



1936-13

Advanced School on Synchrotron and Free Electron Laser Sources and their Multidisciplinary Applications

7 - 25 April 2008

DEVELOPMENTS and
APPLICATIONS of
X-RAY MICROANALYTICAL
TECHNIQUES BASED on X-RAY TUBES
and SR SOURCES

Andrzej Markowicz

IAEA Laboratories
Seiberdforf

Austria

DEVELOPMENTS and APPLICATIONS of X-RAY MICROANALYTICAL TECHNIQUES BASED on X-RAY TUBES and SR SOURCES

Andrzej Markowicz



IAEA Laboratories
Seibersdorf
Austria

ICTP, Trieste, 22 April 2008

Outline

- 1. Introduction
- 2. Micro-analytical techniques
- 3. Selected applications
- 4. Micro-tomography
- 5. Confocal setup
- 6. Sample preparation procedures
- 7. SR-based applications
- 8. Training
- 9. Conclusions



1. Introduction

- Assistance in establishing a modern and safe technological base to implement nuclear techniques and technologies for peaceful applications
- Laboratories at Seibersdorf and Monaco provide experimental support to the Agency's programmes
- Transfer of technologies through Technical Co-operation (TC)



1. IAEA Seibersdorf Laboratories



The IAEA Laboratories are located about 35 km south of Vienna, Austria



1. Introduction (cont.)

XRF Laboratory at Seibersdorf

- R & D
- Analytical services
- Co-ordination and support of research
- Proficiency tests
- Development of QA/QC procedures
- Technical backstopping to TC projects
- Training
- Dissemination of information



2. Facilities and techniques available

Laboratory instruments and techniques

EDXRF spectrometers

Total reflection XRF spectrometers

Micro-beam X-ray fluorescence and absorption technique



2. Secondary, polarizing target spectrometers

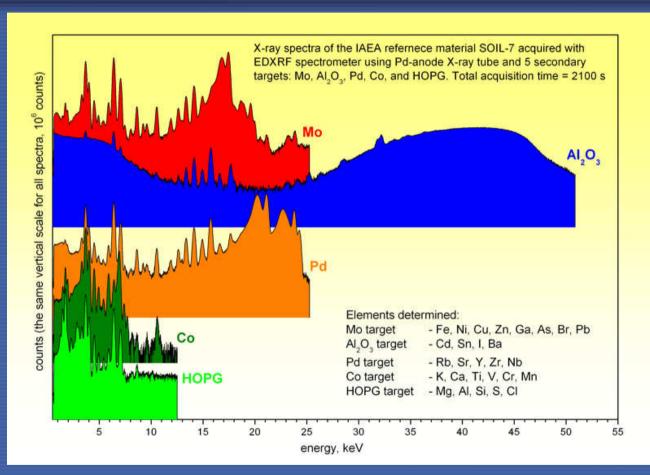




Left: heavy-duty, fully software controlled EDXRF spectrometer (Pd-anode X-ray tube) utilizing 5 secondary targets (Pd, HOPG, Co, Mo, Al₂O₃); right: in-house assembled EDXRF spectrometer (Cr-, Fe-, Cu-, Mo-, Rh-, Ag-, W-anode), manual selection of secondary targets (Mo, Rh, Ti in use, any other solid target possible)



2. Comparison of X-ray spectra



X-ray spectra of a "thick" pellet sample made of Soil-7 reference material excited in the EDXRF secondary target spectrometer.



2. Total reflection X-ray fluorescence spectrometers



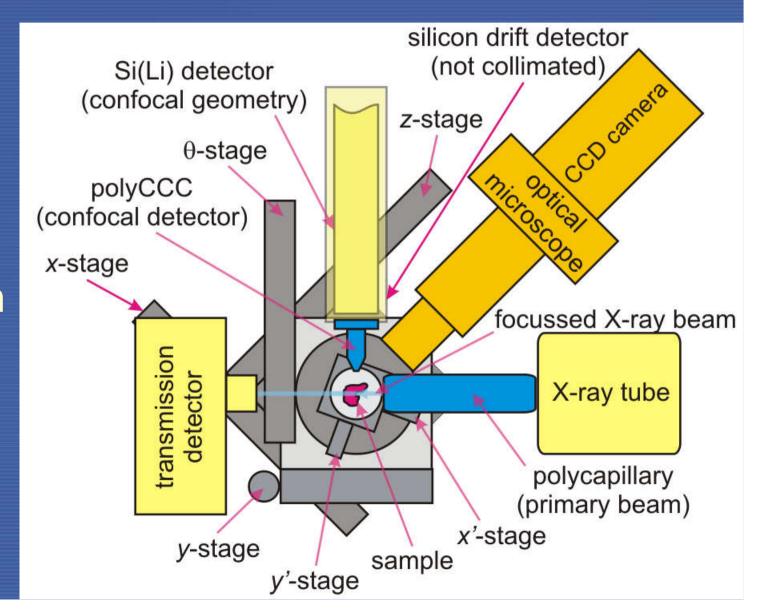


TXRF spectrometers, left: with 12-position sample changer; right: with motorized beam modifier and sample reflector stages



2. Micro-beam X-ray Setup Scheme

spatial resolution: 15 - 40 μm





2.The Micro-Beam X-ray Scanning Spectrometer

Si drift detector (fluorescence) on a stage

optical microscope X-ray tube

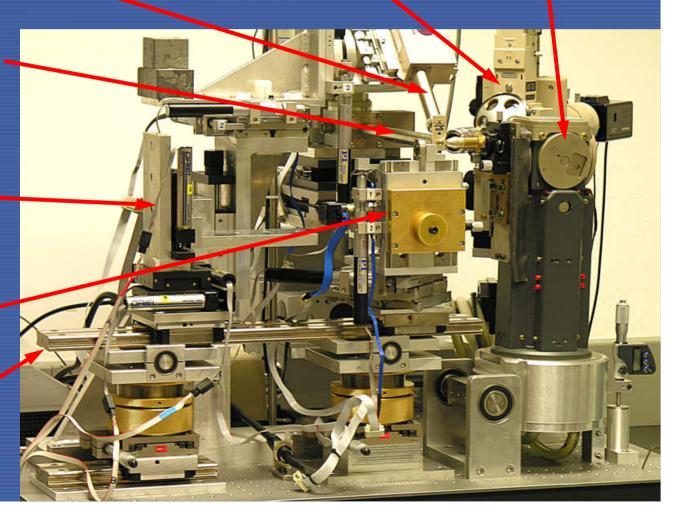
Si(Li) detector (confocal)

