

# Infrared microspectroscopy for environmental science: from the basis to data analysis and results' interpretation

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in Environmental Scientific Projects



# Talk outline

## □ Introduction

- The infrared spectral region
- Basics on vibrational spectroscopy

## □ Instrumentation

- FTIR interferometer
- Vis-IR microscope
- InfraRed Synchrotron Radiation (IRSR)
  - Infrared Synchrotron Radiation sources
  - IRSR Advantages
  - IR Beamline design

## □ FTIR spectroscopy for environmental sciences

## □ Biological information from a FTIR microspetrum

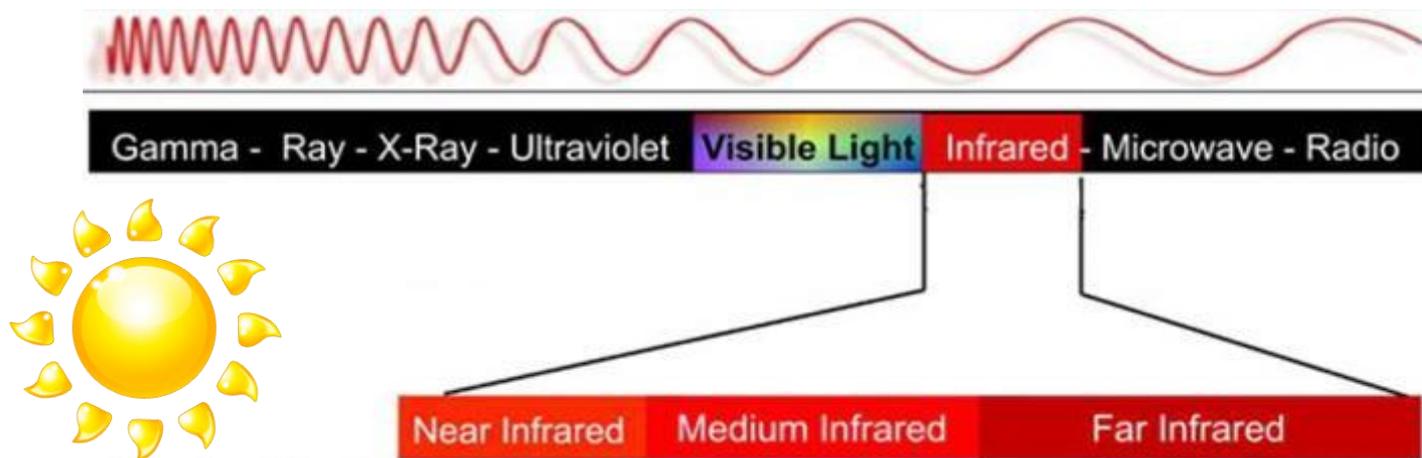
## □ From the sample to the biological information

- Sampling techniques
- Sample preparation
- Data acquisition
- Data analysis

## □ Some examples

- Real-time Chemical imaging of bacterial activity in biofilms
- In Situ FT-IR Microscopic Study on Enzymatic Treatment of poplar Wood Cross-Sections
- Transmissible spongiform encephalopathies

# The infrared spectral region



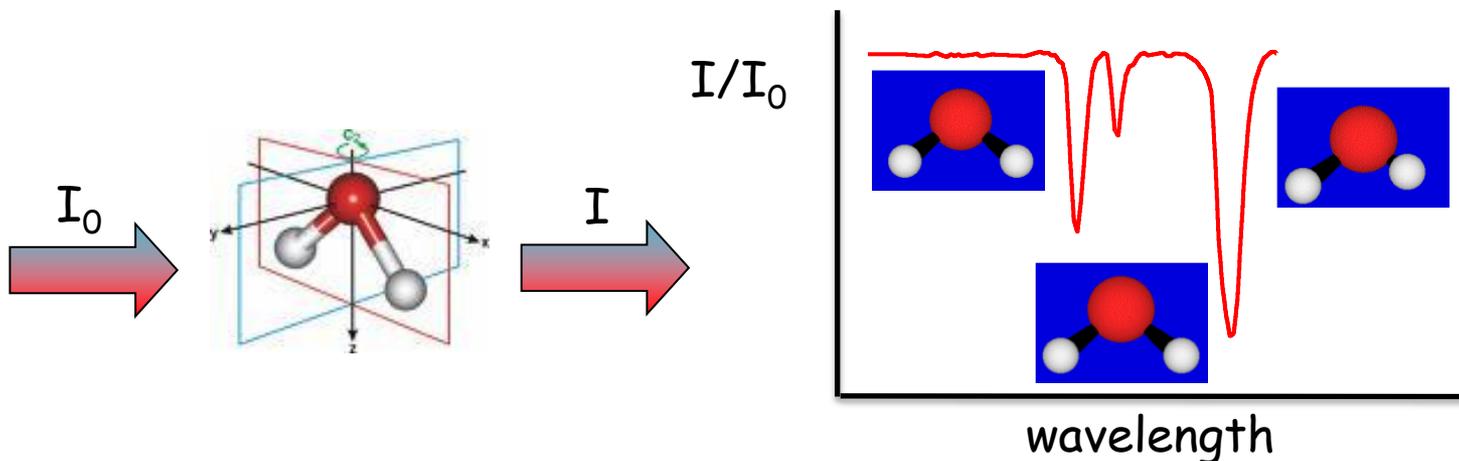
	<b>NIR</b>	<b>MIR</b>		<b>FIR</b>
$\lambda$ ( $\mu\text{m}$ )	0.74	3	30	300
$\nu$ (THz)	400	100	10	1
$\bar{\nu}$ ( $\text{cm}^{-1}$ )	$\sim 13000$	$\sim 3333$	$\sim 333$	$\sim 33$
E (eV)	1.65	0.413	0.041	0.004
E (Kcal/mol)	37	10	1	0.1

The IR spectral range is a wide region of long wavelengths

# Basics on vibrational spectroscopy

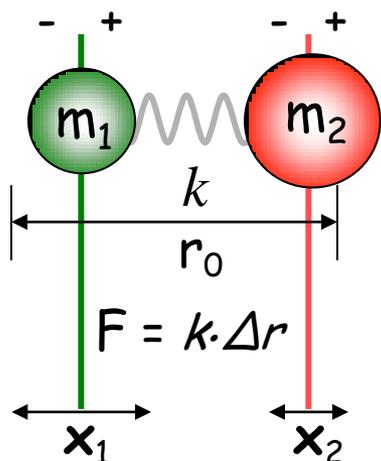
IR spectroscopy is an absorption spectroscopy

Direct resonance between vibrational transition frequency and photon frequency



## Vibrational (stretching) frequencies

For a diatomic molecule or molecular fragment



$$\nu = \frac{1}{2\pi} \sqrt{\frac{k}{\mu}}$$

$$\mu = \text{Reduced mass} = \frac{m_1 m_2}{m_1 + m_2}$$

$k$  = Spring constant

$\Delta r$  = Displacement

# Vibrational energies

## Harmonic approximation

$$E_{vib} = h\nu_0(n + 1/2), \quad \nu_0 = \text{fundamental vibrational frequency} = \frac{1}{2\pi} \sqrt{\frac{k}{\mu}}$$

## Anharmonicity

$$E_{vib} = h\nu_0[(n + 1/2) - x_0(n + 1/2)^2 + \text{higher terms}]$$

$x_0$  = anharmonic constant

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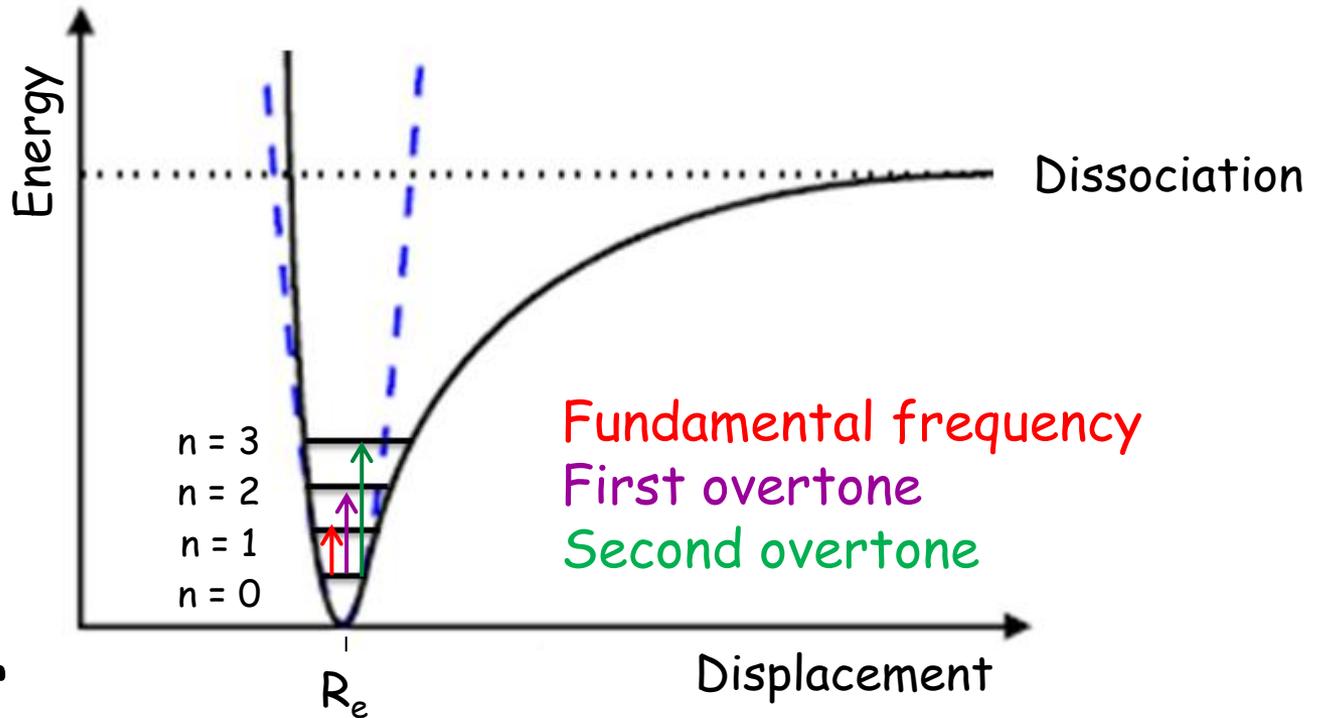
Harmonic oscillator

$$\Delta n = 1$$

—————

Anharmonic oscillator

$$\Delta n = \text{integer}$$

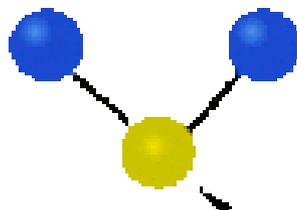


# Fundamental modes of vibrations for polyatomic molecules

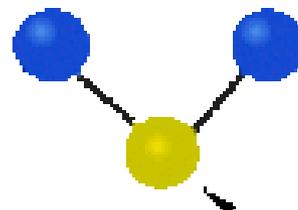
Linear N-atom molecule:  $3N-5$  modes of vibration

