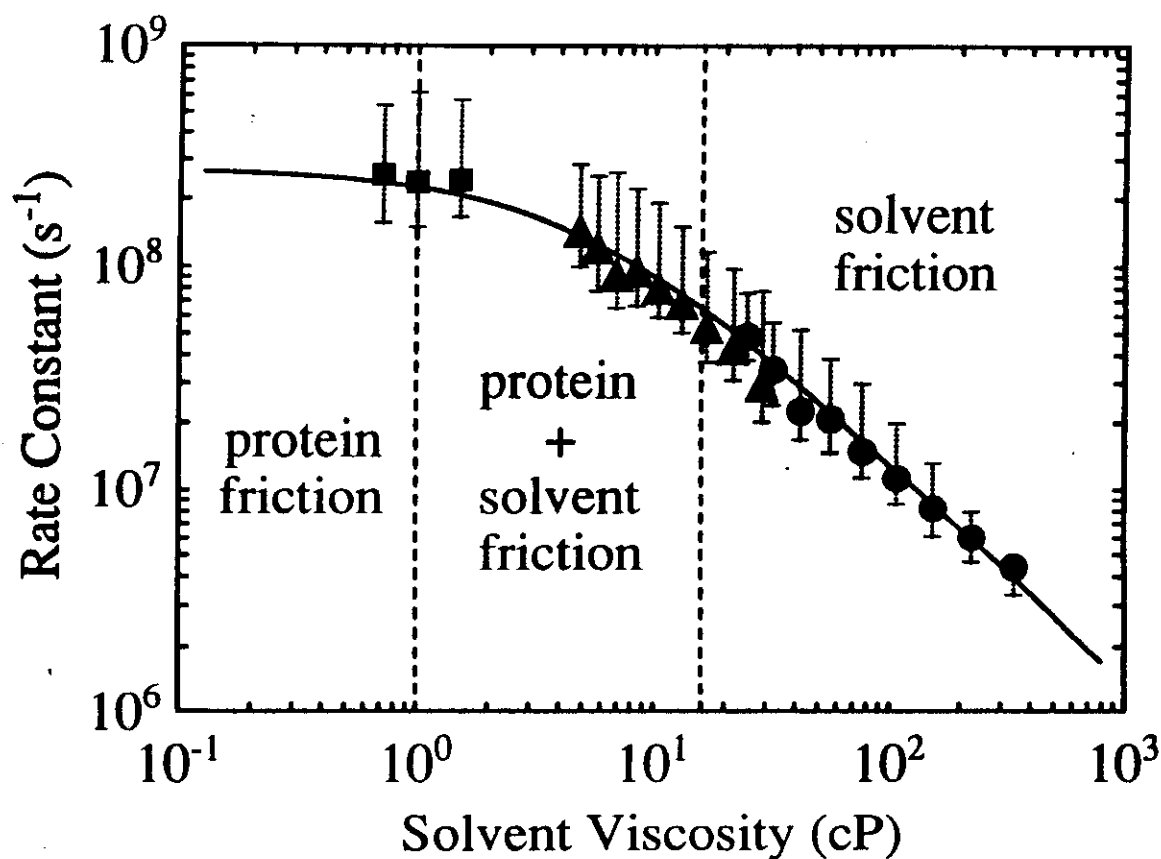
Assuming that protein friction and solvent friction are additive, Kramers' equation is:

$$k = \frac{B}{\alpha\zeta_p + (1-\alpha)\zeta_s} \exp\left(-\frac{E_0}{k_B T}\right)$$

which, using Stokes' law, becomes:

$$k = \frac{C}{\sigma + \eta_s} \exp\left(-\frac{E_0}{k_B T}\right)$$

Myoglobin Conformational Relaxation



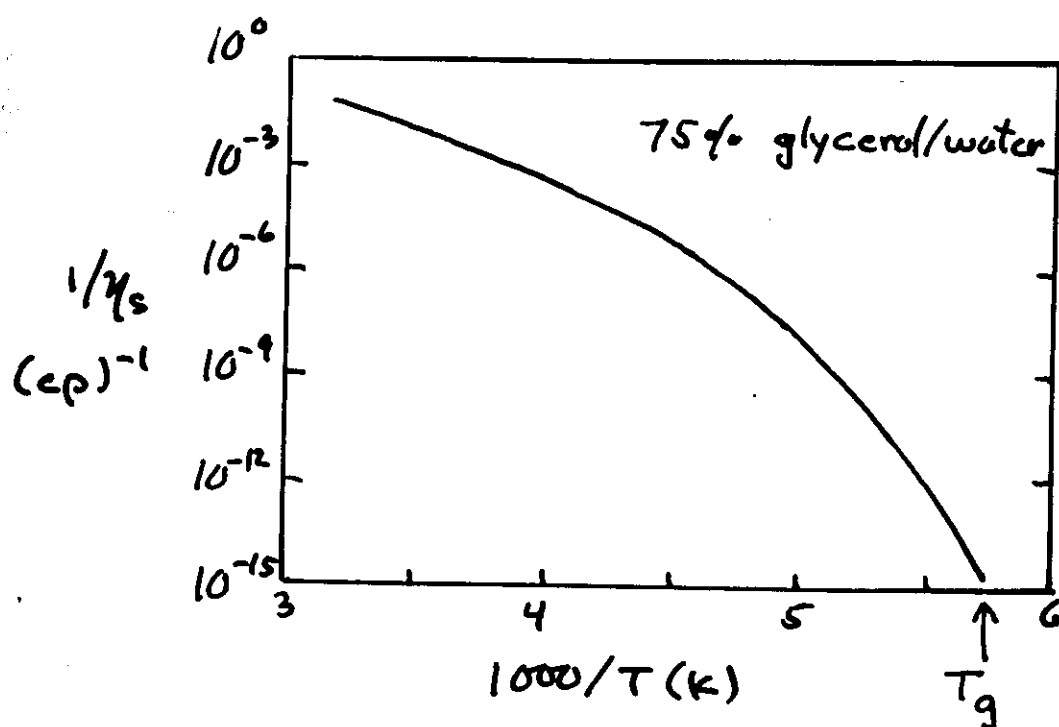
$$k = \frac{C}{\sigma + \eta_s} e^{-E_0/k_B T}$$