transmission detector stage

sample stage

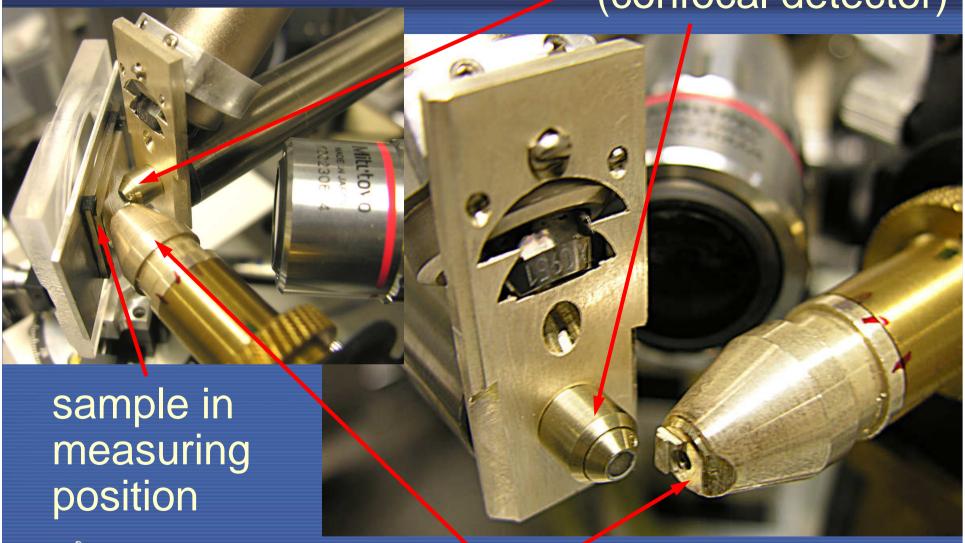
optical bench-





2. Measuring geometry polyCCC

(confocal detector)

