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QM and Hybrid QM-MM Simulation of Biomolecules (Computer Simulation of Ligand Binding and Reactivity of Heme Proteins) Part III

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Computer simulation of ligand binding and reactivity of heme proteins

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In this final talk, we will discuss two complex and interesting problems:

Multiligand chemistry: the M. Tuberculosis TrHbN case

Reaction mechanism in indoleamine and tryptophan dioxygenases

Truncated hemoglobins

Truncated Hb family found in the 90': sequences 20-40 residues shorter than classical globins

Distributed in bacteria, unicellular eukaryotyes and higher plants (no in animals!)

Tertiary structure based on a 2-on-2 \propto helical sandwich (trimmed modification of classical 3-on-3 fold.

Despite minimal size, they may display an inner cavity/tunnel system

TrHbN from M.T. Multiligand chemistry: NO produced by the immune system maybe inactivated by the oxygenated Hb: toxic innocuous NO + Fe(II) $O_2 \rightarrow Fe(III) + [NO_3]^-$ Relevant process in physiological (Hb, Mb) and pathological processes (it is a problem in the design of synthetic hemoglobins)

Experimental kinetic information

Fe(II) + $O_2 \rightarrow Fe(II)O_2$ $k_{on} = 25 \,\mu M^{-1}s^{-1}$ Involves oxygen entry and formation of Fe-O₂ bond

NO + Fe(II) $O_2 \rightarrow Fe(III) + [NO_3]^{-1}$ $k_{ox} = 745 \ \mu M^{-1} s^{-1}$ Involves NO entry and chemical reaction Difficult to understand

Reaction mechanism:

O₂ migration
O₂ binding
NO migration
Reaction of NO with O₂
Product release
Protein reduction

(classical MD) (QM-MM) (classical MD) (QM-MM) (QM-MM and classical MD)

Key reaction: NO + Fe(II) $O_2 \rightarrow Fe(III) + [NO_3]^{-1}$

2 channel system proposed on the basis of inspection of x-ray results (Bolognesi's group at Milano) But how do O_2 and NO migrate?



Long (100 ns) classical MD (Amber 9) of oxy and deoxy proteins indicate PheE15 acts as the long tunnel gate, showing two conformations (open and closed). O_2 coordination "opens" the tunnel for NO migration

Umbrella sampling free energy profiles for channel opening



Long channel closed almost always in deoxy protein. Oxygen binding induces channel opening for second ligand. (NO)!!

A. Bidon Chanal et al, Proteins (2006), 64, 457-464.



Long channel closed (deoxy) vs open (oxy) NO entry to oxygenated protein Short channel closed (oxy) vs open (deoxy) O_2 entry to deoxy protein.

Why does this happen?



GlnE11 moves away upon O₂ binding, pushing PheE15, due to competition in H bonding of oxygen (which carries a significant negative charge)

Oxygen affinity Hydrogen bond with OH of Tyr B10

Effect of TyrB10→PheB10 mutation on k_{off} is reproduced. Affinity is large (consistent with the detoxification role).



isolated Mb TrHb TyrB10 \rightarrow PheB10 -21.4 -25.0 -37.2 -34.3 kcal/mol



Further confirmation: *in silico* mutated proteins

TyrB10-Phe mutant: does not detoxify efficiently . Why?

Ouellet et al, PNAS (2002), 99, 5702.

A. Bidon-Chanal et al, J. Am. Chem. Soc. (2007), 129, 6782.

MD of oxygenated proteins



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TyrB10-Phe (black) GlnE11-Ala (grey) mutants Always closed!!!

Wt



 Our data strongly support the hypothesis that the access of NO to the heme cavity is dynamically regulated by the TyrB10-GInE11 pair, which acts as a molecular switch that controls opening of the ligand diffusion tunnel.

Binding of O₂ to the heme group triggers local conformational changes in the TyrB10-GInE11 pair, which favor opening of the PheE15 gate residue through global changes in the essential motions of the protein skeleton.

Role of the pre A helix in ligand entry modulation

M. Smegmatis TrHbN is almost identical to that of M.T. but lacks a fragment (pre A helix)



Strikingly, M. Smegmatis trHbN does not detoxify efficiently NO!

Simulation shows that deletion of pre A helix results in the trapping in the closed conformation, consistently with experimental results



Lama et al, JBC, 284, 14457, 2009.

Product release



Nitrate should be released to regenerate the enzyme.
We have performed classical MD simulations
QM model system and QM-MM calculations

Marti et al, JACS, 2008, 130, 1688.



