



**The Abdus Salam
International Centre for Theoretical Physics**



1936-40

**Advanced School on Synchrotron and Free Electron Laser Sources
and their Multidisciplinary Applications**

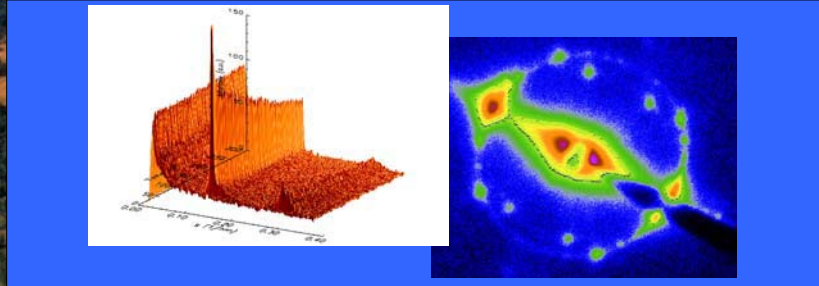
7 - 25 April 2008

SAXS under extreme conditions: Nanomaterials and proteins.

A. Amentisch
Austrian Academy of Sciences.

SAXS under extreme conditions: Nanomaterials and proteins

By H.Amenitsch (amenitsch@elettra.trieste.it)



Sub-millisecond Exposures for Time-Resolved Structure Analysis
on Synthetic and Biopolymers, Colloids, Ceramics, Glasses and
Alloys



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Layout of Presentation

Time-Resolved Measurements:

- How to Trigger the Reactions
- Biological Samples
Muscle, Proteins, Phospholipids
- Material Science
Nanoparticles
Mesoporous Materials

Grazing Incidence Small Angle Scattering (GISAXS/GISAD):

- Surface Diffraction Lipids
- Mesoporous Materials
- Nanoparticles/Nanocrystals



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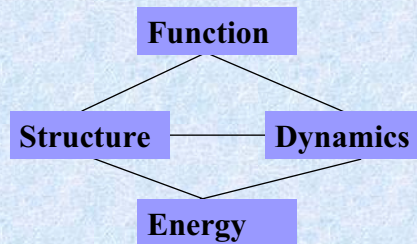


Why?

What is extreme?

- Temperature: mK, 10^3 K, 10^6 K
- Time scales: years, s, ms, μ s
- Pressure: MPa, GPa
- Chemical potential
- Non equilibrium states $\Rightarrow$ Transitions

Scientific Case:



Biology and Biomedicine:

- understand molecular and cellular function
- find ways to cure diseases

Material Science:

- understand macro- and supramolecular assembly
- find new, purpose-designed materials



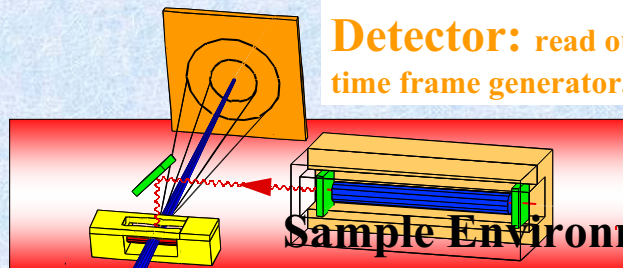
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What do you need?

Detector: read out times,
time frame generator, cps, efficiency



Sample Environment!!!

Optics (10^{12} - 10^{14} ph/s)



Source



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How to trigger transitions?

- T-jump (heating): Erbium Glass Laser (ms)
“heat exchanger”
- T-cool jump: “heat exchanger” (s)
- p scans (jumps): High pressure cells (ms)
 - hydrostatic pressure
 - diamond anvil cells
- Stopped-flow cells, jet-cells (μ s-ms)
- M.C.Ramachandra et.al. Biophysical Journal 74, (1998), 2714
- Segel DJ et.al. , JOURNAL OF MOLECULAR BIOLOGY, 288, 489, (1999)
- Pollack L et.al., PNAS, 96,10115, (1999)
- Akiyama S et.al., PNAS, 99,1329, (2002)
- Rössle M et.al. *Biomacromolecules*, 4, 981-986, (2003)
- Batch reactor(s)
- Magnetic field
- Shear experiments
- Mechanical stretch (μ s-ms-s)

How it all began - Muscle Contraction

September 1970: DESY
Rosenbaum, Holmes & Witz, *Nature* (1971), 230,434

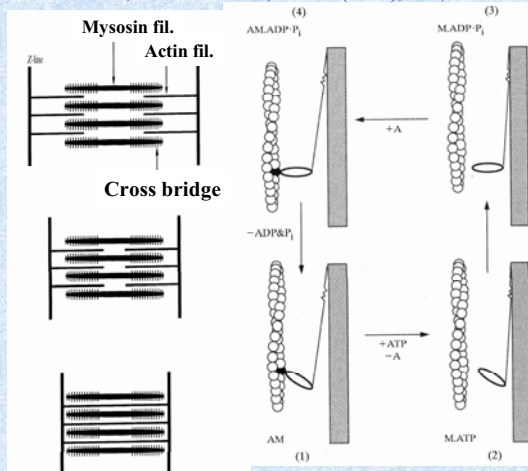


Fig. Muscle Contraction thick (myosin)-, thin-(actin) fibers are interdigitating K.C Holmes *Acta Cryst. A*54, (1997), 789