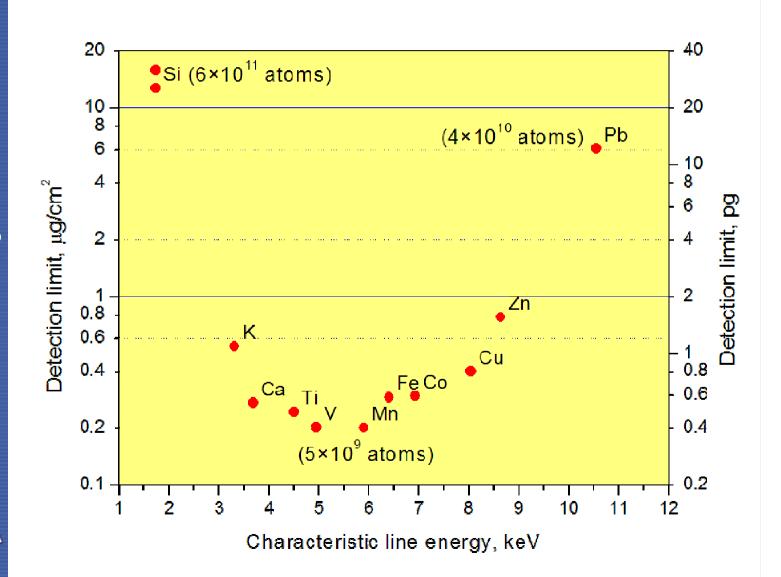




polycapillary (primary beam) 12

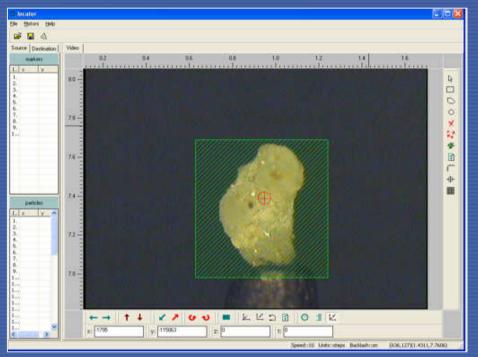
2. Detection Limits of Elements

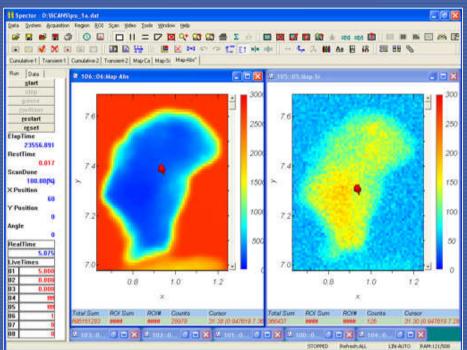
 $40 \mu m$ t = 1000s





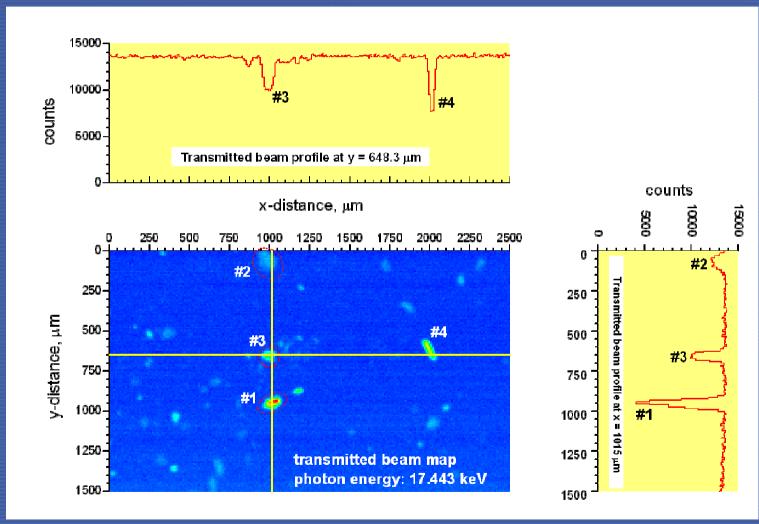
2. Data acquisition software

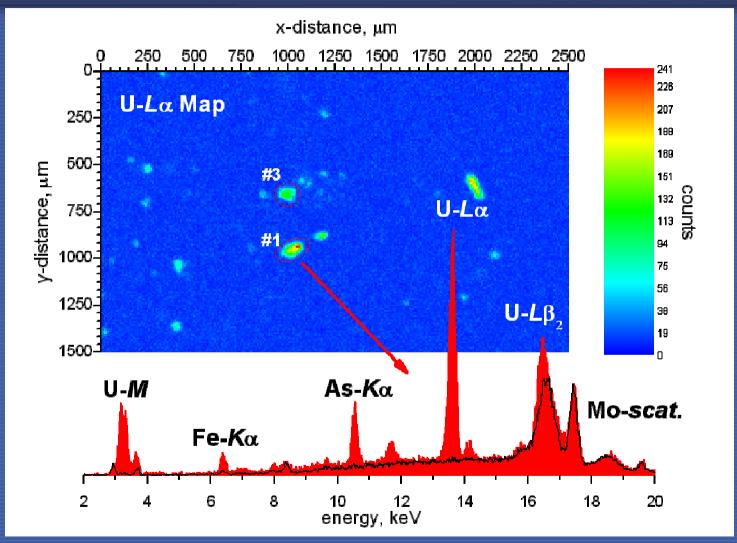




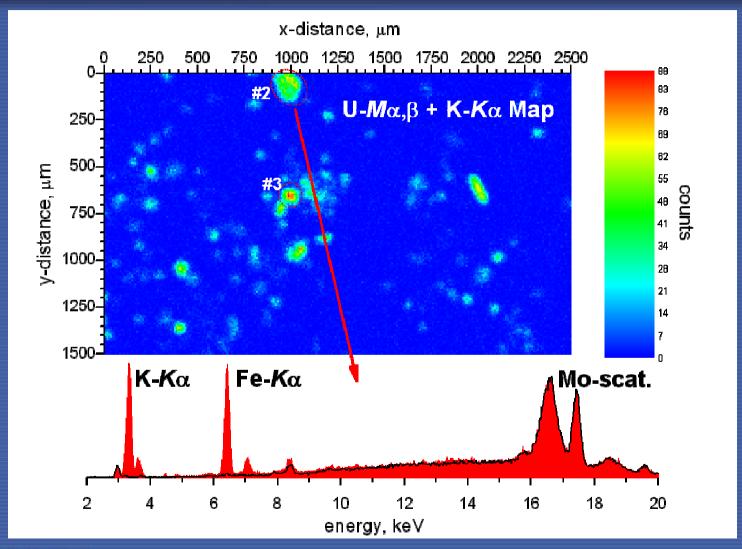
SPECTOR-LOCATOR: microscopic image of the sample (×150) and the collected X-ray absorption/fluorescence images



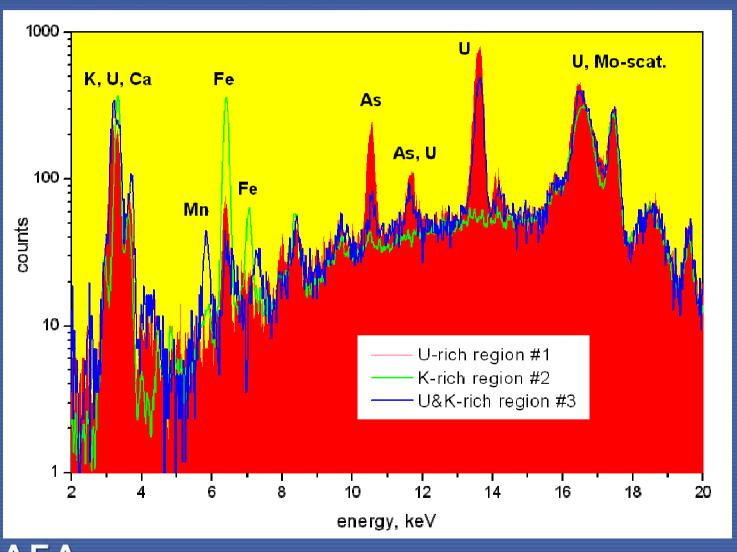




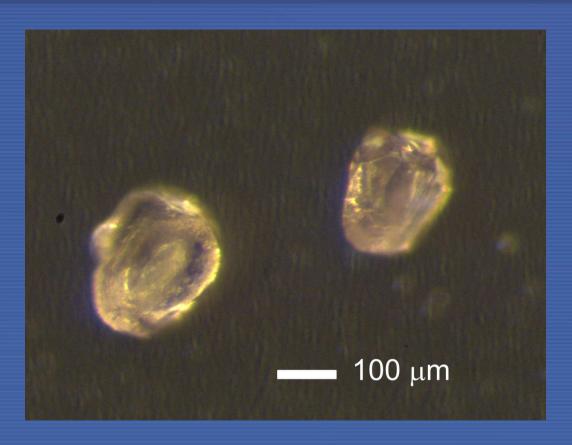








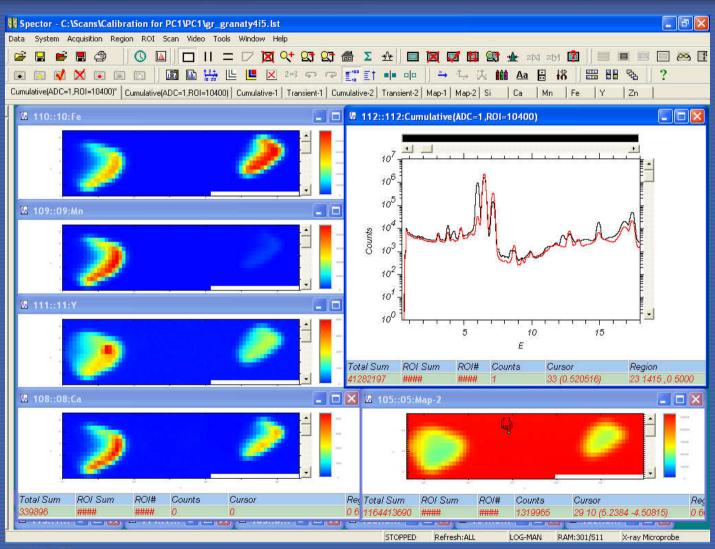
3. Identification of minerals



Two "similar" garnets: almandine-spessartine (left) and almandine (right)