Non linear N-atom molecule:  $3N-6$  modes of vibration

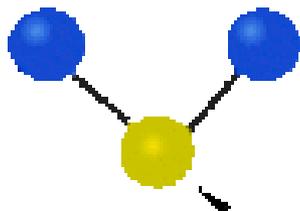
**Symmetric  
Stretching**



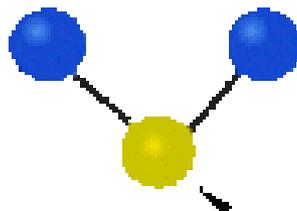
**Antisymmetric  
Stretching**



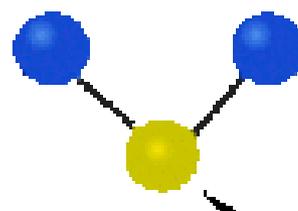
**Scissoring**



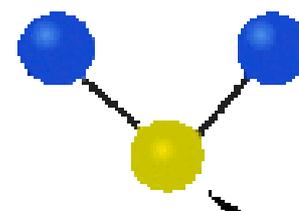
**Rocking**



**Wagging**



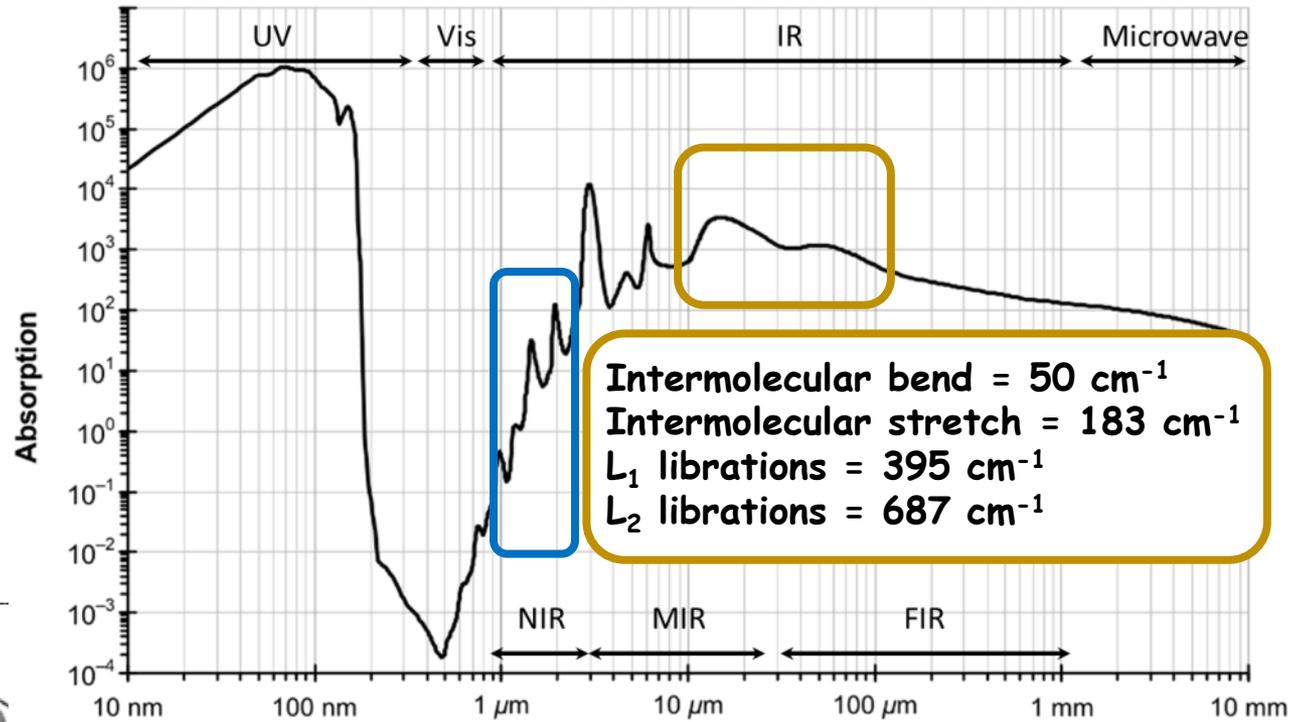
**Twisting**



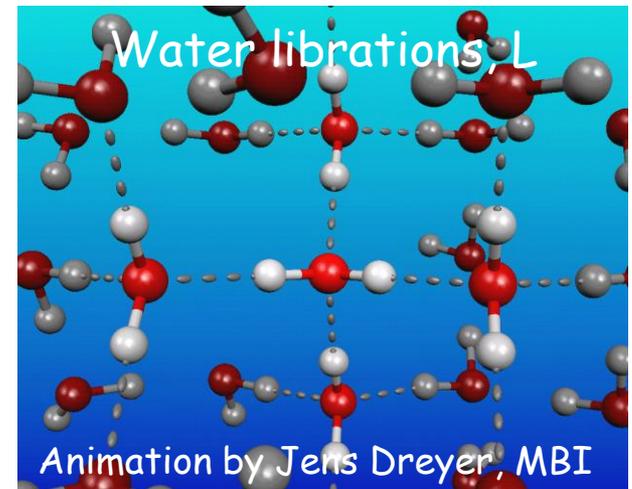
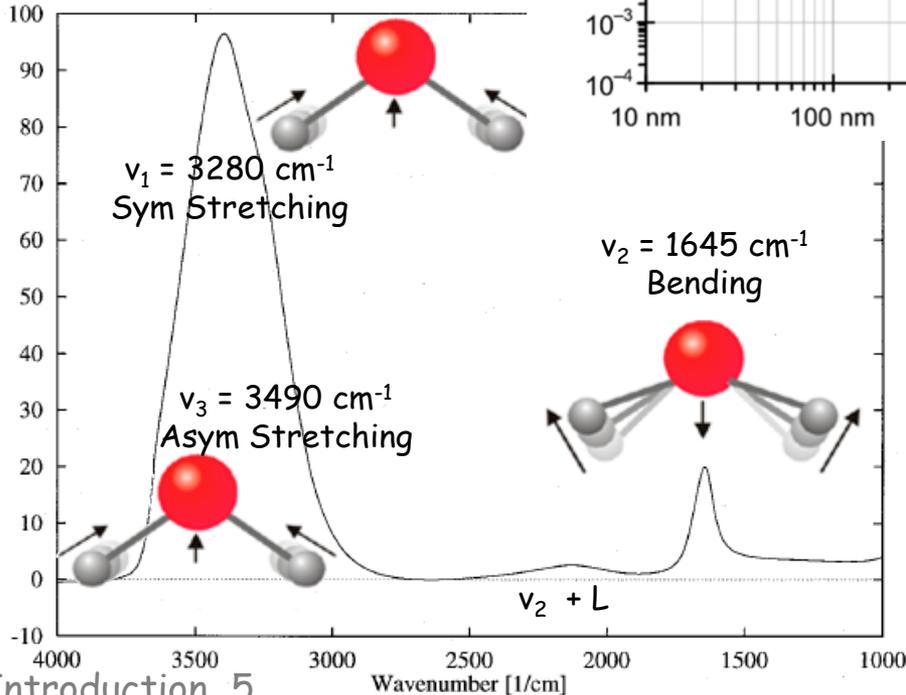
**Only the vibrational modes that produce a change of the molecular dipole moment are IR active**