Fig. Lymn-Taylor cycle. (Lymn, Taylor *Biochemistry*, (1971)10, 4617 Myosin-cross-bridge is bound in rigor (1) ATP binds->quick dissociation (2) ATP->ADP + P (hydrolysis) binding of myosin to actin 90 up (3) release of components, rowing to (1)

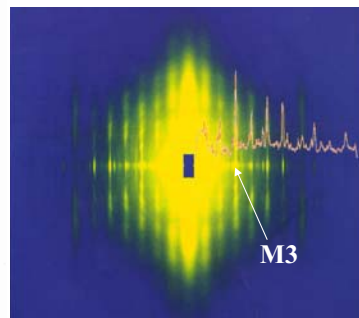
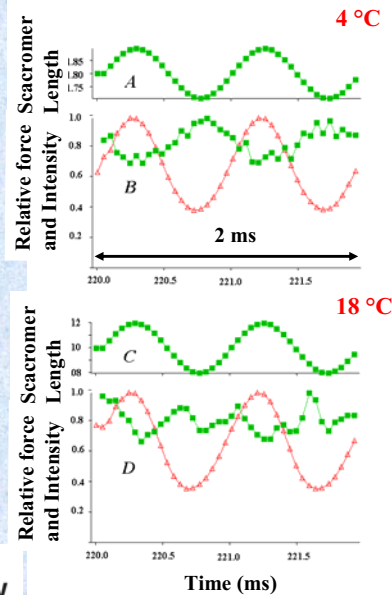


Fig. Diffraction pattern of life skeletal frog muscle Cover page: Yagi, et.al. *J.Synchrotron. Rad* (1996), 3, 247

How it all began - Muscle Contraction



Rana temporaria (tibialis anterior fibers)
Frequency: 1 kHz
Time resolution: 50 μ s

Frequency: 3 kHz
Time resolution: 16 μ s

Literature:

Bagni MA, et.al., BIOPHYSICAL JOURNAL
80, 2809, (2001)

Griffiths P J, et.al. PNAS, 99, 5384, (2002)

Piazzesi G, et.al. NATURE 415, 659, (2002)

Reconditi M, et.al. NATURE 428, 587, (2004)



- BEAMLINE AT ELETTRA
Bernstorff, M. Rappolt & P. Laggner



Biomechanics on Human Aorta: Motivation

Macroscopic
response

Model

Micro- and nano-
structure

Pathology, Clinics

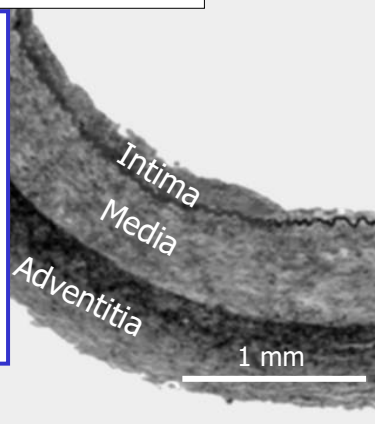
Characterization of vascular disease
Effects of aging
Identification of therapeutic targets (Balloon
Angioplasty)

Graft design

Biomimetic materials

Functional tissue engineering

Mechanobiology



Cross section of a human artery



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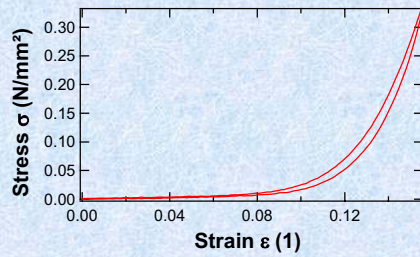
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Human Aorta: Mechanical Parameters

Macroscopic

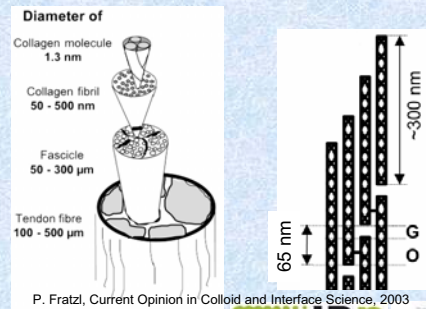
- geometric deformation
- stress
- strain