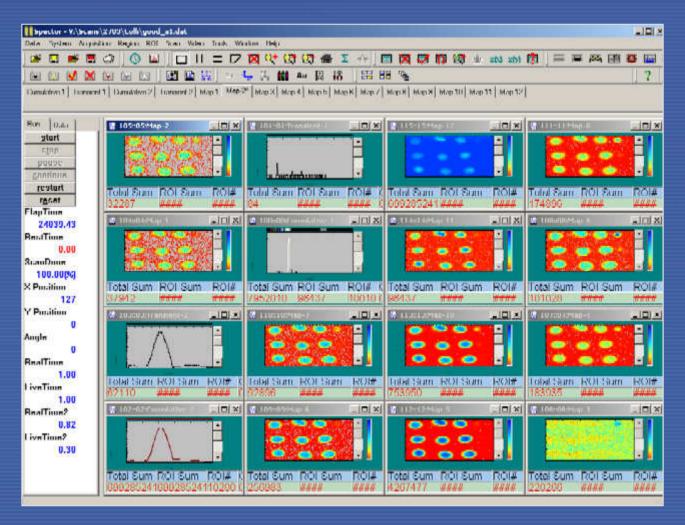


3. Identification of minerals



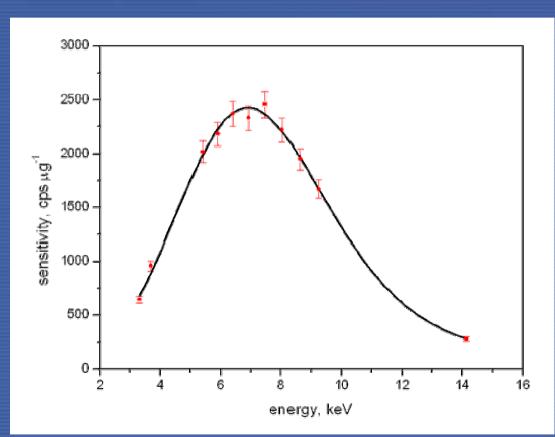
- Based of the cumulative X-ray fluorescence spectrum of the MERCK CertiPUR standard the sensitivity curve is established. The thickness of the standard residue, confirmed by the data from transmission measurements, was well within the "thin sample" approximation – the sensitivity values were calculated assuming a "thin sample" model.
- The sensitivities obtained in step 1 are used to calculate the element concentrations in the dried residue samples prepared from the candidate RM. The obtained concentrations are corrected for absorption, using the data from transmission measurements.







Determination of the sensitivity calibration curve:



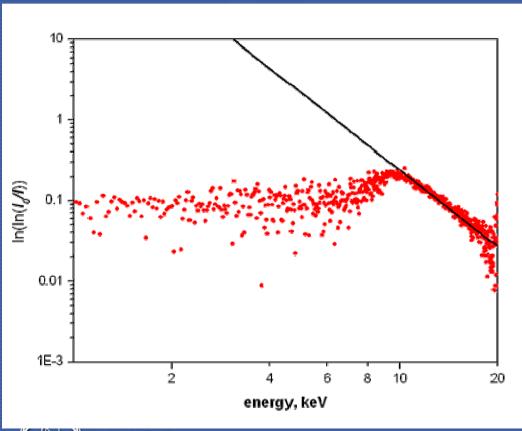
$$(I)_S = S(Z)m$$

$$(I)_{x} \approx S(Z_{x})m_{x}F(E_{eff},E_{x})$$

$$c_{x} = \frac{m_{x}}{m_{u}}$$



$$F(E_{eff}, E_x) = \frac{1 - \exp\{-((\mu \rho d)_{eff} + (\mu \rho d)_x)\}}{(\mu \rho d)_{eff} + (\mu \rho d)_x}$$



 Estimation of the absorption correction factor



	Known Concentration	Determined Concentration
Ti	0.884% (0.082)	0.82 % (0.055)
Cr	352 ppm (22)	288 ppm (50)
Mn	1757 ppm (58)	1670 ppm (90)
Fe	7.91 % (0.24)	6.8 % (0.3)



4. CT Principle – Parallel Beam Projections

2D/3D CT scan (absorption and/or fluorescence)

Absorption correction of the fluorescence sinograms

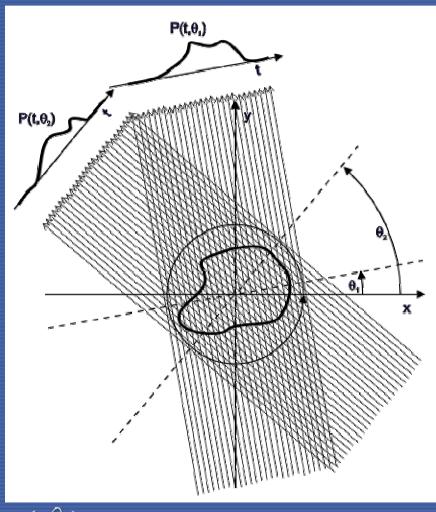
Construction of interpolated sinograms for 3D scans

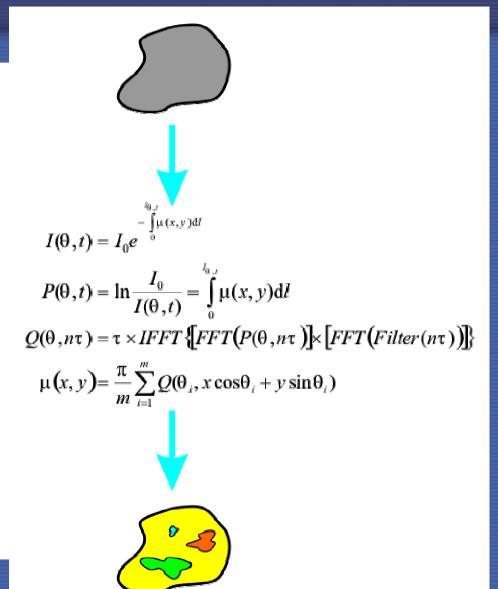
Reconstruction of the object cross-section(s) by filtered back projection algorithm