# The liquid water absorption spectrum

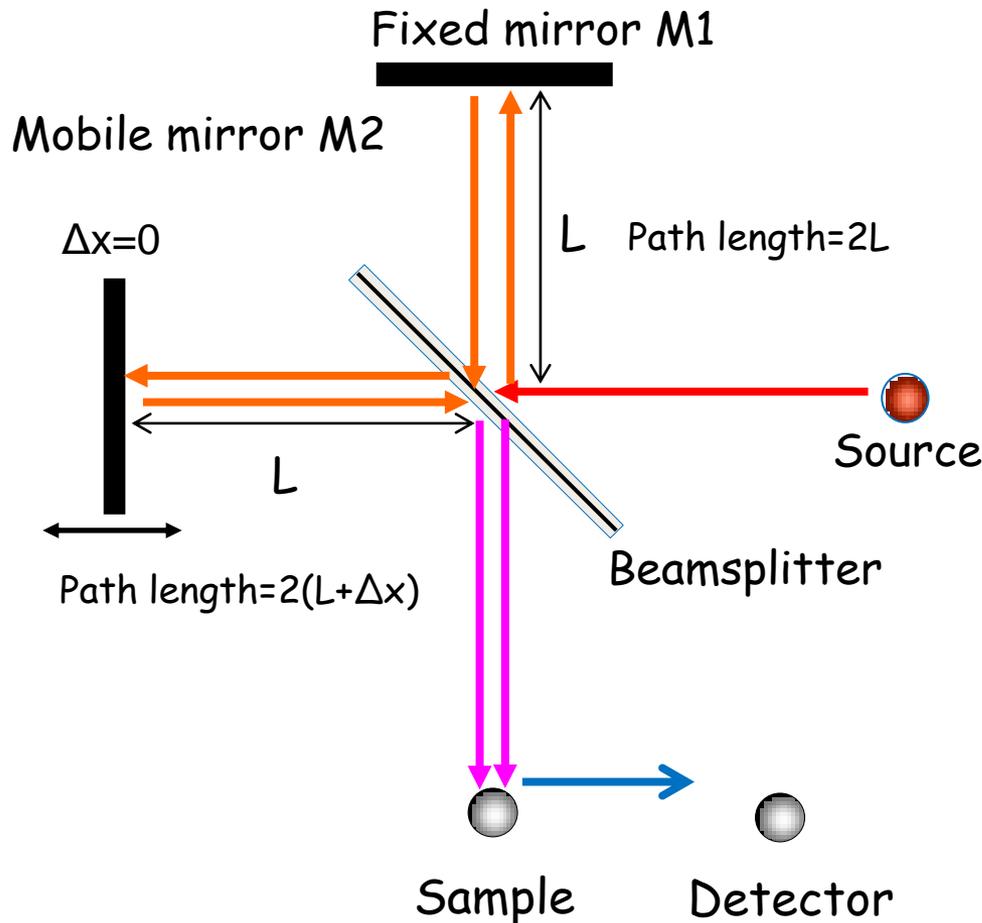
Overtone  
and  
combination bands



Intermolecular bend = 50 cm<sup>-1</sup>  
Intermolecular stretch = 183 cm<sup>-1</sup>  
L<sub>1</sub> librations = 395 cm<sup>-1</sup>  
L<sub>2</sub> librations = 687 cm<sup>-1</sup>



# FTIR interferometer



## Conventional sources

NIR: Tungsten lamp  
MIR: Glow bar (SiC)  
FIR: Hg-Arc

## Beamsplitters

NIR:  $\text{CaF}_2$   
MIR: KBr  
FIR: Mylar, Silicon

## Detectors

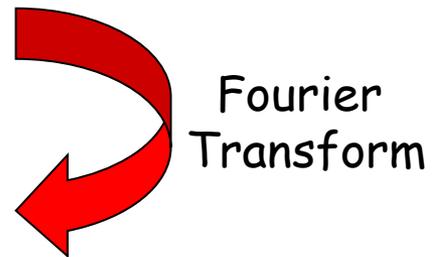
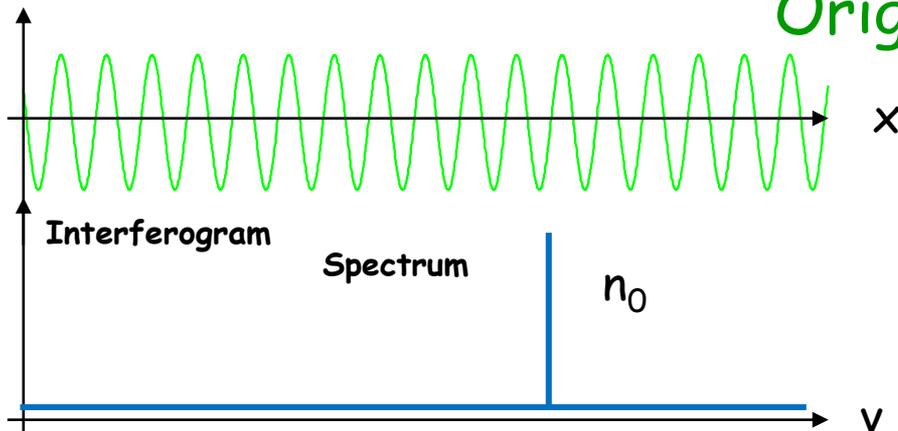
NIR - InGaAs, InSb, Ge, Si room temperature detectors  
MIR: Room temperature DLaTGS  
Nitrogen cooled MCT  
FIR - He Cooled Silicon Bolometer  
Room temperature DLaTGS

## Spectral Resolution

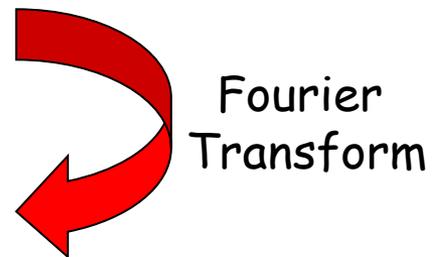
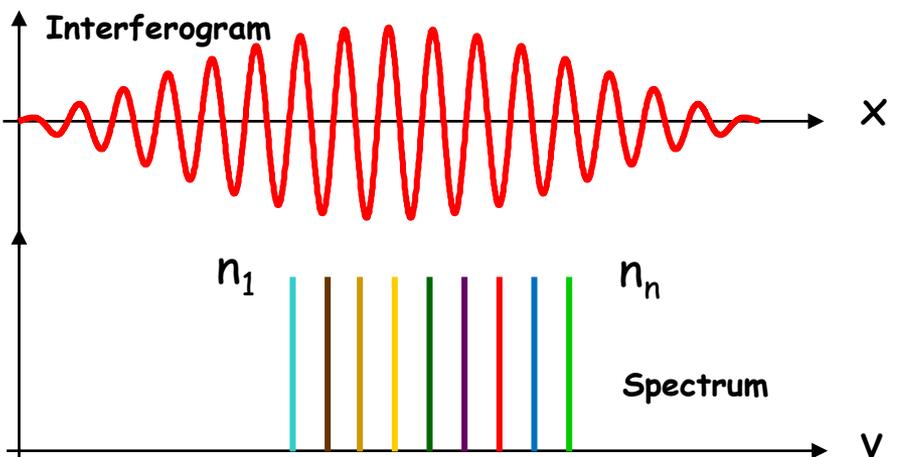
$$\Delta\nu \sim 1/\Delta x \text{ (cm}^{-1}\text{)} \sim 0.001 \text{ cm}^{-1} \sim 1 \text{ meV}$$

# Origin of the interferogram

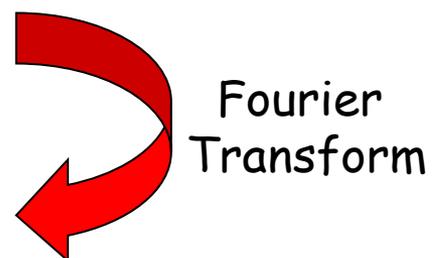
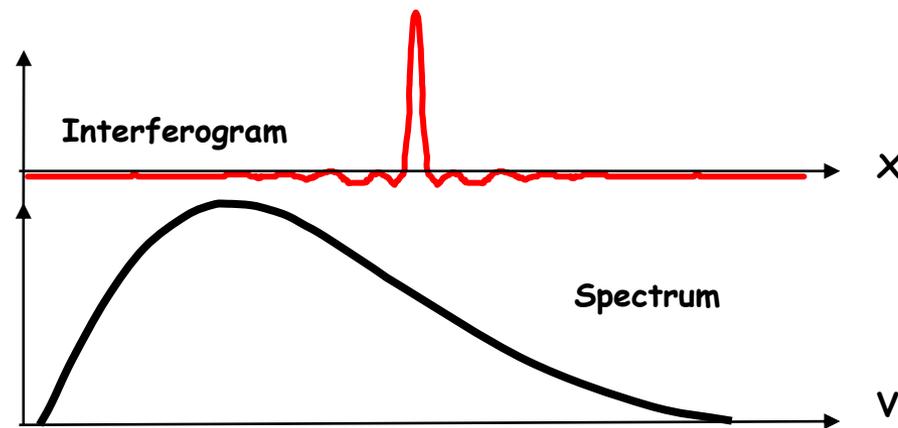
Monochromatic source



Policromatic source



Continuum source



# Advantages of FTIR interferometers

- **Jacquinot's throughput advantage**
  - All source wavelengths are measured simultaneously in an interferometer, whereas in a dispersive spectrometer they are measured successively. A complete spectrum can be collected very rapidly and many scans can be averaged in the time taken for a single scan of a dispersive spectrometer.
- **Fellgett's Advantage (Multiplex advantage)**
  - For the same resolution, the energy throughput in an interferometer can be higher than in a dispersive spectrometer, where it is restricted by the slits.
- **Connes' accuracy advantage**
  - The wavenumber scale of an interferometer is derived from a HeNe laser that acts as an internal reference for each scan. The wavenumber of this laser is known very accurately and is very stable. As a result, the wavenumber calibration of interferometers is much more accurate and has much better long term stability than the calibration of dispersive instruments.