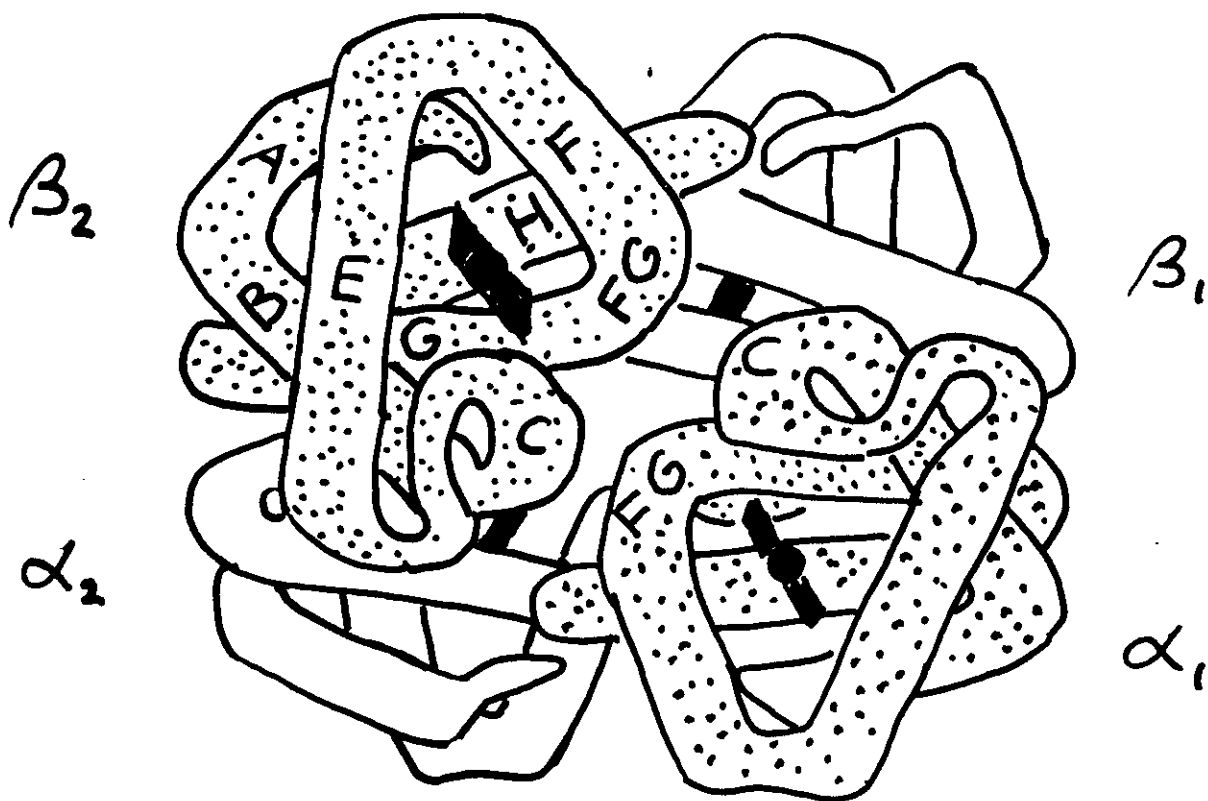
$$\sigma = 4 \text{ cP}$$

$$E_0 = 2.4 \text{ kcal/mole}$$

$$C = 7 \times 10^{10} \text{ cP/sec}$$

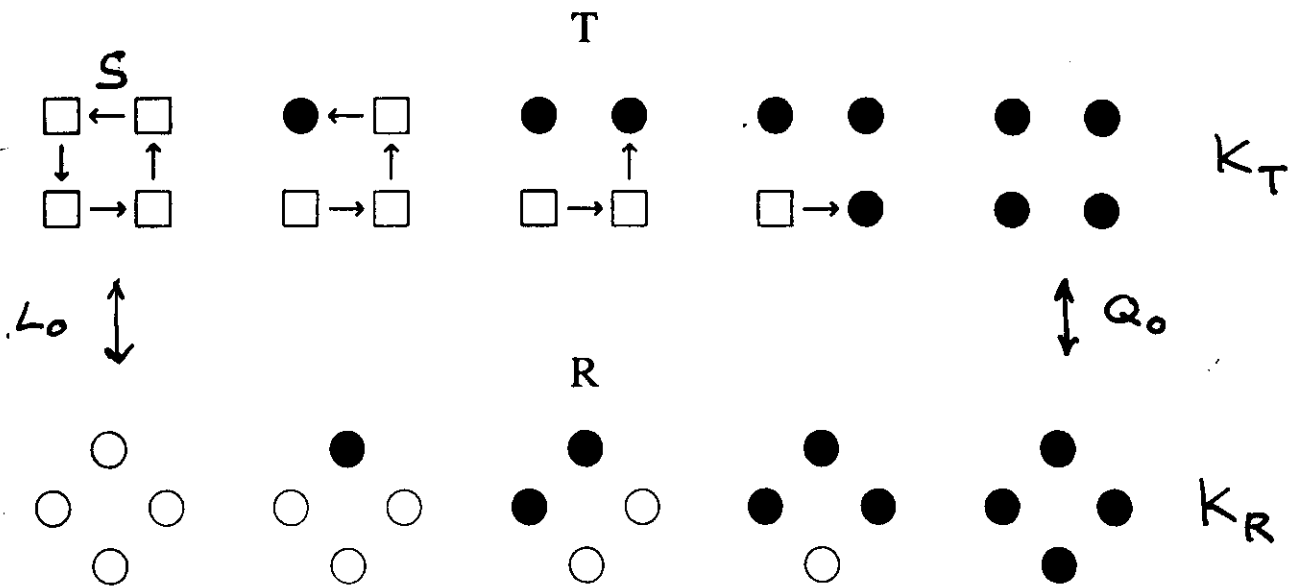