Volumetric rendering, 3D reconstruction, 3D mapping of element distribution, volume and area measurements



4. CT Principle – Parallel Beam Projections

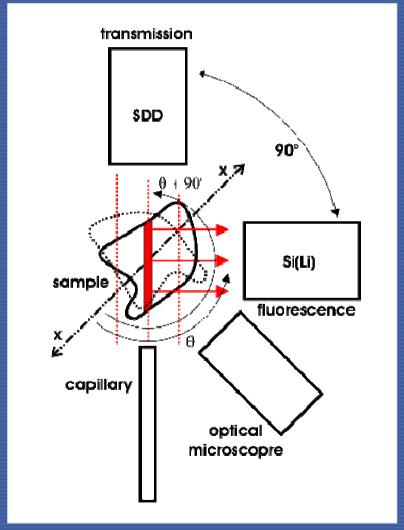






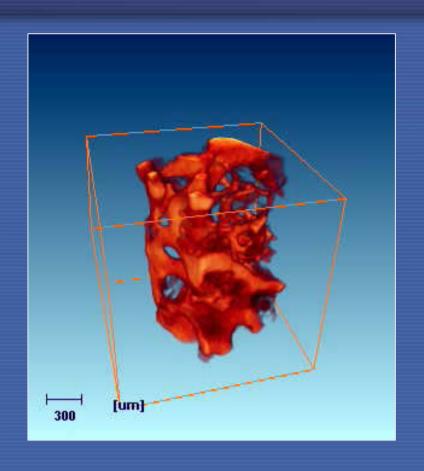
4. CT Absorption and XRF Imaging

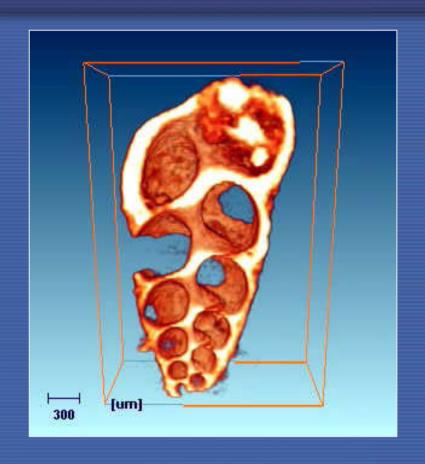
Simultaneous X-ray absorption and X-ray fluorescence imaging in a "pencil" beam geometry





4. CT Absorption and XRF Imaging

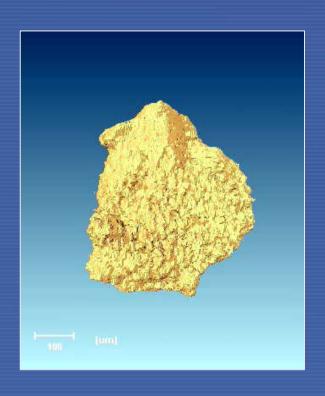


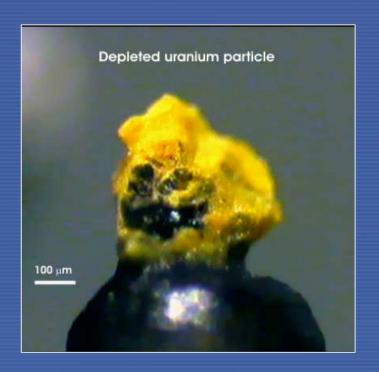


Reconstructed volumetric models of an osteoporotic bone fragment (left) and a shell fossil (right). CT absorption scan mode.



4. Depleted Uranium Particle

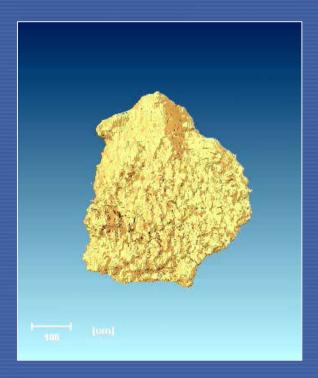


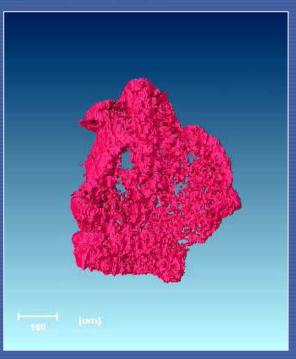


Reconstructed volumetric model (left) of a DU grain (right) obtained in absorption mode



4. Depleted Uranium Particle





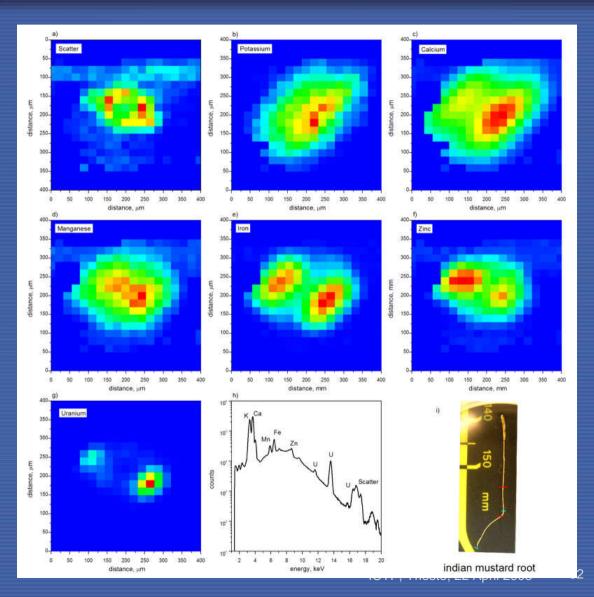


Reconstructed volumetric models of a DU grain (left to right): absorption, U-La not corrected for absorption and U-La corrected