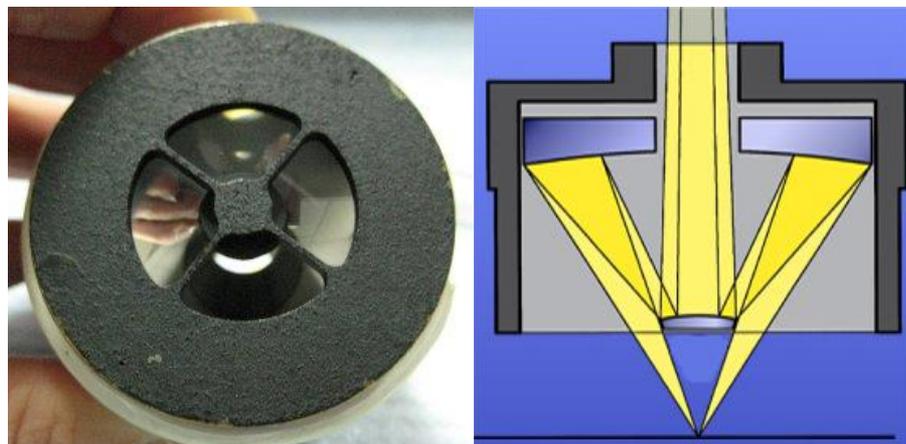
As a result, a FTIR spectrum can be measured with the same signal-to-noise ratio than a dispersive spectrum in a much shorter time (seconds rather than minutes) and spectral subtractions can be carried out without frequency errors

# Vis-IR microscopes

Spatially resolved chemical information on heterogeneous samples are obtained by coupling FTIR spectrometers with specially designed Vis-IR microscopes



Schwarzschild objective



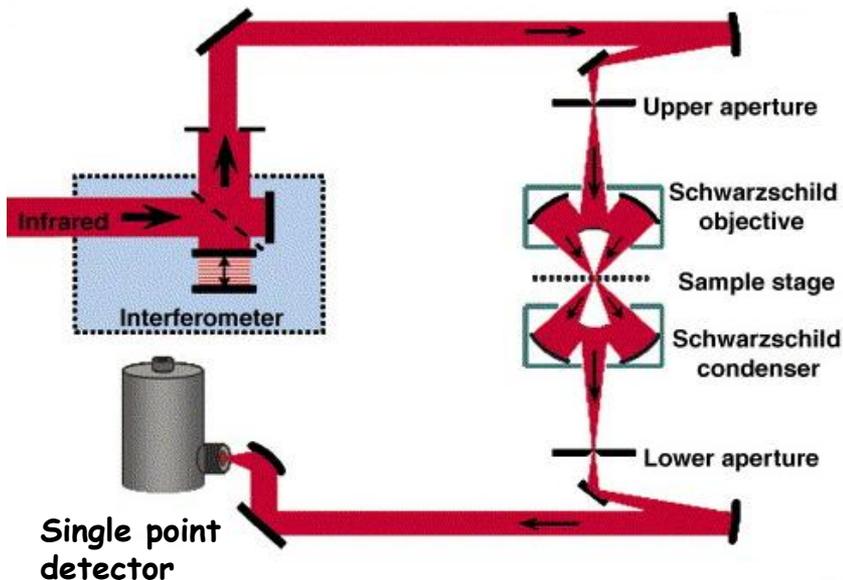
*The highest achievable lateral resolution,  $\delta$ , is diffraction limited*

Objective NA	Wavelength	$\delta$
0.4	10 $\mu\text{m}$ ( $1000\text{cm}^{-1}$ )	$\sim 15 \mu\text{m}$
	2.5 $\mu\text{m}$ ( $4000\text{cm}^{-1}$ )	$\sim 4 \mu\text{m}$
0.65	10 $\mu\text{m}$ ( $1000\text{cm}^{-1}$ )	$\sim 9,5 \mu\text{m}$
	2.5 $\mu\text{m}$ ( $4000\text{cm}^{-1}$ )	$\sim 2,5 \mu\text{m}$

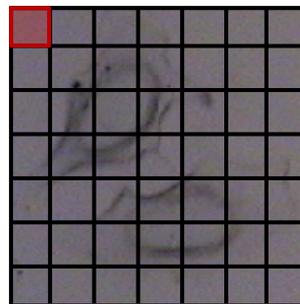
$$\delta \approx 0.61 \lambda / NA$$

# FTIR mapping versus FTIR imaging

Mapping layout

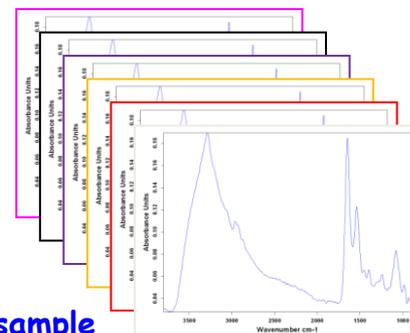
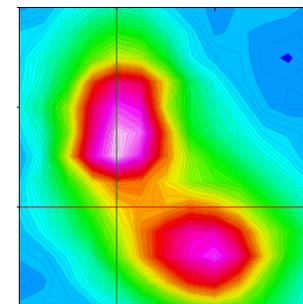


Optical sample image

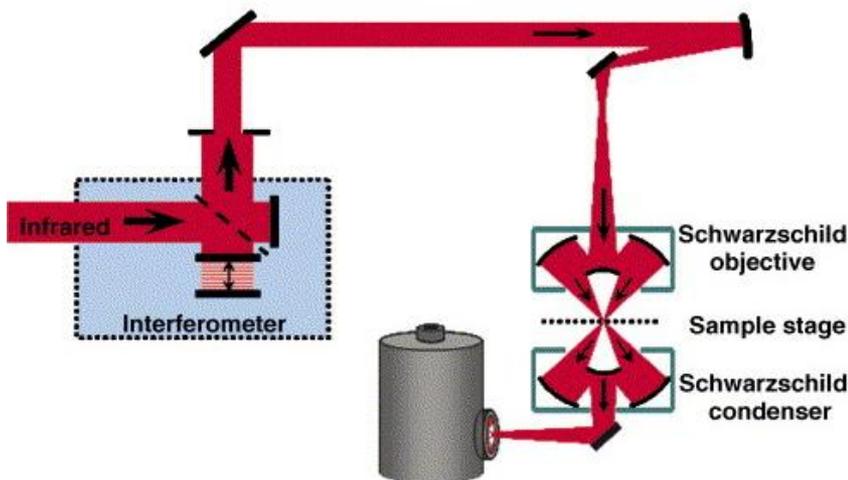


- The map pixel size is provided by the aperture settings
- Sequential acquisition: Slow imaging of large sample areas
- Low detector noise, high SNR

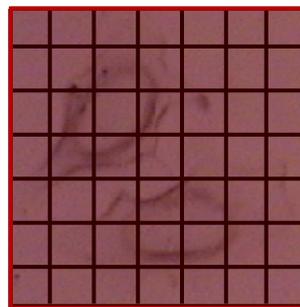
Chemical sample image



Imaging layout



Optical sample image

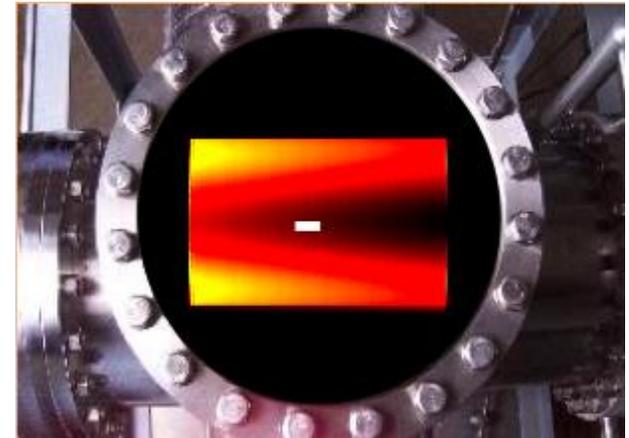
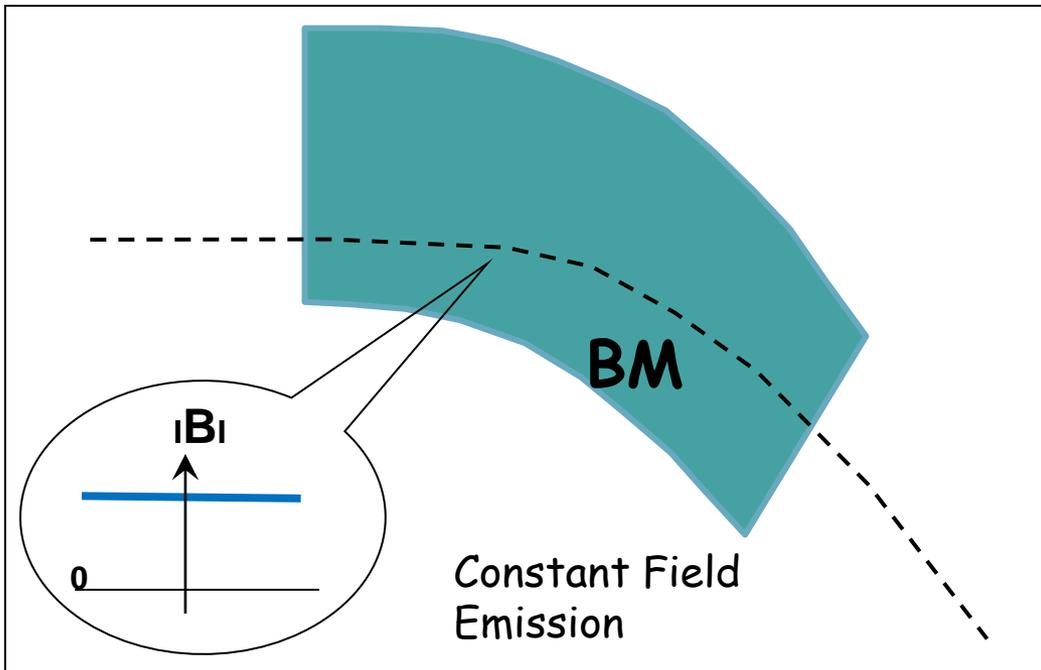


- The image pixel size is provided by the individual pixel of the detector
- Parallel acquisition: Fast imaging of large sample areas
- High detector noise, low SNR

# InfraRed Synchrotron Radiation (IRSR) \_ Sources

## Standard Bending radiation

Emitted during the circular trajectory in the bending magnet (BM) due to the constant magnetic field,  $B$