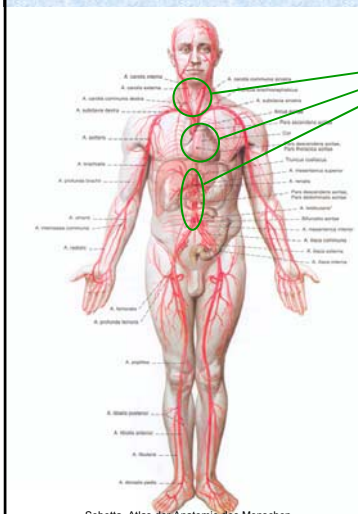
Nanoscopic

- fiber – matrix composite
- fiber alignment
- fiber strain

Collagen - The most abundant protein



Human Aorta: Sample Preparation



An artery, cleaned from
surrounding tissue

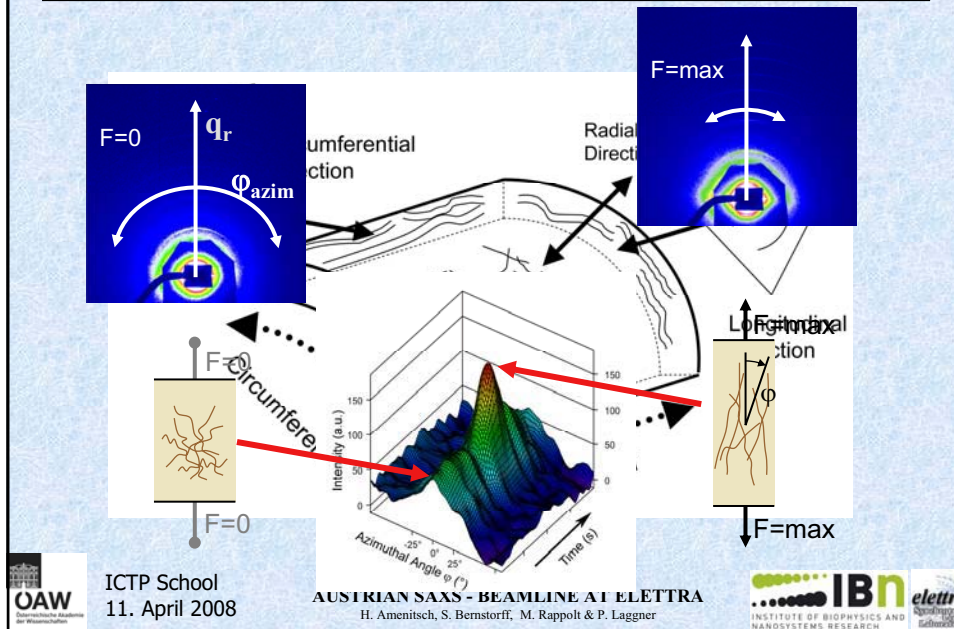


After dissection into
its major layers

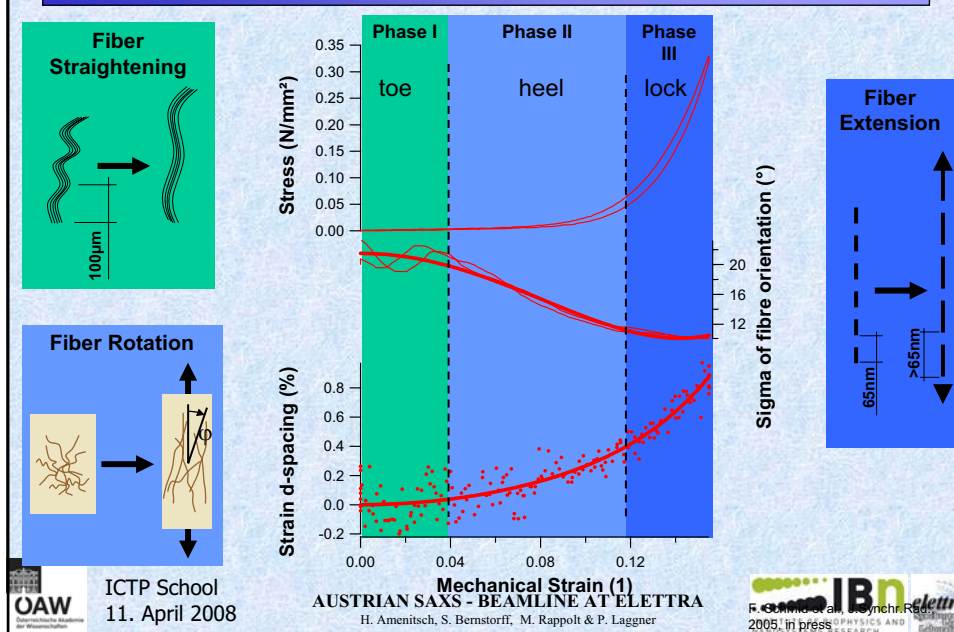


The final
sample

Human Aorta: Collagen Fiber Orientation



Nano – Macro Coupling



T-jumps: Phospholipid Phase Transition

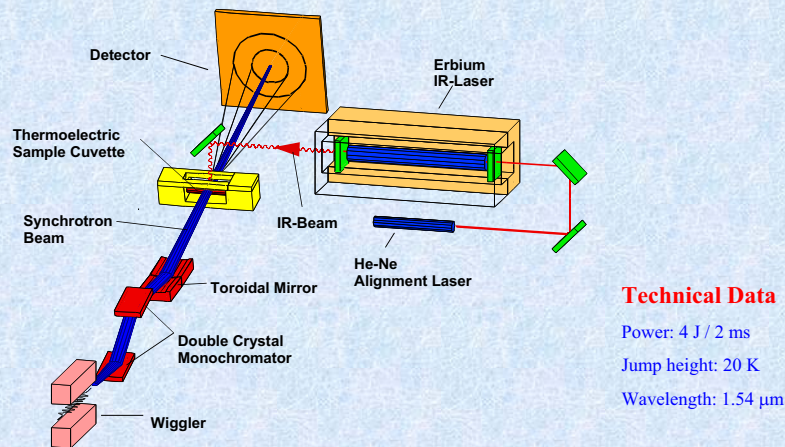


Fig. Experimental set-up of the T-jump device at the SAXS beamline (ELETTRA).