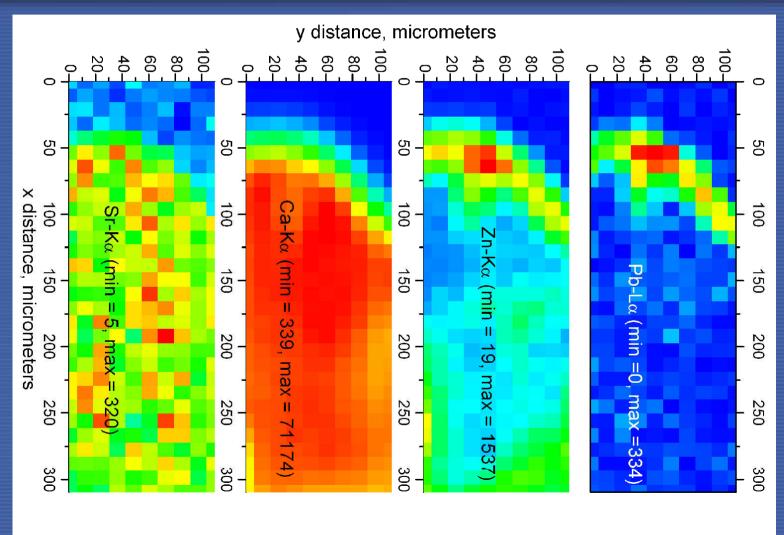
5. Depth profiling of element distribution with confocal setup

a) - g) distributions of the intensities of scatter peaks, K-Ka, Ca-Ka, Mn-Ka, Fe-Ka, Zn-Ka, and U-La obtained from cross-sectional confocal X-ray fluorescence scan through the root tip. Scan parameters: step size dx = 22.124 μ m, dy = 20.000 μ m, scan size 19x21 pixels, spectra acquisition time per pixel = 500s for "live pixels", 1 s for others, spatial resolution of the confocal setup FWHM = 60 μm, Mo-anode X-ray tube operated at 45kV/40 mA; h) cumulative X-ray spectrum (sum of all individual pixel spectra) collected during the confocal scan; i) photograph of the indian mustard root.





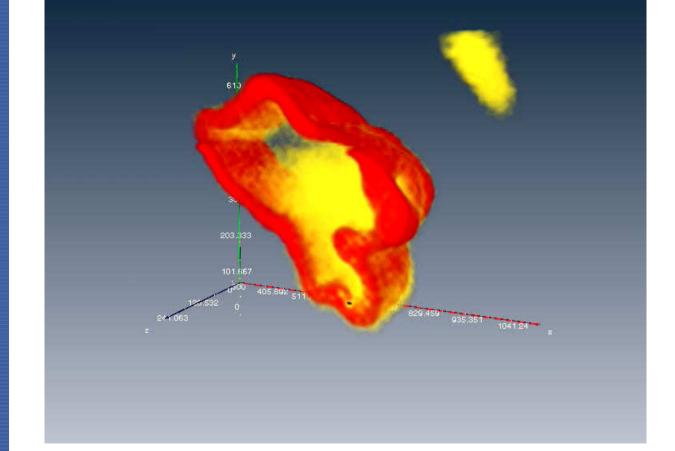
5. Distribution of elements in human bone obtained by confocal scanning





5. Fe-rich plaque covering rice root

red – iron yellow – potassium





6. Sample preparation procedures

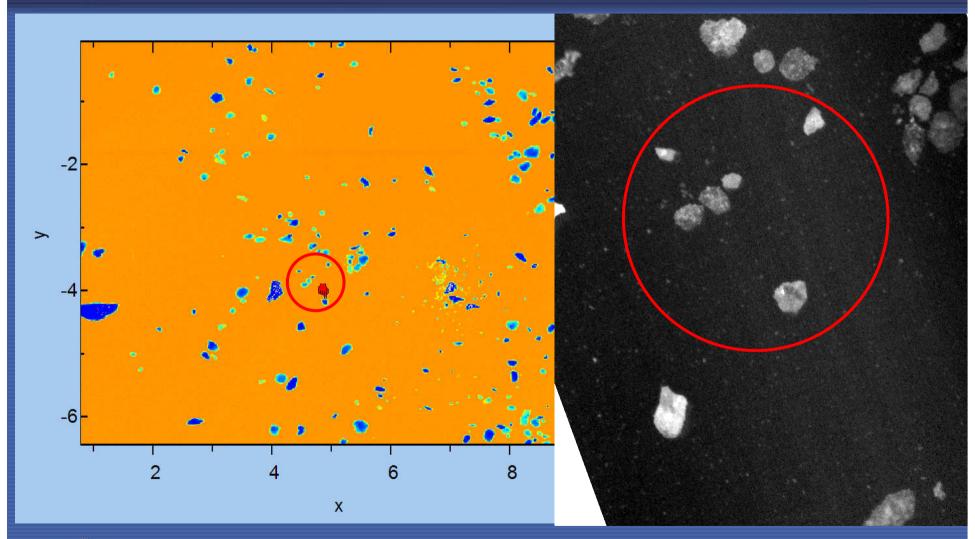
For micro-analytical techniques (micro XRF and micro-tomography)