Natural opening angle

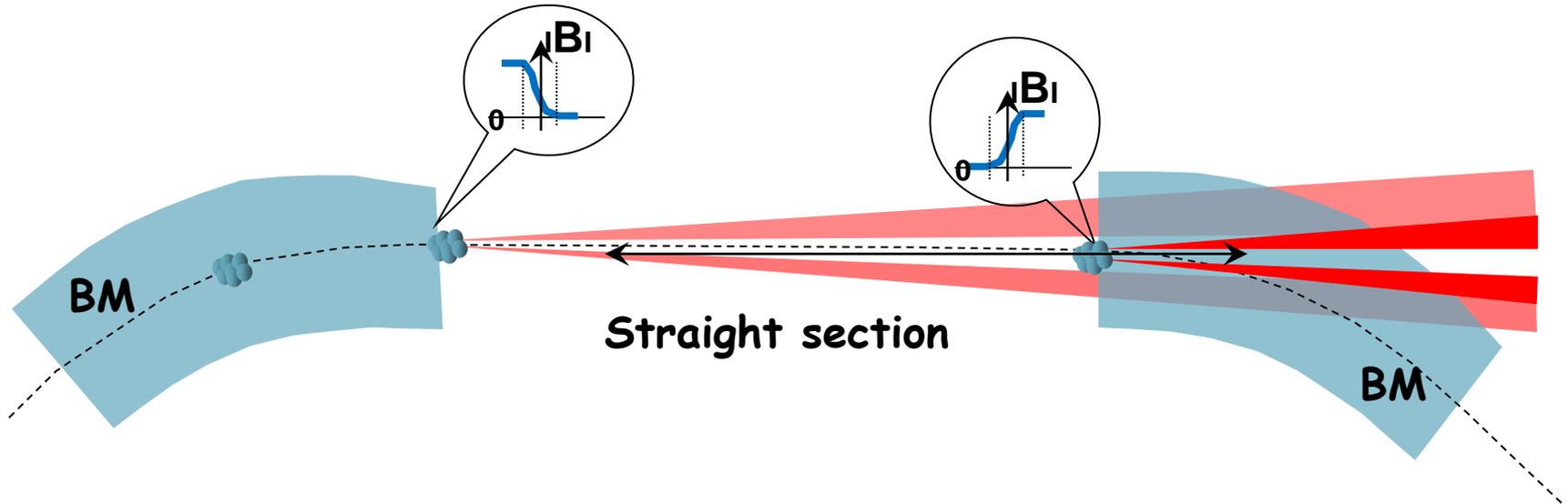
$$\Theta_{V-NAT}(\text{rad}) = 1.66 (\lambda / \rho)^{1/3}$$

$$P(\lambda) = 4.4 \cdot 10^{14} \times I \times \Theta_H \times bw \times (\rho / \lambda)^{1/3} \text{ photons s}^{-1}$$

$I$  (A) is the ring current,  $\Theta_H$  (rads) the horizontal collection angle,  $bw$  (%) the bandwidth,  $\lambda$  ( $\mu\text{m}$ ) the wavelength, and  $\rho$  (m) the radius of the bending

# Edge Emission

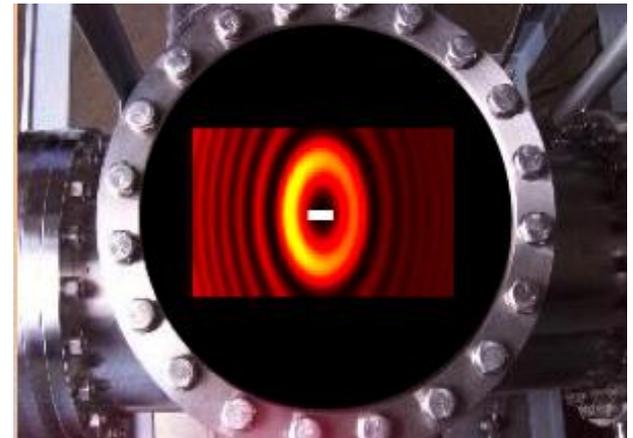
Emitted at the entrance (exit) of a bending magnet due to the rapid variation of the B field



In the Far-Field approximation:

$$P = \alpha \times I \times \gamma^4 \Theta^2 / (1 + \gamma^2 \Theta^2)^2 \text{ photons s}^{-1}$$

$I$  is the current in amperes,  
 $\Theta$  (rads) the emission angle  
(concentrated in  $\Theta_{\text{max}} \sim 1/\gamma \sim 10$  mrad)

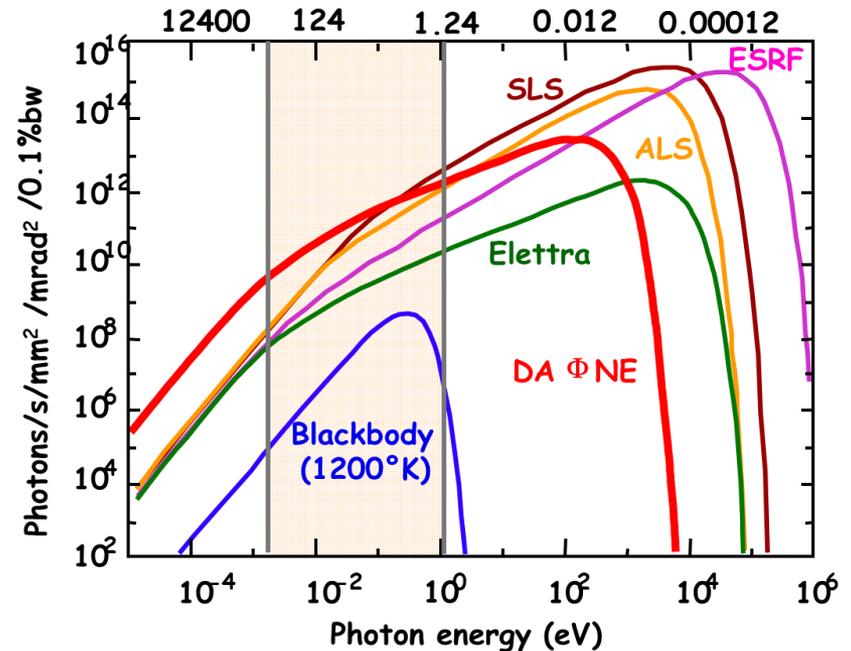
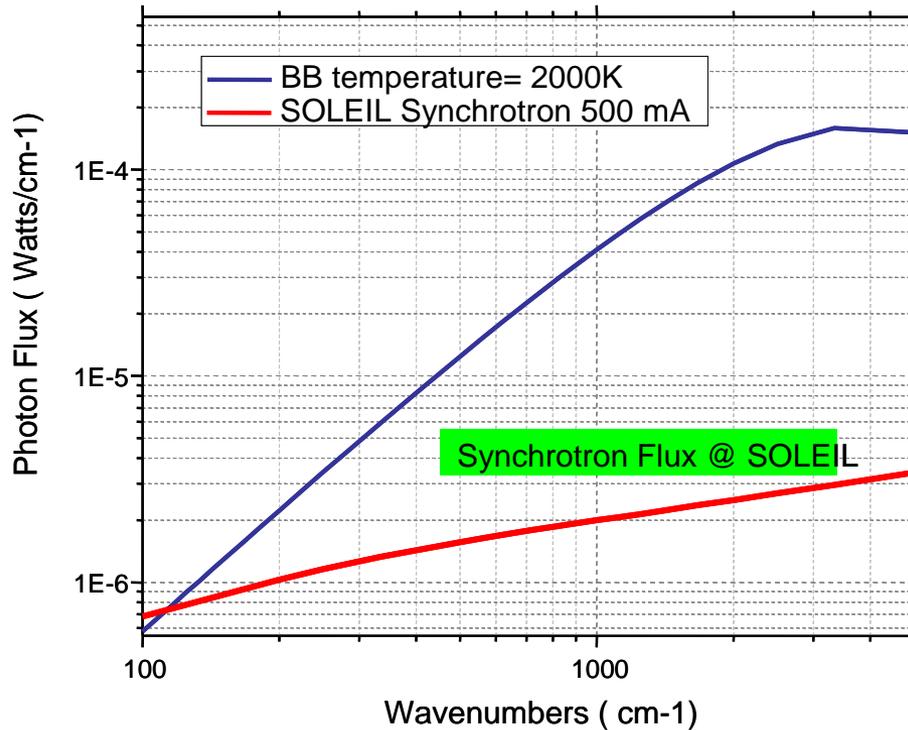
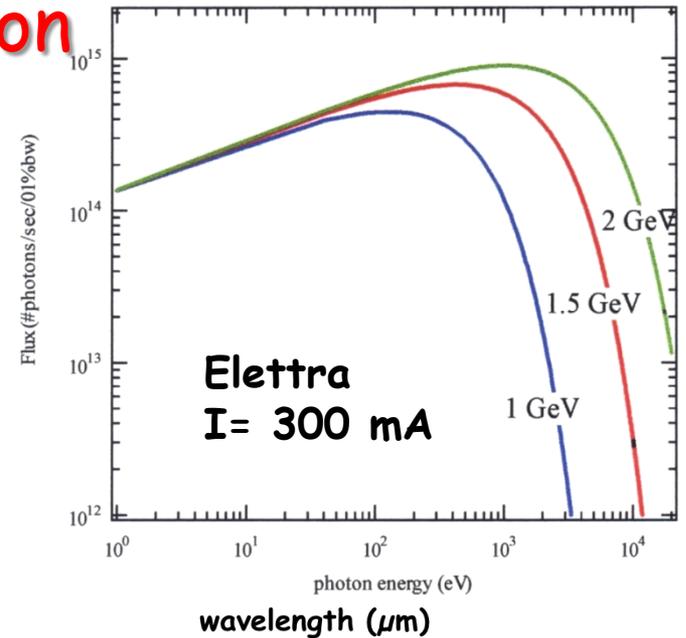