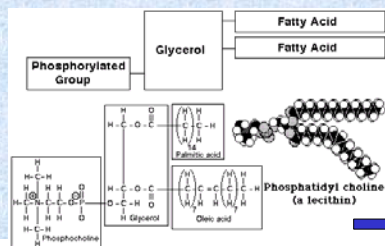
What is a biological / model membrane ?

The boundaries of cells are formed by biological membranes, the barriers that define the inside and the outside of a cell.



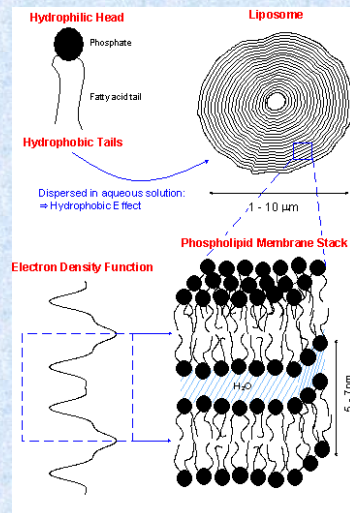
Phospholipids are the major components of biological membranes that form the structural matrix into which proteins are imbedded.

STRUCTURE



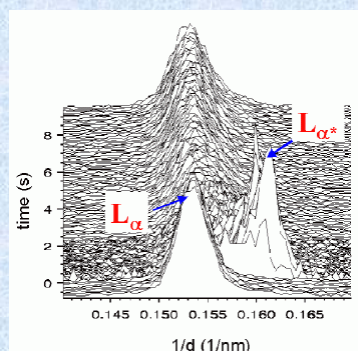
In aqueous solution: self assembly into, e.g., unilamellar vesicles

T-jumps: Phospholipid Phase Transition



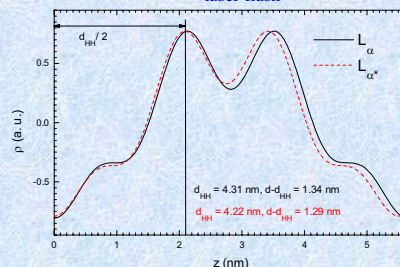
The formation of a phospholipid membrane. Phospholipids aggregate spontaneously into ordered supra-molecular structures in the presence of water. This can be explained in simple terms by the fact that phospholipids feature a hydrophilic headgroup (attracting water) and hydrophobic hydrocarbon-chains. The average 1- dimensional repeat distance d , i.e., bilayer plus waterlayer of the depicted liquid crystalline phase (L_α) is in the range of 5-7 nm. The electron density distribution of a bilayer (bottom left corner) has maxima in the headgroup regions and a minimum at the methyl terminus of the hydrocarbon-chains. The dashed rectangle marks the part of the electron density distribution shown in the fig below.

T-jumps: Phospholipid Phase Transition

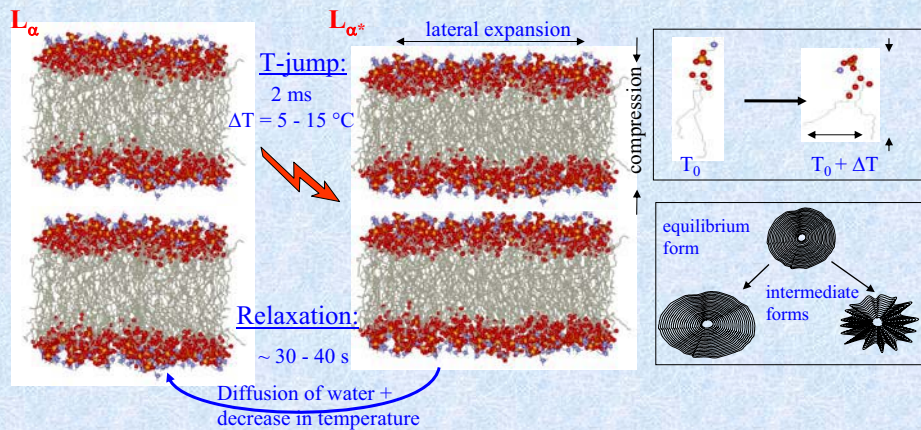


The first order diffraction peaks of a phospholipid sample during a T-jump experiment (time resolution = 5 ms). The IR-laser was triggered at time zero.