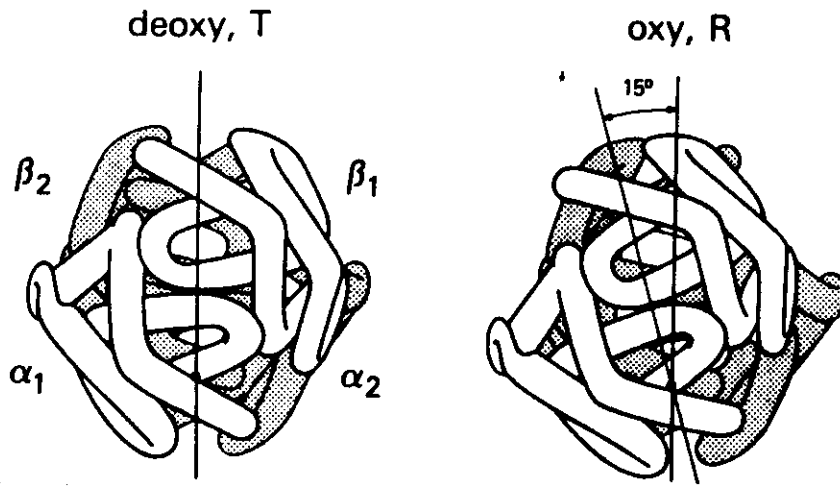
$$k = \frac{c}{\phi + \eta_s} e^{-E_0/RT}$$





MWC - PSK Model

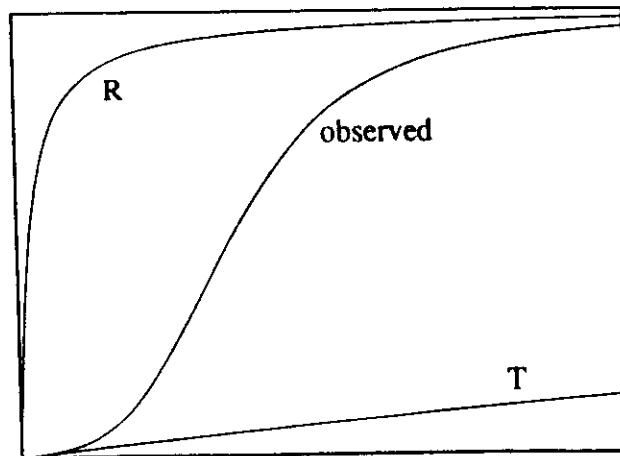
Monod
Wyman
Changeux
Perutz
Szabo
Karplus