# InfRared Synchrotron Radiation (IRSR) \_ Characteristics

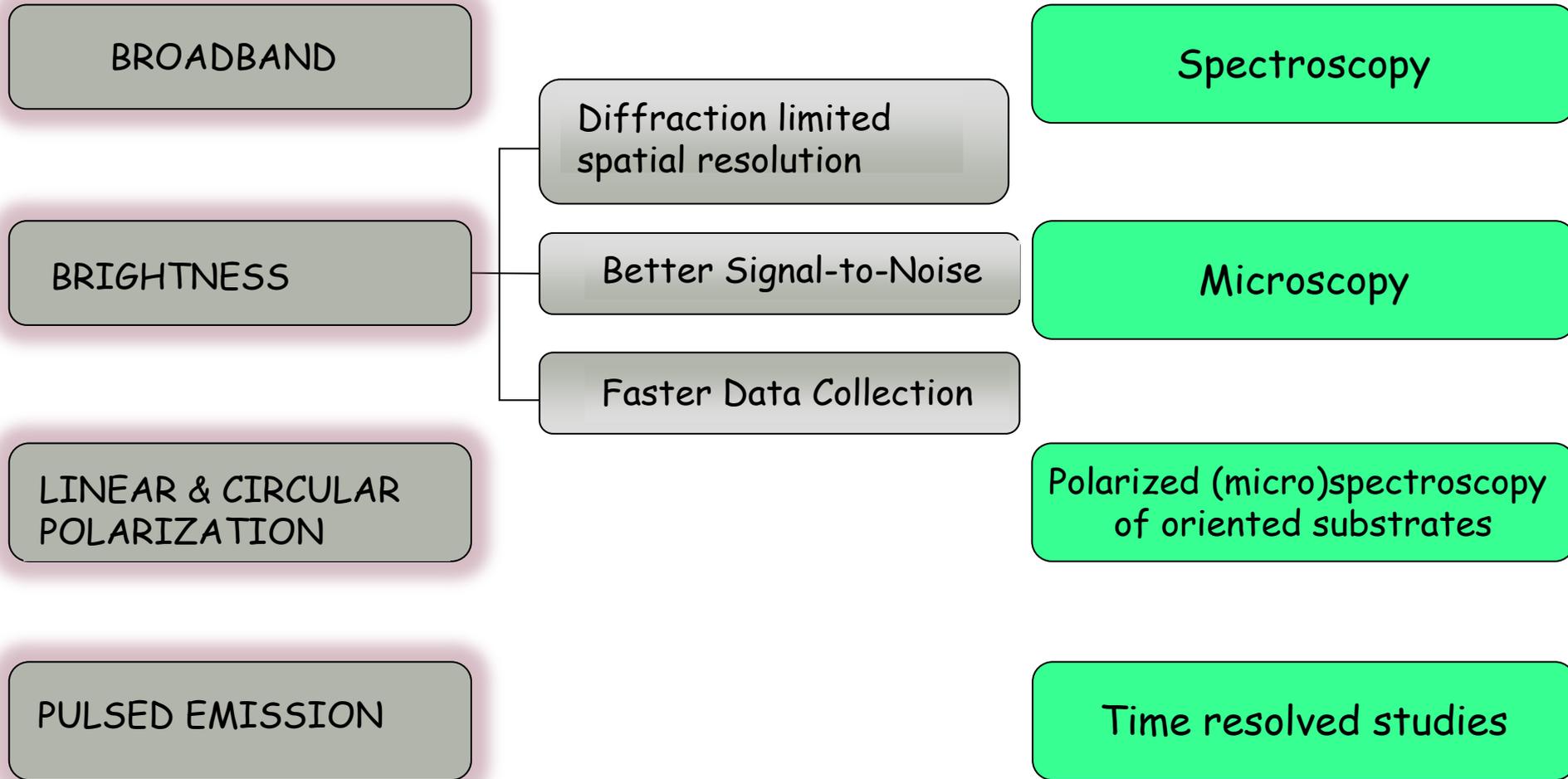
The IRSR Flux and Brilliance depend on:

- beam current
- source size/emittance
- extraction aperture
- transmission optics

Instead scarcely depend on the machine energy



# Infrared Synchrotron Radiation (IRSR) Advantages

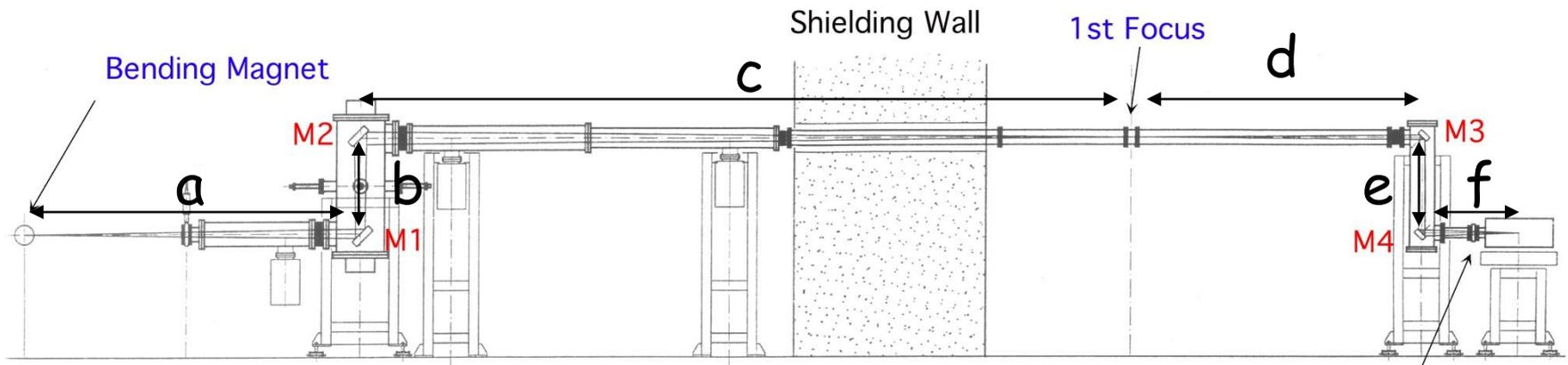


# IR beamline design

Conventional IR beamline layout: SISSI@Elettra

Radiation is collected over a solid angle of  
**65 mrad (H) × 25 mrad (V)**

M1 Plane mirror  
M2 Ellipsoidal mirror  
M3 Plane mirror  
M4 Ellipsoidal mirror



$$a = 3.5 \text{ m} \quad d = 1.5 \text{ m}$$

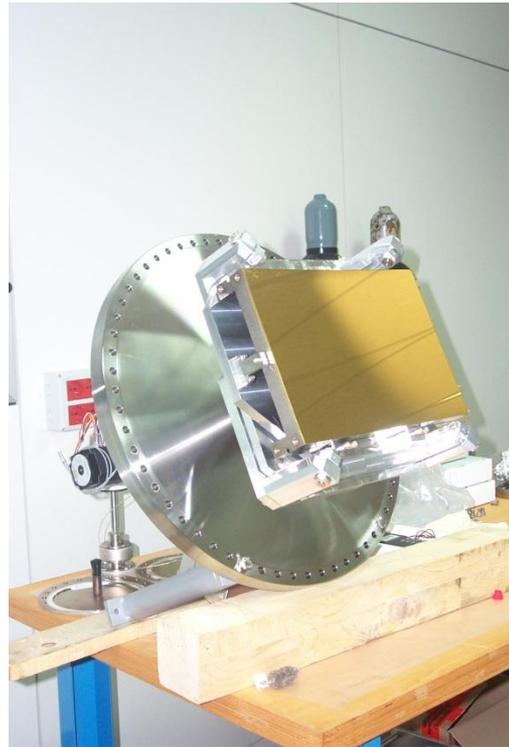
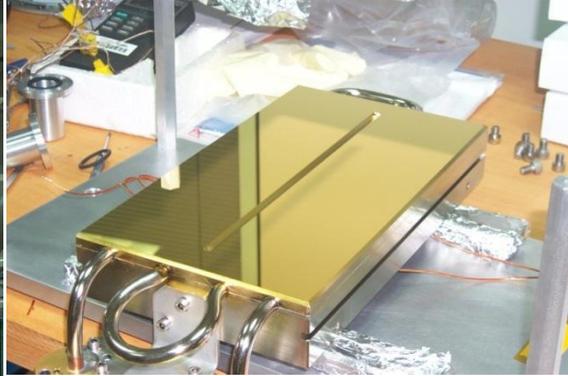
$$b = 1.0 \text{ m} \quad e = 1.0 \text{ m}$$