Individual particles

Biological materials

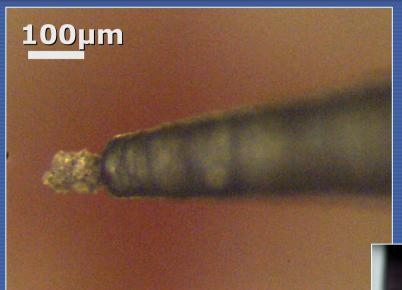


6. Individual Particles





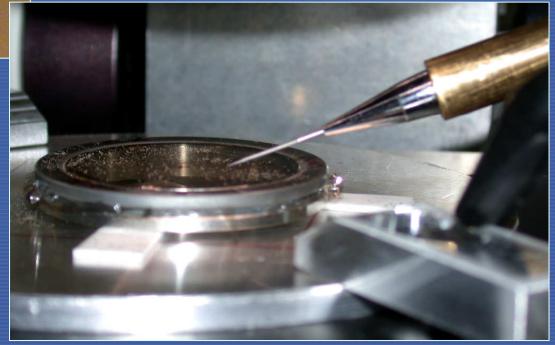
6. Individual Particles



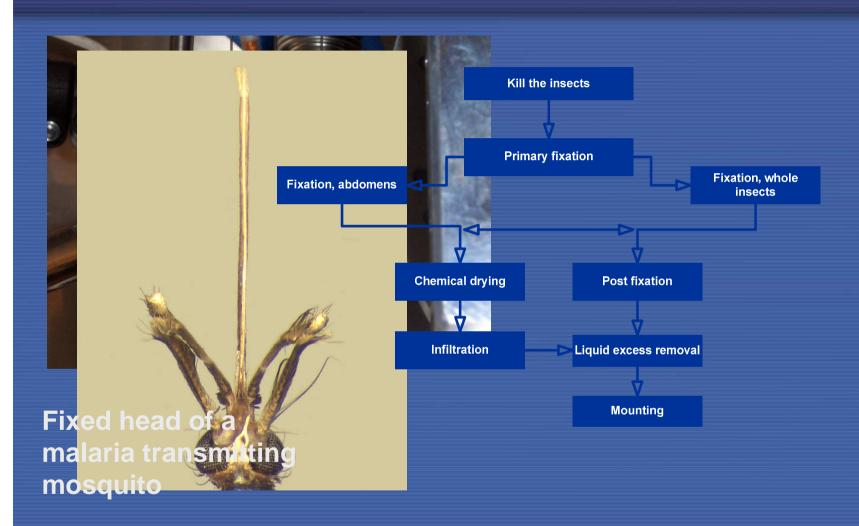
Loading the needle with a selected POI in the µ-manipulation system

DU particle from a contaminated soil, loaded on a sharpened graphite needle



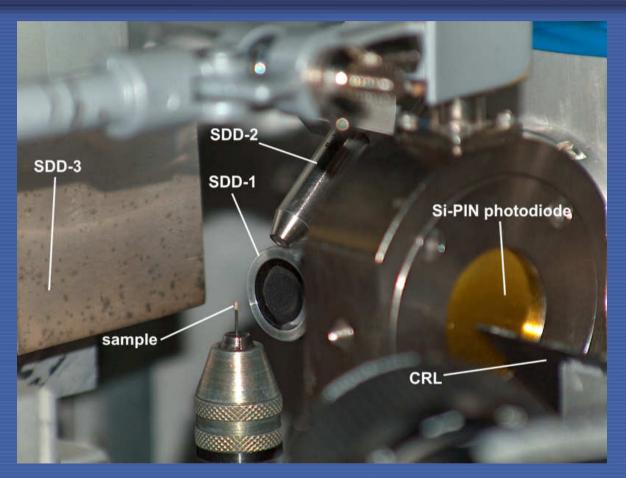


6. Biological Materials





7. Micro-beam X-ray spectrometer installation at the ANKA synchrotron Fluo-Topo beamline



The IAEA micro-beam X-ray scanning spectrometer setup installed at the ANKA Fluo-Topo beamline, FZK Research Centre, Karlsruhe, Germany.



7. Angstrom Quelle Karlshure (ANKA)

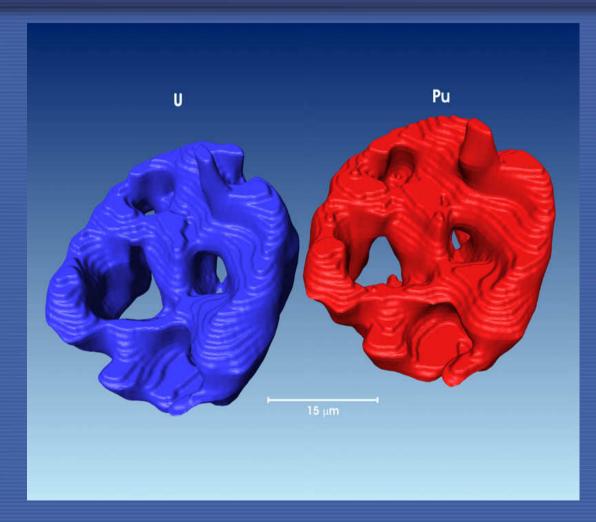
The IAEA set-up at the synchrotron beamline







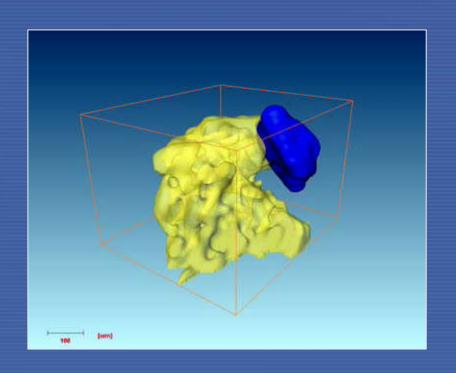
7. X-ray fluorescence micro tomography

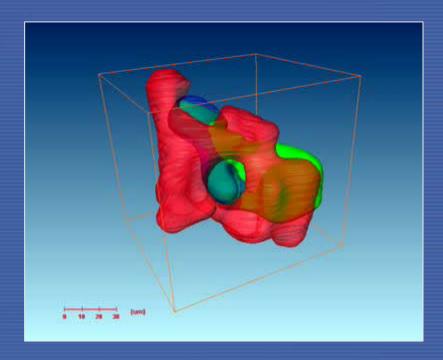


3D reconstruction of the spatial distributions of uranium and plutonium in a U/Pu-rich particle.