MWC
 K_R, K_T, L_0
 $L_i = L_0 c^i$

SK
 K_R, S, Q_0

Fractional Saturation



Oxygen Pressure

$$K_T = K_R / S$$

$$L_0 = Q_0 S^4$$

Linear Free Energy Relation

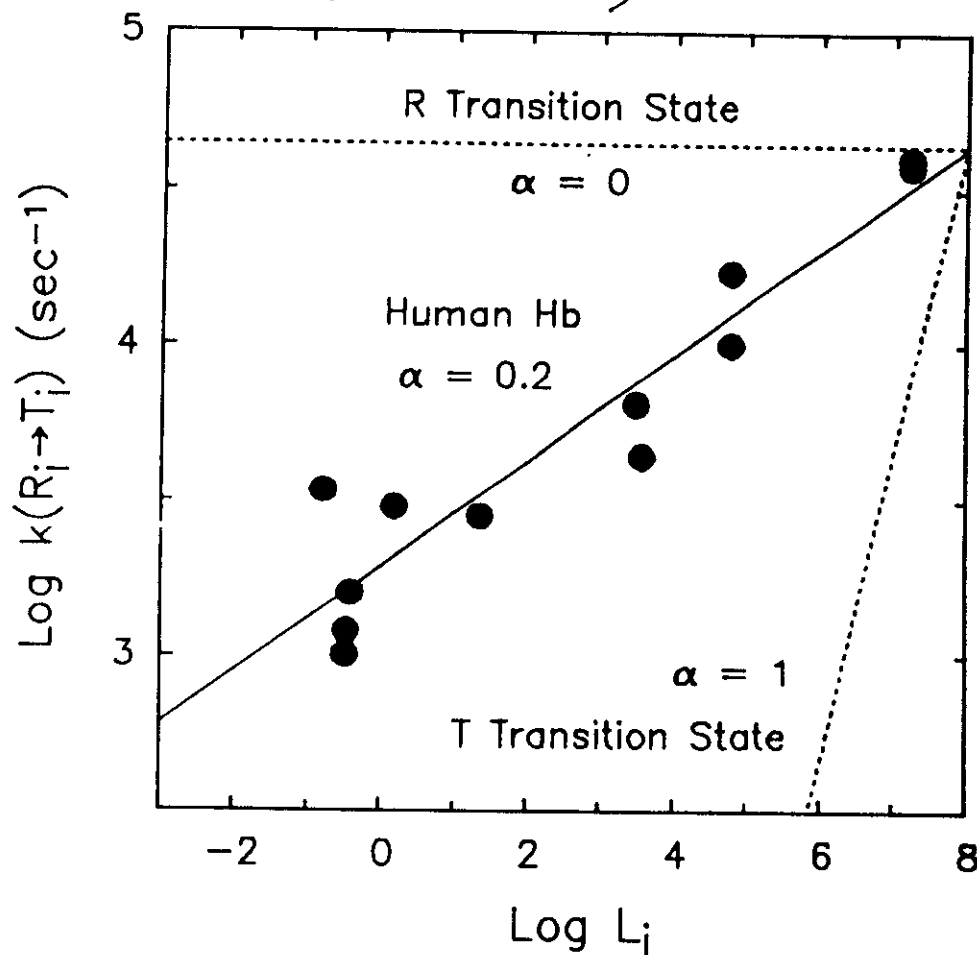
$$\delta \Delta G^\ddagger = \alpha \delta \Delta G$$

For two-state allosteric model

$$k(R_i \rightarrow T_i) = \gamma (L_o c^i)^\alpha$$

$$k(T_i \rightarrow R_i) = \gamma (L_o c^i)^{\alpha-1}$$

$$(d = c^{-\alpha})$$

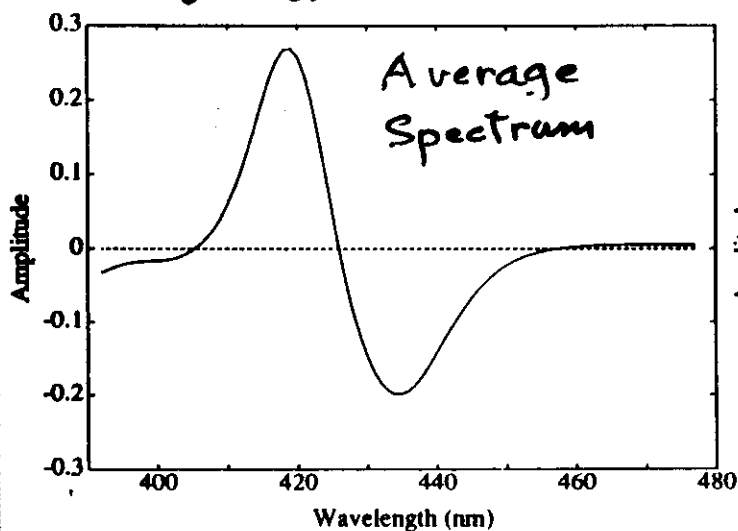


Transition state appears at 20% of the distance along the reaction path from R to T.

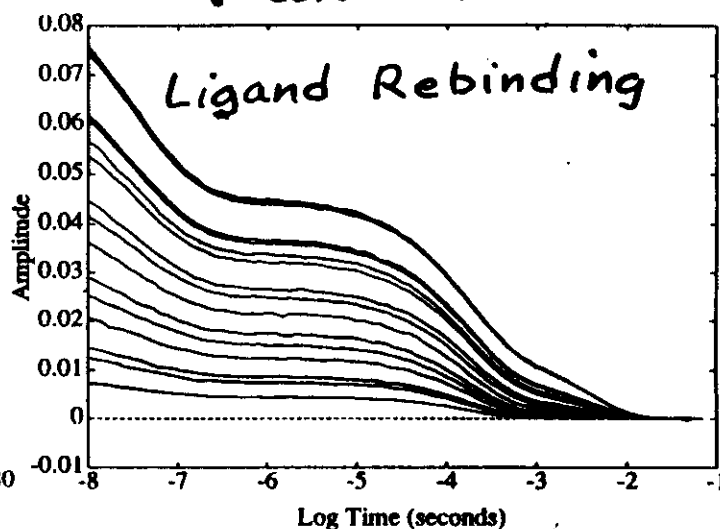
Singular Value Decomposition (of isotropically-averaged data)