$$c = 11.5 \text{ m} \quad f = 2.5 \text{ m}$$

Final Focus and  
Diamond Window

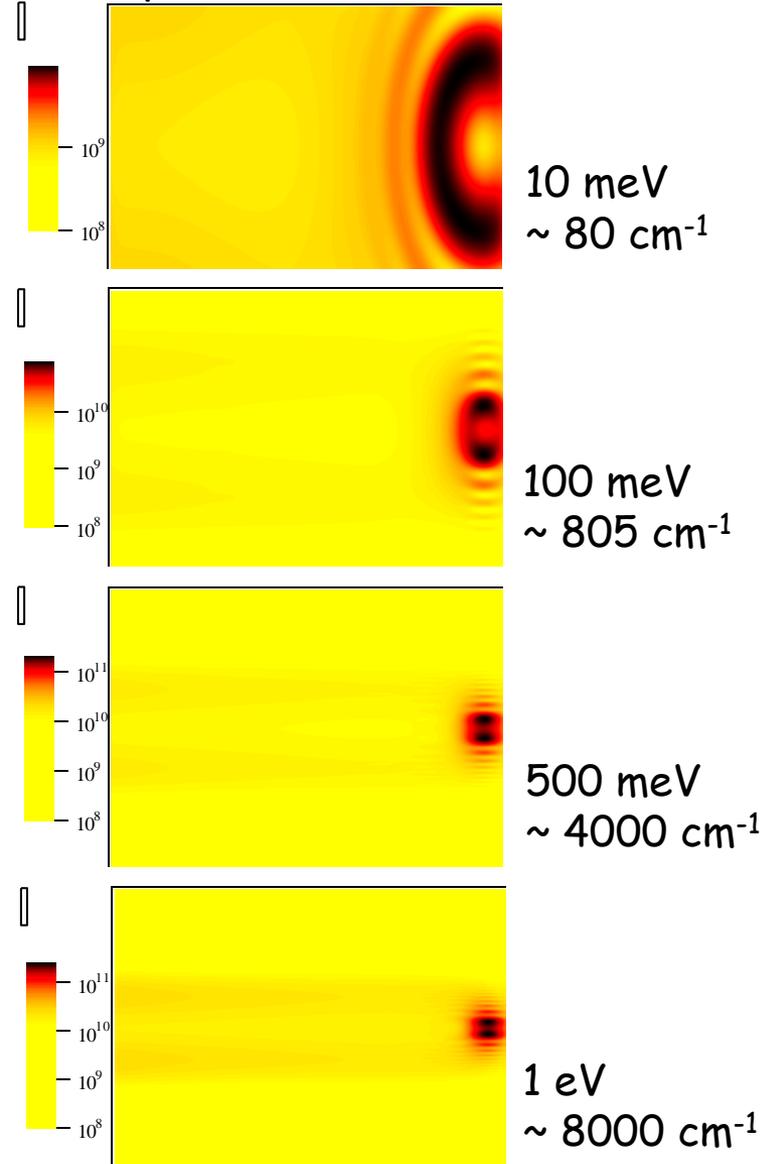
# SISSI first chamber

The water-cooled extraction mirror (M1)



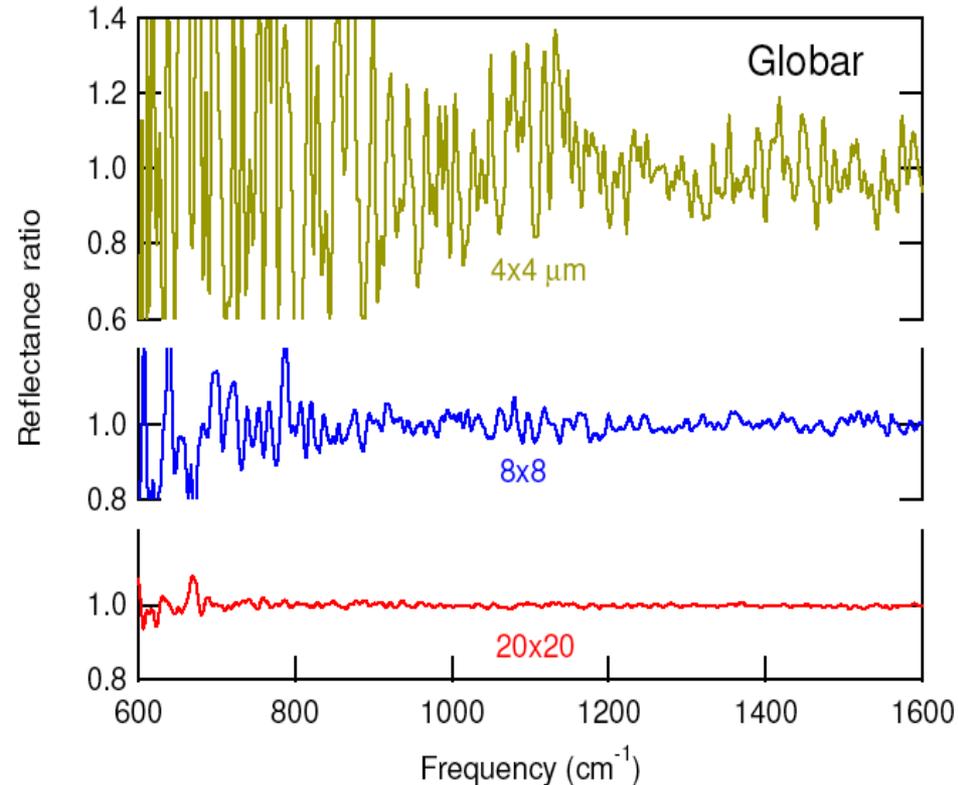
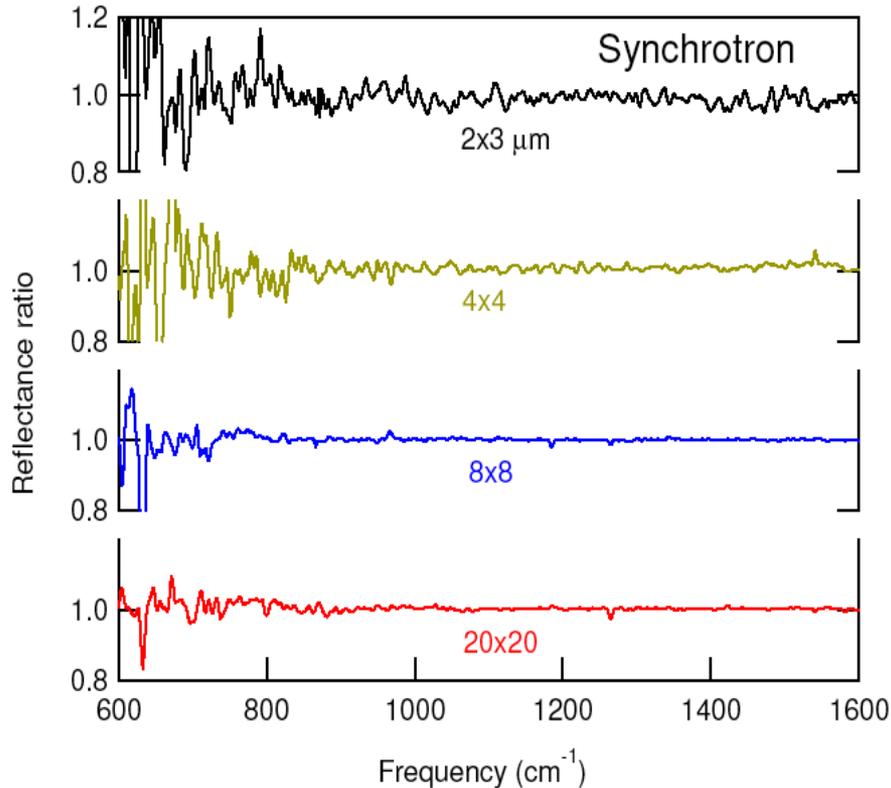
The focusing Ellipsoidal mirror (M2)

Intensity distribution on the extraction mirror (photons\*0.1%bw/mm<sup>2</sup>)



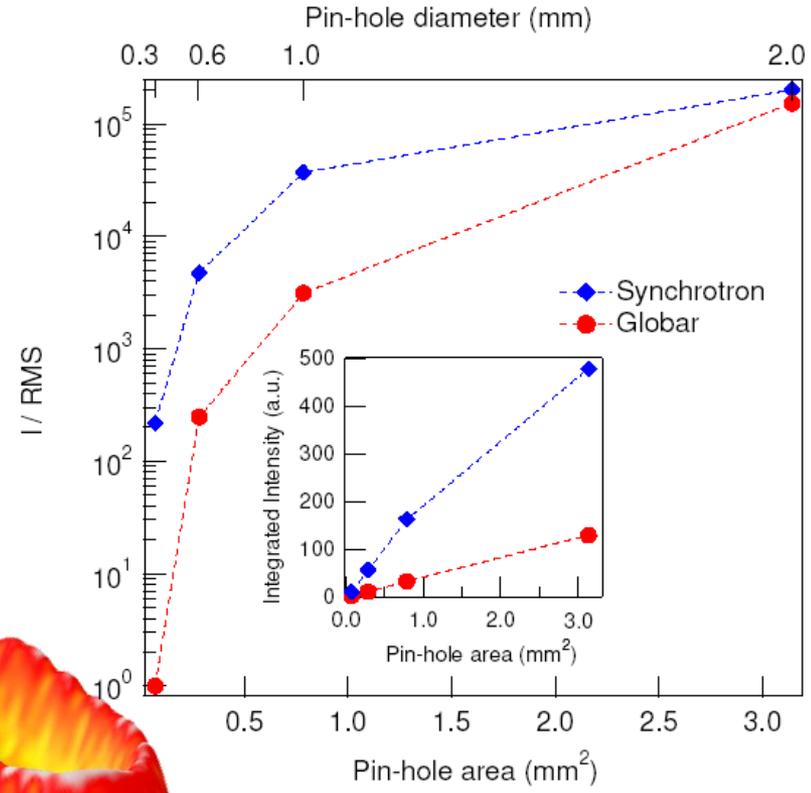
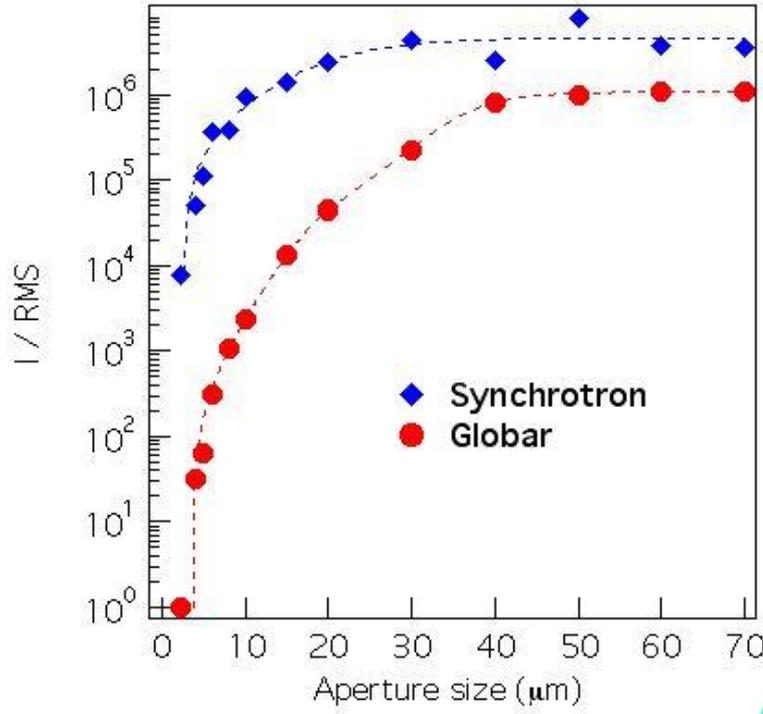
# MIR performances of SISSI@Elettra