7. Pu/U-rich particles – X-ray tomography

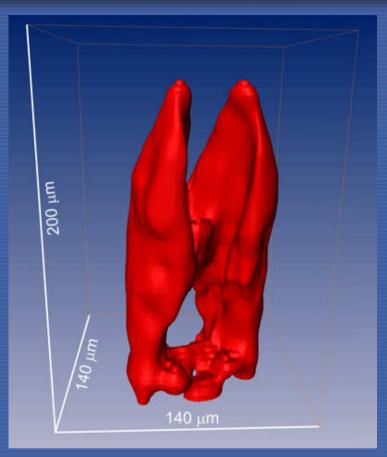




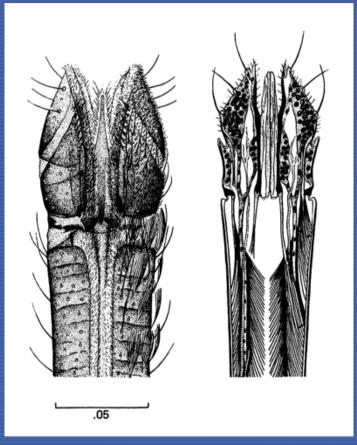
Reconstructed volumetric distributions of elements in individual "hot particles": left - plutonium rich particle (blue) attached to a coral matrix (yellow) and right — a U/Pu rich particle (green/blue) embedded in sediment matrix (red)



7. X-ray fluorescence micro tomography

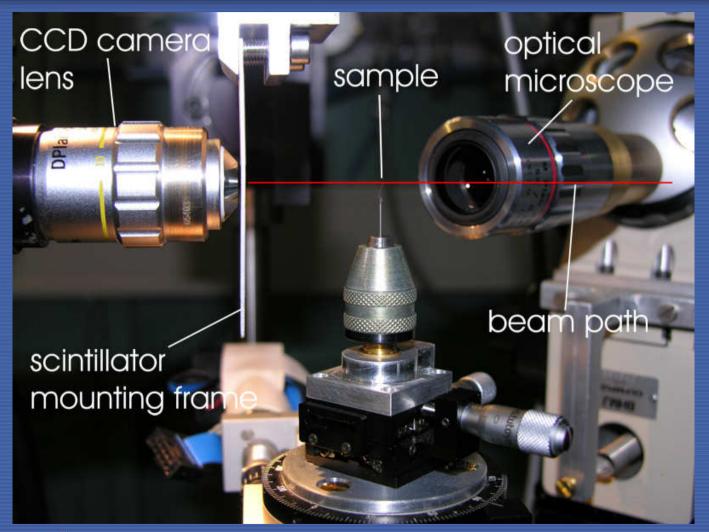


AEA



Left: tomographic reconstruction of the last segments of the proboscis of a female mosquito (Anopheles); right: anatomical drawing of Aedes Aegypti, adopted from Jobling, Boris and David James Lewis, Anatomical Drawings of biting flies, British Museum of Natural History, London, in association with the Wellcome Trust, 1987. The filtered backprojection algorithm was used to reconstruct the uranium-fixed proboscis of a female mosquito.

7. X-ray phase-contrast enhanced micro tomography

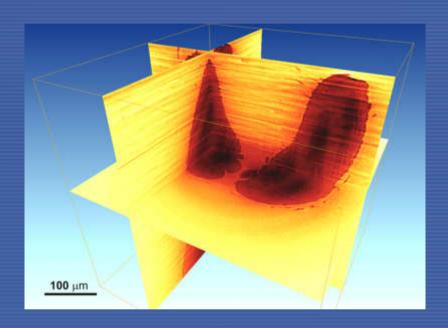


Experimental setup established at the ANKA synchrotron Fluo-Top beam line for investigating the morphology of malaria transmitting mosquitoes



7. X-ray phase-contrast enhanced micro tomography



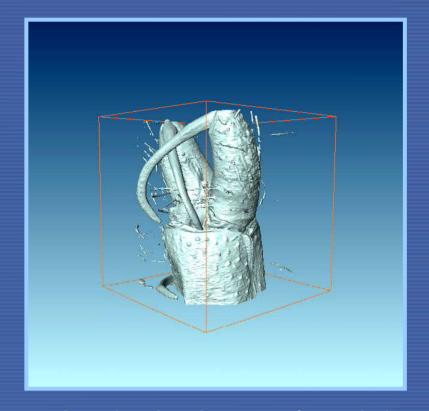


Left: single X-ray phase-contrast enhanced projection image of an abdomen of irradiated male mosquito; right: reconstructed volumetric cross-section through the mosquito abdomen



7. X-ray phase-contrast enhanced micro tomography





Left: single X-ray phase-contrast enhanced projection image of an abdomen of irradiated male mosquito; right: reconstructed volumetric model through the mosquito abdomen



7. Phase-contrast X-ray imaging of live specimens





8. Training

Model XRF laboratory for basic and advanced training

- Training to develop practical skills
- Develop analytical procedures for special applications in IAEA MSs
- Research & development
- QA/QC procedures incl. uncertainty budget



9. Summary and conclusions

- Integration of R&D with training and direct analytical services to IAEA MSs
- Extension of applicability range and improvement in analytical performance of XRF are major concern
- Emphasis on <u>versatility</u> of XRF (important for XRF labs in developing countries)
- Emphasis on <u>usefulness</u> of the results for XRF laboratories



9. Summary and conclusions

- X-ray tube based micro-analytical techniques are well suited for characterization of minute samples, and heterogeneous objects.
- Quantitative analysis is possible but more difficult as compared to the analysis of bulk samples.
- When combined with synchrotron radiation very low amount of substance can be detected, coherent beam allows performing phase-contrast imaging of weakly absorbing objects, e.g. tissue, biomedical samples.



Acknowledgements

The results presented were obtained with collaboration of:

- D.Wegrzynek, E. Chinea-Cano, S. Bamford, C. A. Malcolm and M. Helinski, IAEA Laboratories Seibersdorf
- P. Wobrauschek, C. Streli, N. Zoeger, Vienna University of Technology, Atominstitut, Austria
- M. Eriksson, M. Betti, S. Toeroek, European Commission Joint Research Center, Institute for Transuranium Elements, Karlsruhe, Germany
- R. Simon, S. Staub, T. Weitkamp, A. Rack, Forschungszentrum Karlsruhe GmbH, Institute for Synchrotron Radiation, Eggenstein-Leopoldshafen, Germany



THANK YOU

E-mail: A.Markowicz@iaea.org