$$A = U S V^T$$

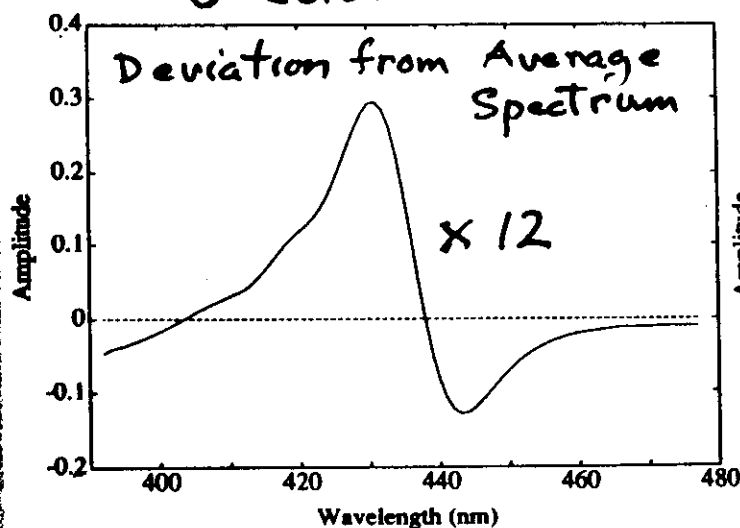
U Column 1



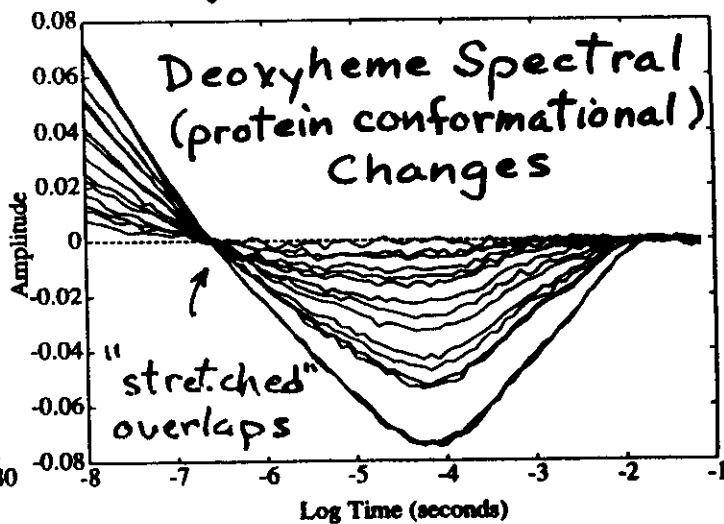
V column 1



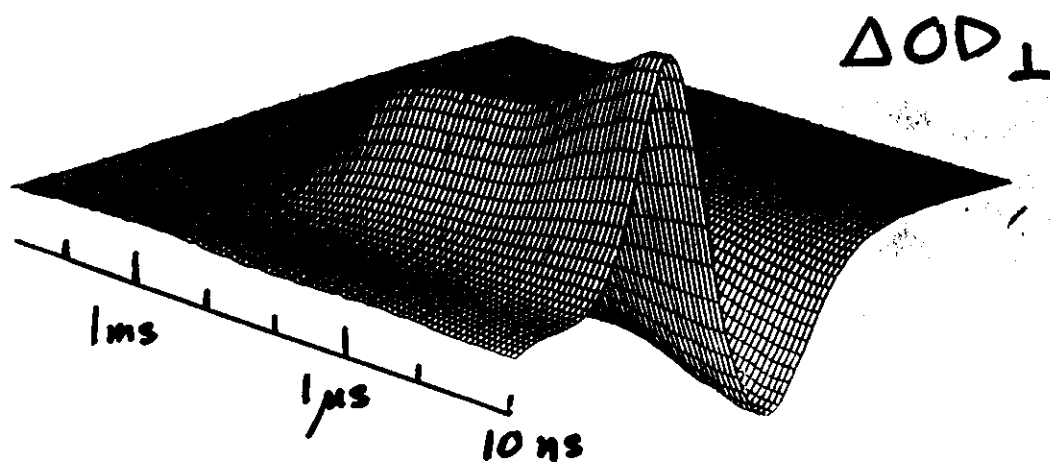
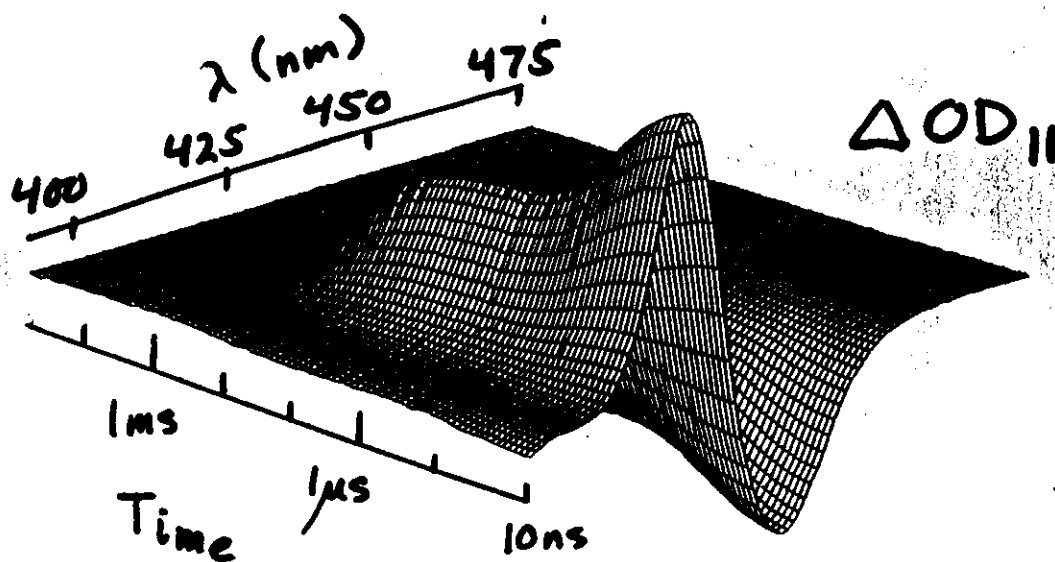
U column 2