Superimposed electron density distributions of the original L_α -phase (straight line) and of the intermediate phase L_{α^*} (dashed line) immediately after the laser flash

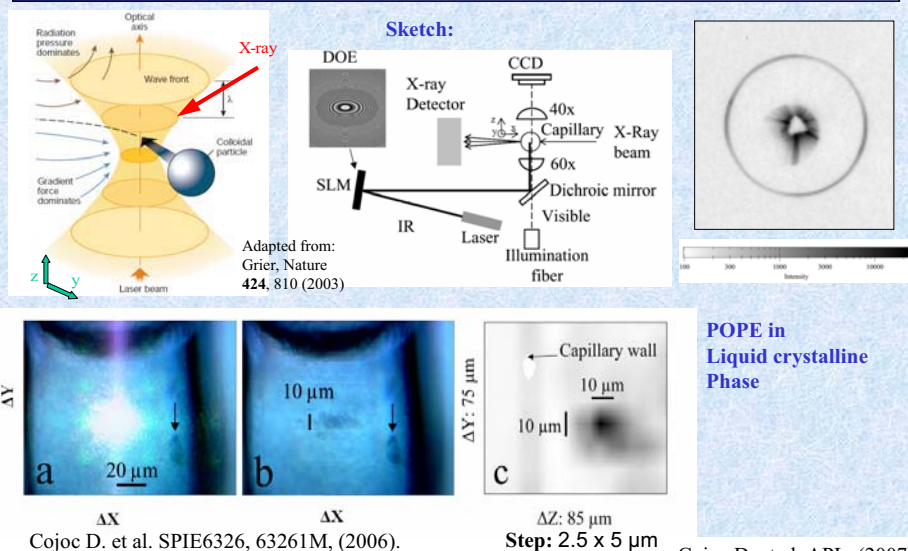


T-jumps: Phospholipid Phase Transition



G. Pabst, M. Rappolt, H. Amenitsch, S. Bernstorff & P. Laggner, *Biophys. J.*, (2000)

Current Developements: Optical Tweezer



Mesoporous Materials: Bulk MCM-41

In-situ study of the Formation of the MCM-41 Structures using liquid crystal templating mechanism

P. Ågren, *J. Phys. Chem. B*, (1999), 103, 5943

Aim:

Influence of the co-surfactant and its concentration on the phase behaviour of the TEOS synthesis.

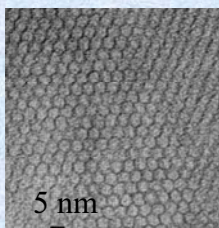


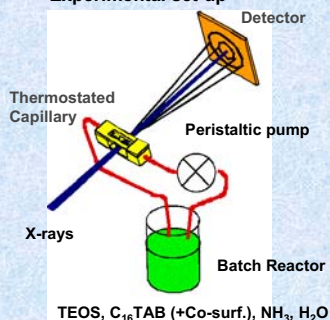
Fig. Representative electron transmission micrograph of a MCM-41 structure depicting the mesoporous hexagonal nano-structure.

Industrial applications: -adsorbents

-ion-exchangers

-catalysts

Experimental set-up



TEOS Tetraethylorthosilicate
C₁₆TAB Hexadecyltrimethylammonium bromide



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Hosts for technologically advanced materials

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Mesoporous Materials: Bulk MCM-41

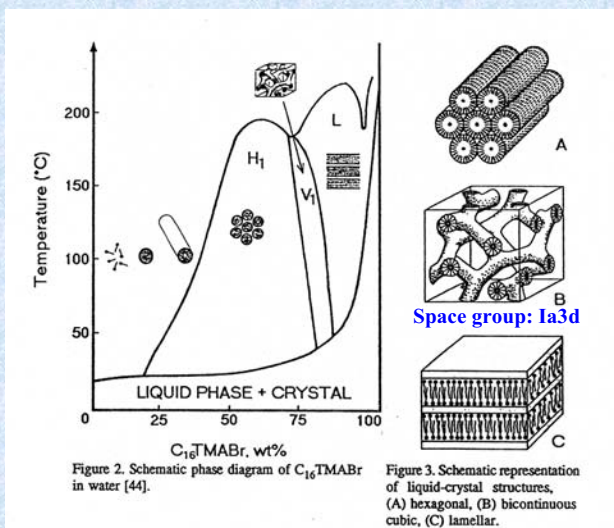


Figure 2. Schematic phase diagram of C₁₆TABr in water [44].

Figure 3. Schematic representation of liquid-crystal structures, (A) hexagonal, (B) bicontinuous cubic, (C) lamellar.

MCM-41

MCM-48

MCM-50

From: Sayari,
Studies in Surface
Science and
Catalysis (1996),
Vol 102, 1



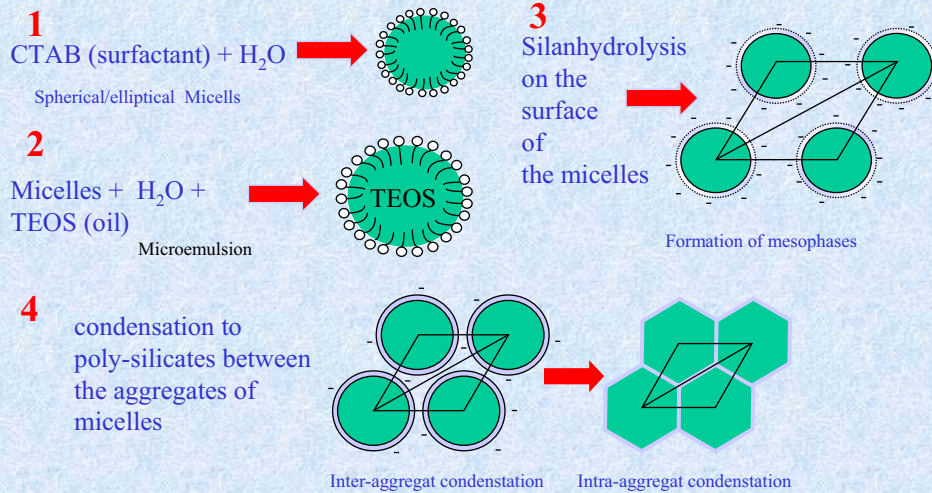
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Model for synthesis of mesophases in the system: TEOS/CTAB MCM-41



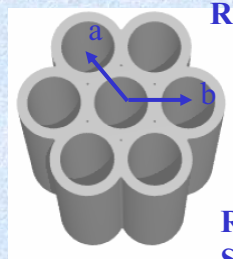
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How to obtain real space information?