QM-MM calculations: heme+ligand+water+TyrB10+ GInE11; just heme+ligand QM subsystems

High spin (sextet) is ground state at LACV3P*/B3LYP level for the large system



This electrostatic screening loosens the Fe-nitrate bond (barrier less than 4 kcal/mol), compared to 18 kcal/mol in the reduced model

Finally, once the bond is broken, classical MD simulations indicate that nitrate exits the protein very easily, but not along the (apolar) tunnels used for (NO, O_2) ligand entry.



Free energy profile for ligand exit



Minimum: interaction with ThrE2, key in assisting exit

Tryptophane and Indoleamine dioxygenases (TDO, IDO)



TDO in liver, IDO widespread distribution IDO target of anticancer drugs

Differences between IDO and TDO

Localization and structure

- IDO is ubicuous and monomeric (PM ≈ 45 kDa)
- TDO is tetrameric and located mainly in liver (PM≈ 167 kDa) Sequence identiy <10%

Catalysis:

- IDO reacts with L-Trp, D-Trp and other indoles
- TDO is more specific for *L-Trp*

3. Function:

- IDO is induced by *IFN-\gamma* and is related with immune response regulation
- TDO is implied in Trp metabolism in liver



IDO is expressed in placenta and in cancer cells to reduce T cells proliferation
It has been observed that its inhibition leads to a reduction in tumoral growth

Development of selective inhibitors for IDO (which do not inhibit TDO) is of great interest

Samelson-Jones, B. *et al. Biochemistry*. (**2006**) Munn, D.H. et al.. Science 281, 1191–1193 (1998).

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Human IDO



Monomer with two domains

Heme group in each monomer

High structural similarity

X. Campestris TDO



- Homotetramer
- One heme group per monomer
- Binds 8 Trp molecules per tetramer

Relevant questions associated to these proteins

Reaction mechanisms. Are there any differences?

How is substrate stabilized in the active site, and how this explains the stereoselectivity?

Which are the key residues in substrate binding and catalysis?

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How can we answer these questions?

 Flexible Docking (Autodock) to obtain enzyme-substrate initial structures
 Classical MD simulations (Amber) to analyze
 Interactions due to ligand binding

Free energy binding calculations using continuum solvent methods (*MMGBSA*).
 QM-MM (*Hybrid* – *SIESTA*) calculations for the reaction mechanism and oxygen affinity

Oxy – L-Trp adduct in TDO



Arg 117, Tyr 113, His55 y Phe 51 stabilize Trp in active site

L-Trp vs D-Trp in TDO



xcTDO_{wt}-O₂/D,L-Trp

	xC-TDO	
Donor-Acceptor Pairs	L-Trp	D-Trp
Trp O – R 117 Νε	100	100
Trp OXT – R117NH1	100	100
Trp O – Y113 OH	100	100
H55 Νε2 - Trp Νε1	100	95
Trp OXT - T254 NH	100	85
T254 OH - Trp NH ₃ +	98	14
Binding energy (kcal/mol)	-51.7	-46.1
Km (mM)	0.114	16

Oxy – L-Trp adducts in IDO: *Two conformations for L-Trp in active site*



D-Trp in IDO



Just one conformation (conf 2) was stable

>D-Trp, the structure is similar to that found for L-Trp Cf_2 , but lacks the interaction with R231.

Previously proposed reaction mechanisn



- Based on isolated Trp O₂ reactivity
- Replacing the NH moiety for S or O, inhibitors for IDO and TDO are found
- 1(N)-Methyl-Trp is a TDO inhibitor but reacts with IDO.
- H55A and H55S TDO present a lower activity towards Trp



Reaction starts with a direct attack to C2 to yield an epoxide and a ferryl intermediate







- TDO and IDO active sites present significant differences regarding size and flexibility
- IDO presents two conformations for L-Trp binding, while in TDO just one conformation is found
- The first step consists on a direct attack to C2 Trp to yield an epoxide and a ferryl intermediate
- Reaction in IDO probably continues with an attack of the ferryl to C2, epoxide opening, assisted by Trp-NH₃⁺.

Take Home Message:

Complex problems regarding chemical reactivity and catalysis in proteins requiere of a smart and careful combination of purely quantum, classical, and QM-MM strategies

Both careful consideration of the chosen model (level of theory) and sampling are necessary to obtain meaningful results

Choosing adequately benchamarks and comparison cases is essential to validate results

A deep interplay of computer simulation and experiment is an excellent way of getting the best possible insight into a given problem

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