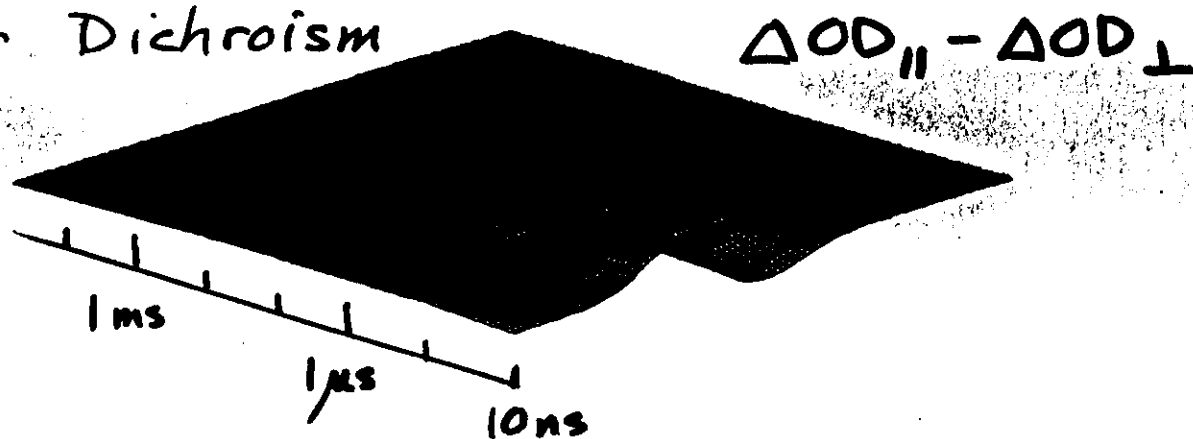
V column 2

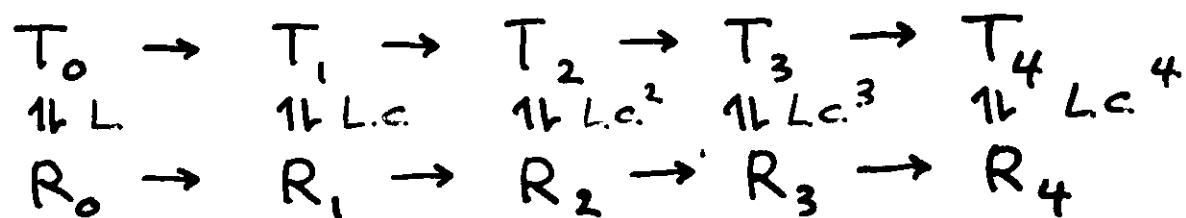


Polarized Spectra

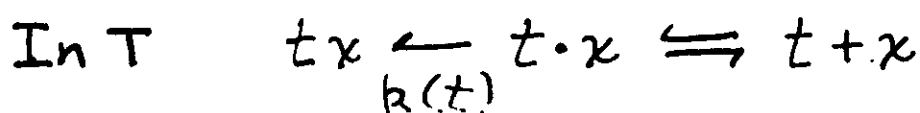
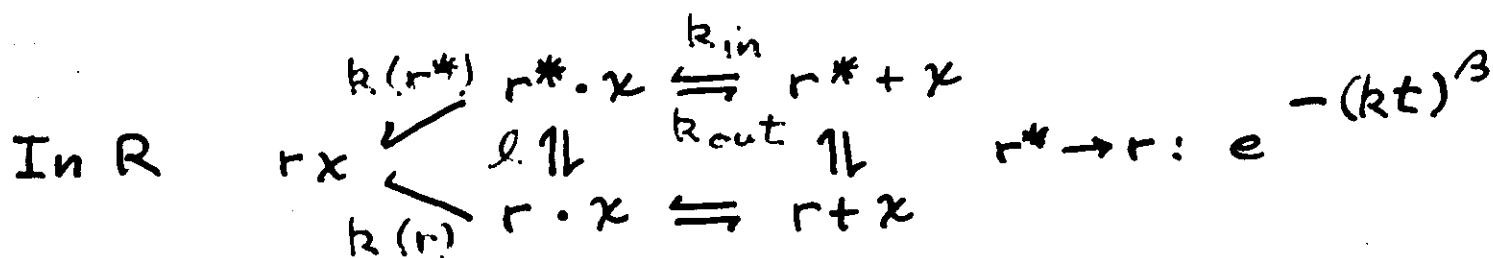


Linear Dichroism





$$k(R_i \rightarrow T_i) = \chi (Lc^i)^\alpha \quad (\text{Gibson } d = c^{-\alpha})$$