Hex:
P6m

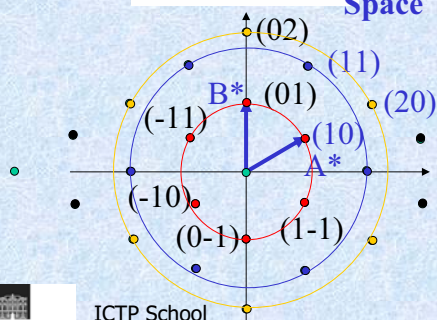


Real Space

$$\rho(\vec{r}) = \rho_{average} + \sum_{\vec{q}} A_{\vec{q}} \cdot \cos(\vec{q} \cdot \vec{r})$$

$$|\vec{q}| = \frac{4 \cdot \pi}{\sqrt{3} \cdot a} \cdot \sqrt{h^2 + k^2 + h \cdot k}$$

Reciprocal Space



$$I(|\vec{q}|) = K \cdot M(h, k) \cdot |A_{\vec{q}}|^2 \cdot |\vec{q}|^{-2}$$

$$\pm A_{\vec{q}} \rightarrow ?$$

(10)	6
(11)	6
(20)	6
(21)	12

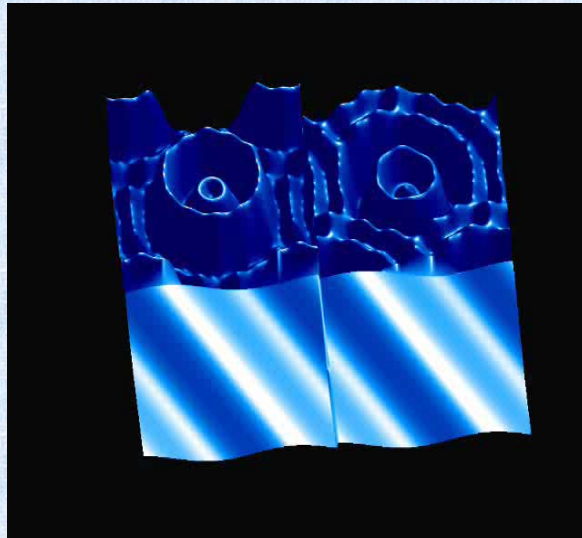


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How to obtain real space information?



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Mesoporous Materials: Bulk MCM-41

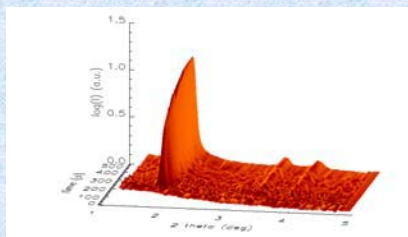
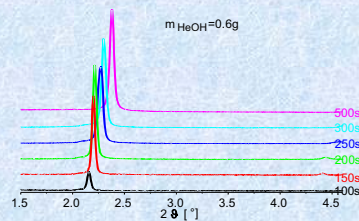
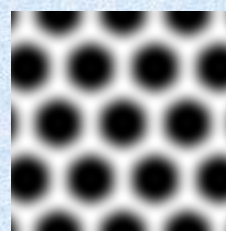


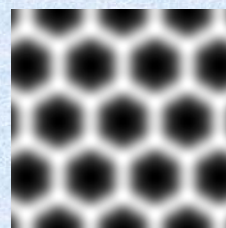
Fig. : Time-resolved diffraction pattern of the TEOS synthesis
Time resolution: 0.3 s/frame, Transition: micellar solution -
ordered phases (standard synthesis: hexagonal D = 4.67 nm)



Calculated Electron Density



After 150s



Final Structure



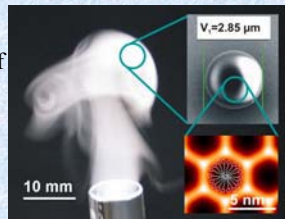
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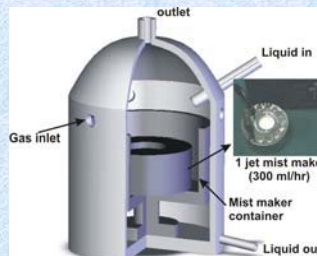


SAXS in Gasphase – Aerosol Synthesis of Mesoporous Materials

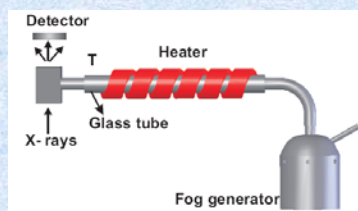
Evaporation
Induced Self
Assembly
(EISA)
J.Brinker
et.al