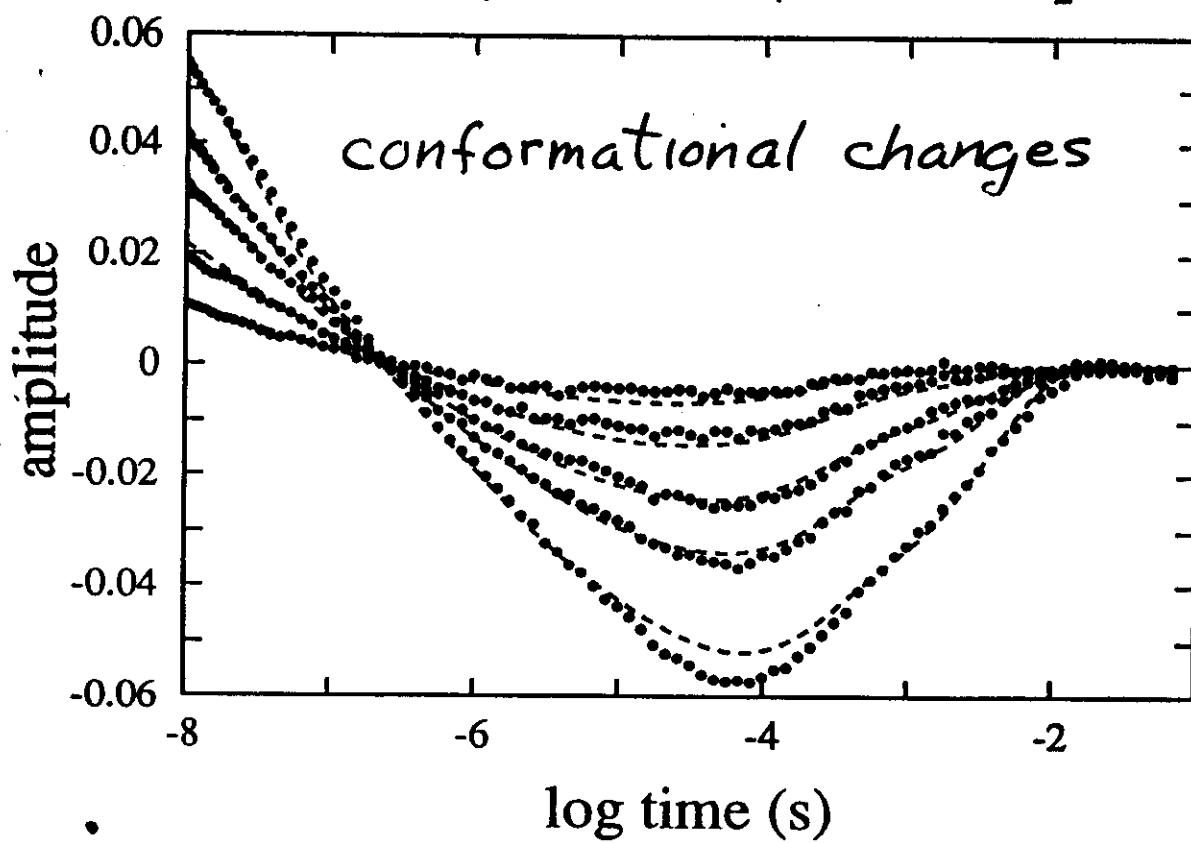
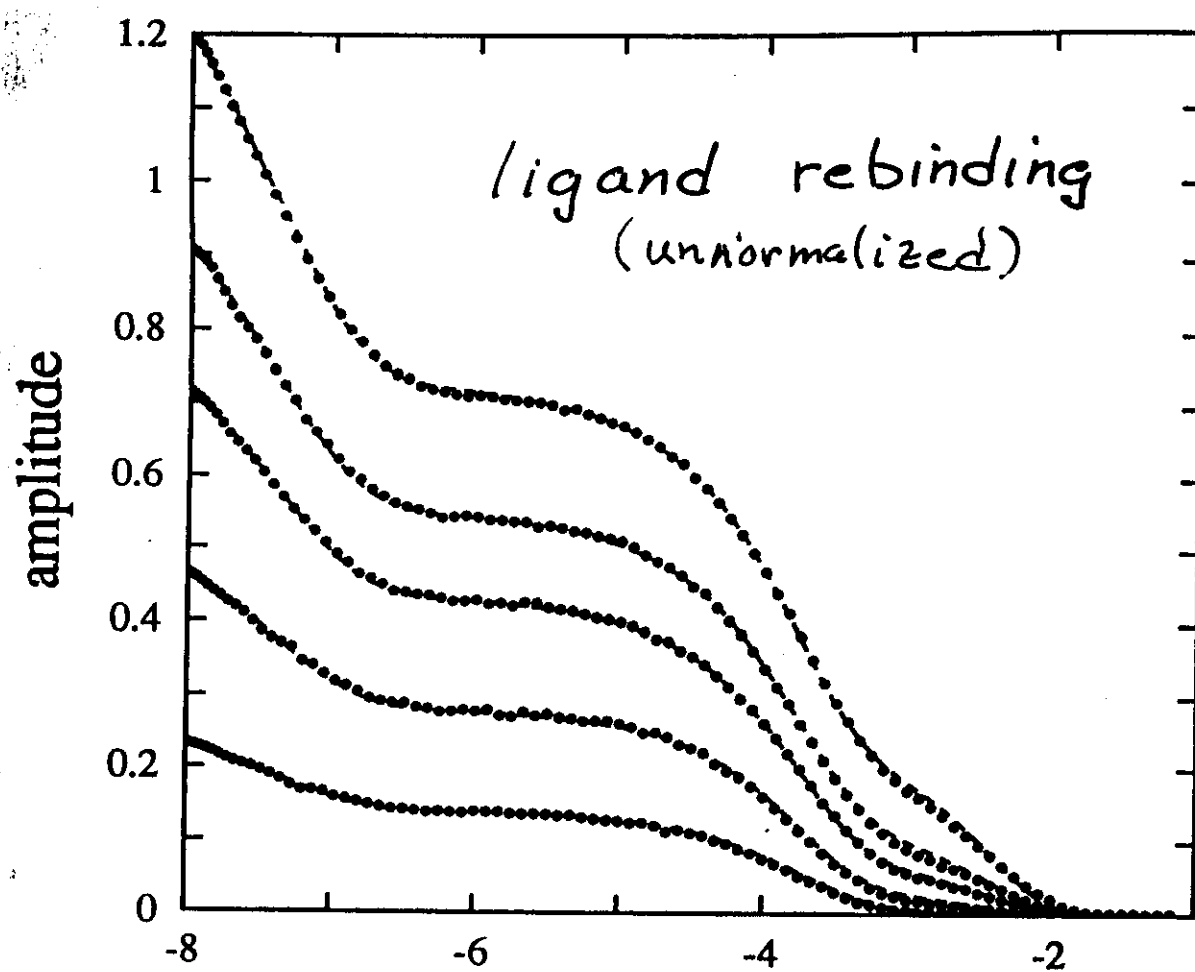
L, c, χ, α	} quaternary rates
k, β, ℓ	
$k(r^*), k(r), k(t)$	
k_{in}, k_{out}	
	tertiary rates
	ligand binding rates

85 distinguishable species, 3 spectra

12 model parameters

to describe

37 experimental parameters, 2 spectra



PARAMETERS OBTAINED FROM FITS WITH TWO-STATE MWC ALLOSTERIC MODEL WITH TERTIARY TRANSIENT

$$L = 2 \times 10^7$$

$$(\equiv [T_O]/[R_O])$$

simultaneously fit to oxygen
binding curve of Imai

$$c = 0.002 \text{ (CO)}, 0.003 \text{ (O}_2\text{)}$$

$$(\equiv p50(T)/p50(R))$$

$$l = 15$$

$$(\equiv [r]/[r^*])$$

r^* is only 6% populated at
equilibrium, i.e. "transient"

$$k_{gem}(r^*) = 4 \times 10^7 \text{ s}^{-1}$$

$$k_{gem}(r) = 4 \times 10^5 \text{ s}^{-1}$$

tertiary relaxation has marked
effect on geminate rebinding
rate, as in myoglobin

$$k_{gem}(t) = 1 \times 10^4 \text{ s}^{-1}$$

quaternary relaxation also has
marked effect on geminate
rebinding rate

$$k_{in} = 1 \times 10^8 \text{ M}^{-1}\text{s}^{-1}$$

$$k_{out} = 7 \times 10^6 \text{ s}^{-1}$$

geminate yield determined by
ligand exit rate

$$k(R_3 \rightarrow T_3) = 500 \text{ s}^{-1}$$

value constrained to be within
a factor of two of the value
obtained by Ferrone from
modulation experiments at very
low light intensities

$$\alpha = 0.2$$

linear free energy exponent,
same as value reported by Eaton
et al., 1991

$$\text{yields } k(R_0 \rightarrow T_0) = 4 \times 10^4 \text{ s}^{-1}$$

$R \rightarrow T$ rate increase by a factor
of $15^{0.2} = 1.7$ for every r^*
subunit

$$k(r^* \rightarrow r) = 3 \times 10^6 \text{ s}^{-1}$$

$$\beta = 0.3$$

overlaps quaternary relaxation
because of small β

Eaton

Lecture III

Sickle Cell Hemoglobin

Kinetic Equations of Double Nucleation Model

$$\frac{dc_p}{dt} = \underset{\text{hom.}}{k_+ c c_{i^*}} + \underset{\text{het.}}{k_+ c c_{j^*}}$$

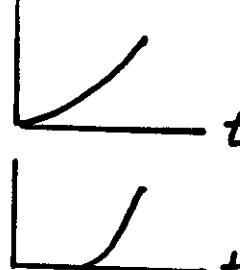
assuming monomer $\rightleftharpoons$ nucleus equilibrium

$$\frac{dc_p}{dt} = k_+ K_{i^*} c^{i^*+1} + k_+ K_{j^*} \phi (c_0 - c) c^{j^*+1}$$

$$-\frac{dc}{dt} = k_+ c_p c - k_- c_p$$

integrating linearized rate equations, $c(t) \sim c_0$

$$c_0 - c(t) = A [\cosh(Bt) - 1]$$

$$= \begin{cases} \frac{1}{2} AB^2 t^2 & Bt \ll 1 \\ \frac{1}{2} Ae^{Bt} & Bt \gg 1 \end{cases} \quad \begin{array}{l} c_0 - c \\ t \end{array}$$


$$A = \frac{K_{i^*} c_0^{i^*+1} \text{ hom}}{K_{j^*} \phi c_0^{j^*+1} \text{ het}}$$

$$B^2 = k_+^2 (c_0 - c) \phi K_{j^*} c_0^{j^*+1}$$

Domain
Formation
Rate
(mM s^{-1})