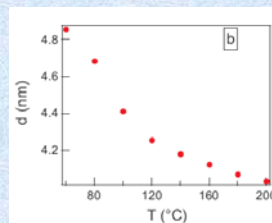
Boississiere, C., et.al. 2003. *Chemical Communications*



Aerosol Generator



Aerosol Setup



Results

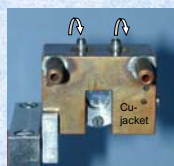
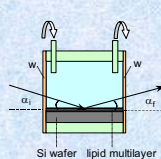


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I. Shyjumon et al., *Rev. Sci. Instrum.*, (2008)
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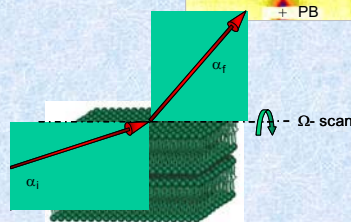
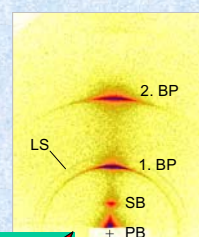


Surface Diffraction Lipids – Surface Chemistry



Sketch and photograph of the
sample cell in transmission
geometry for GISAXS.

Sketch of the exp. set-
up and 2D diffraction
pattern of POPC and
0.5 M LiCl

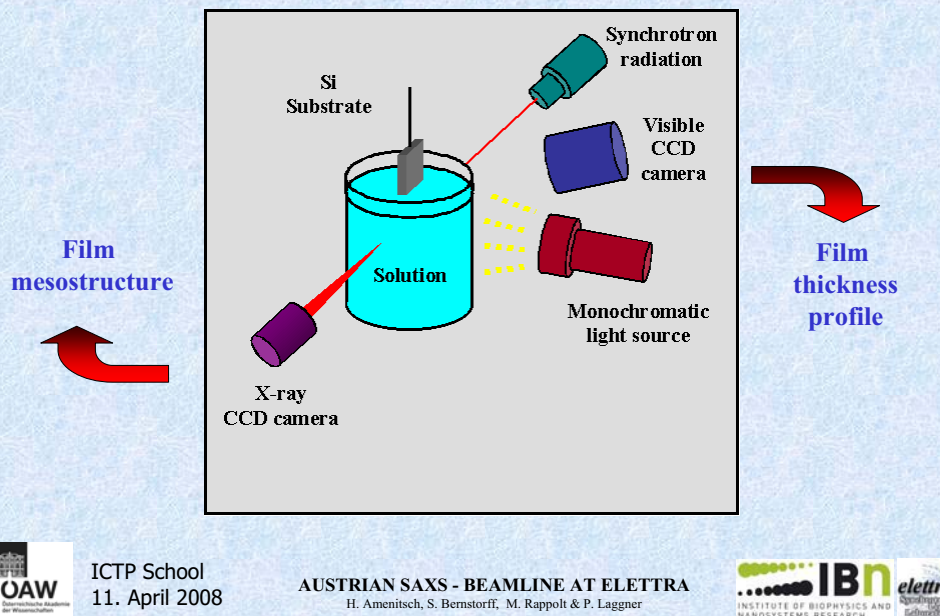


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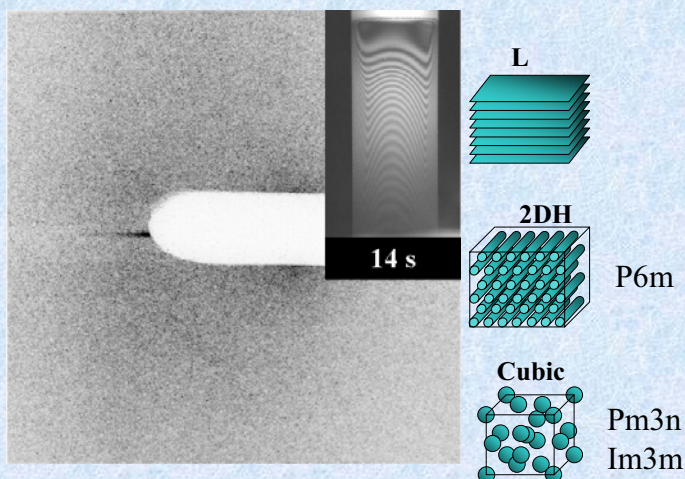


The Self-Assembly of thin films by *in situ* SAXD and interferometry

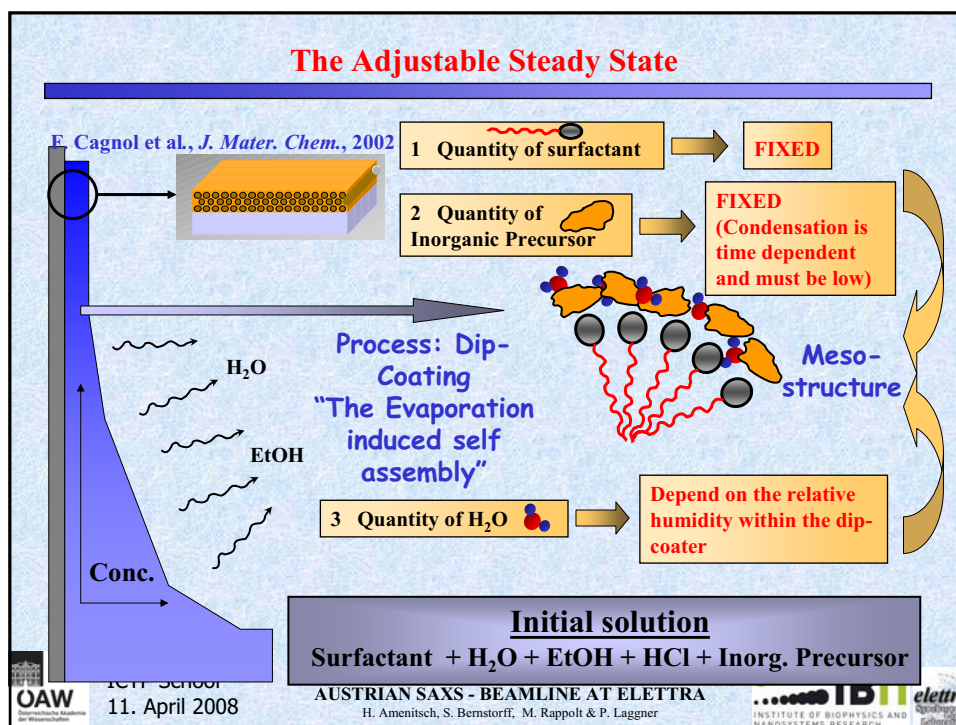


Surface diffraction: Formation of aligned mesoporous thin films

CTAB / Si = 0,18
 H_2O / Si = 5
 HCl / Si = 0.15
 Ageing time
 Relative Humidity



Grosso D, et.al., CHEMISTRY OF MATERIALS 14, 931,(2002)
 Grosso D, et.al., Nature Materials, (2004)



Further Reading

Annual Reports SAXS Beamline:

- Time resolved studies
- Surface diffraction
- GISAXS

SAXS:

Books: Guinier
Glatter, Kratky
Feigin, Svergun

SAXS Conferences: every 3-4 years

Software: D.Svergun, **ATSAS**, O.Glatter, **GIFT**
J.Ivlasky, **Basic Suit**

Surface Diffraction/GISAXS

Book: M. Tolan, X-ray scattering from soft-matter thin film. Springer Tracts in Modern Physics 148, Springer, Berlin, 1999.

Paper: Rauscher M, Salditt T, Spohn H
GISAXS –Distorted Born Wave Approximation
PHYS REV B 52, 16855 (1995)

Software: Lazzari R, ISGISAXS: program, J APPL CRYSTALLOGR 35: 406, (2002)
http://www.esrf.fr/computing/scientific/joint_projects/IsGISAXS/isgisaxs.htm

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elettra

Conclusion

Biology: Single Muscle Fiber Diffraction
Human Aorta
Biomembranes

Nucleation and Growth

Mesoporous Materials: Bulk, Surfaces

-Outlook: USE of NEW DETECTORS!
Use of coherence in SAXS!
(photon correlationspectroscopy, single particle imaging)
Use of new sources FEL's, new nm-beams
Think for yourself of new ways
to use SAXS and SR

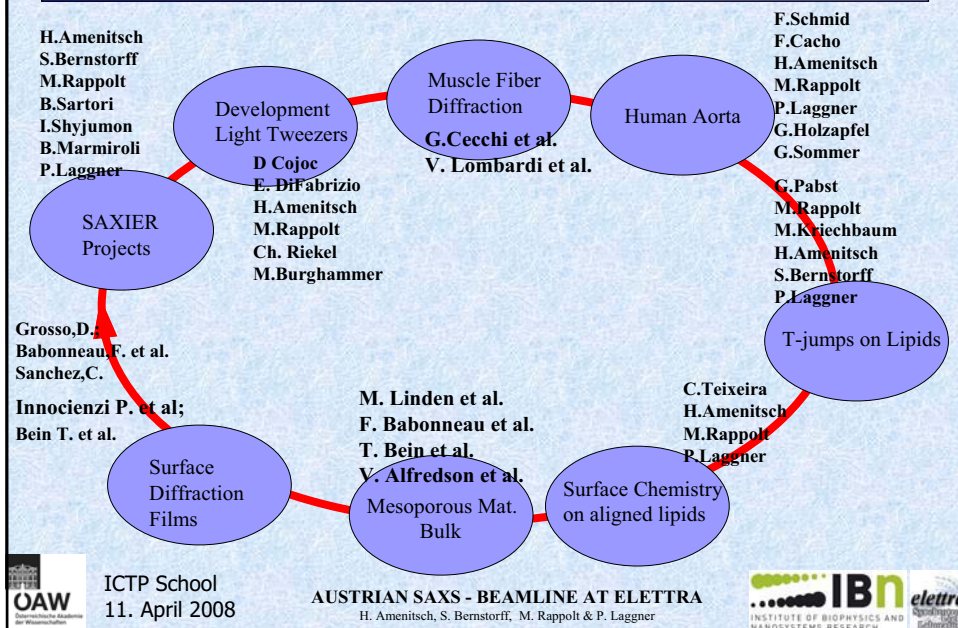


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Acknowledgement



Acknowledgement