Diffraction-limited lateral resolution is practically achievable only by exploiting the brightness advantage of SR



N<sub>2</sub> cooled MCT detector, 128 scans, 4 cm<sup>-1</sup> spectral resolution

# Figure of merit of SISSI@Elettra

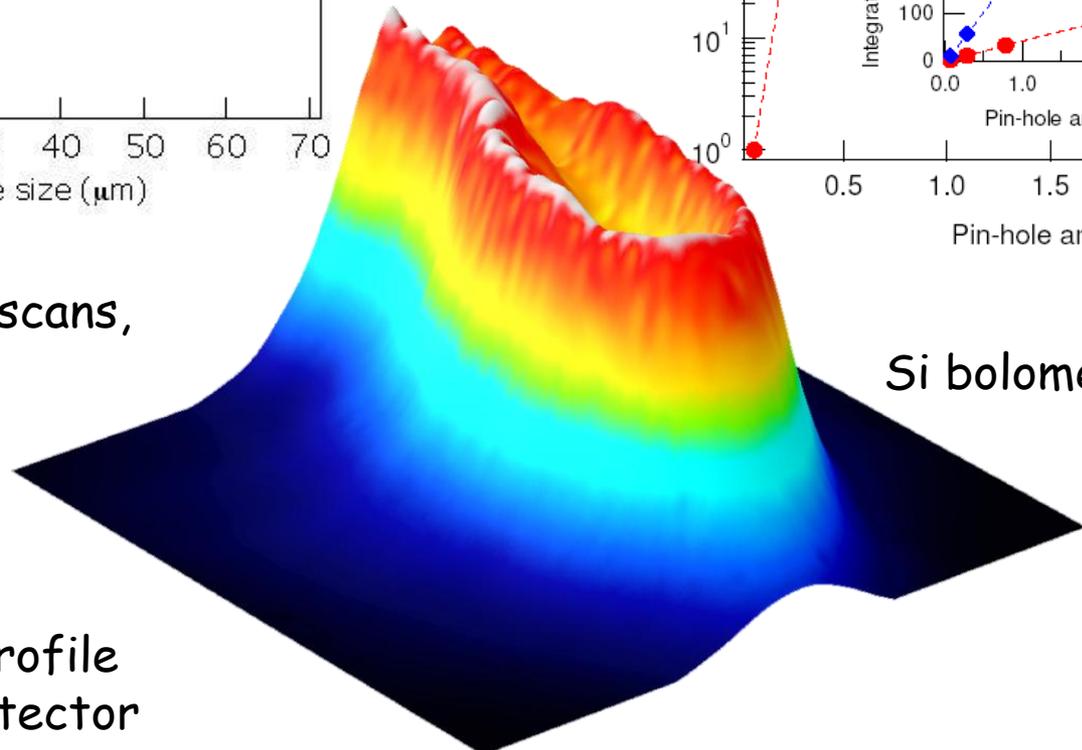


MIR

MCT detector, 128 scans,  
4 cm<sup>-1</sup>

FIR

Si bolometer, 128 scans,  
4 cm<sup>-1</sup>



SISSI MIR beam profile  
imaged with FPA detector

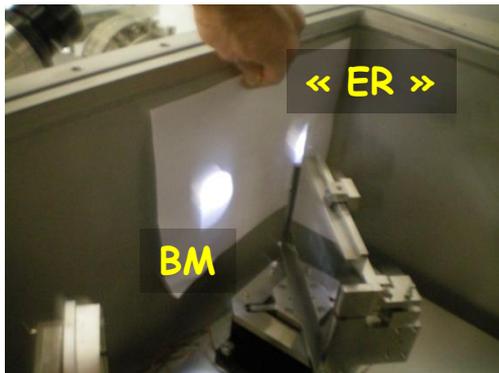
# Beamline branches simultaneously working

At SISSI beamline, the two branches can not operate simultaneously

More recent beamlines in newer 3<sup>rd</sup> generation SR facilities split BM and ER contributions for the simultaneous operation of two branches

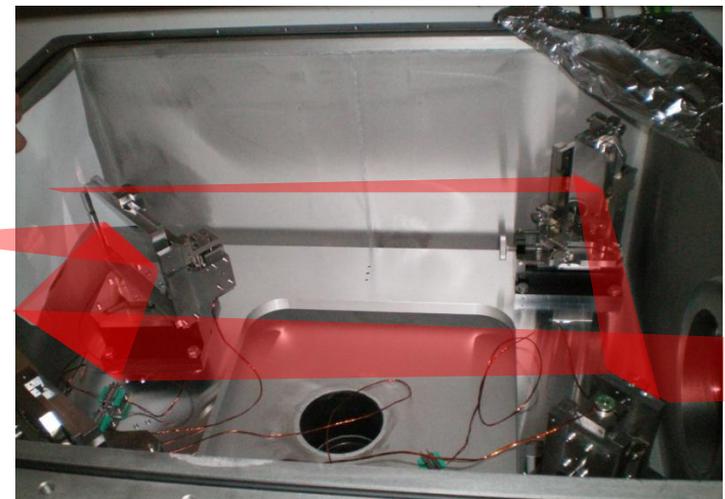


SMIS  
@  
Soleil



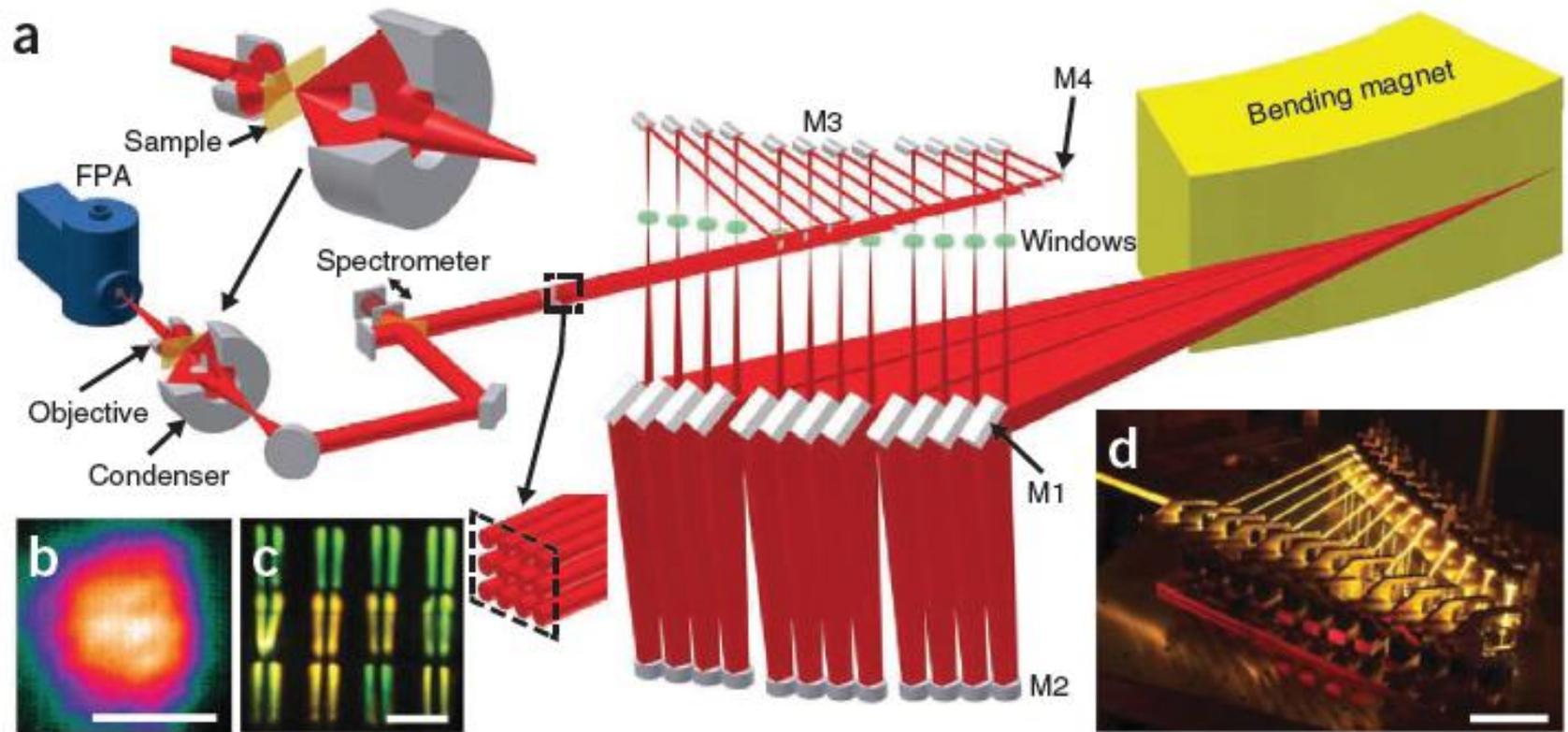
Microscope 2  
Branche ER

Microscope 1  
Branche BM



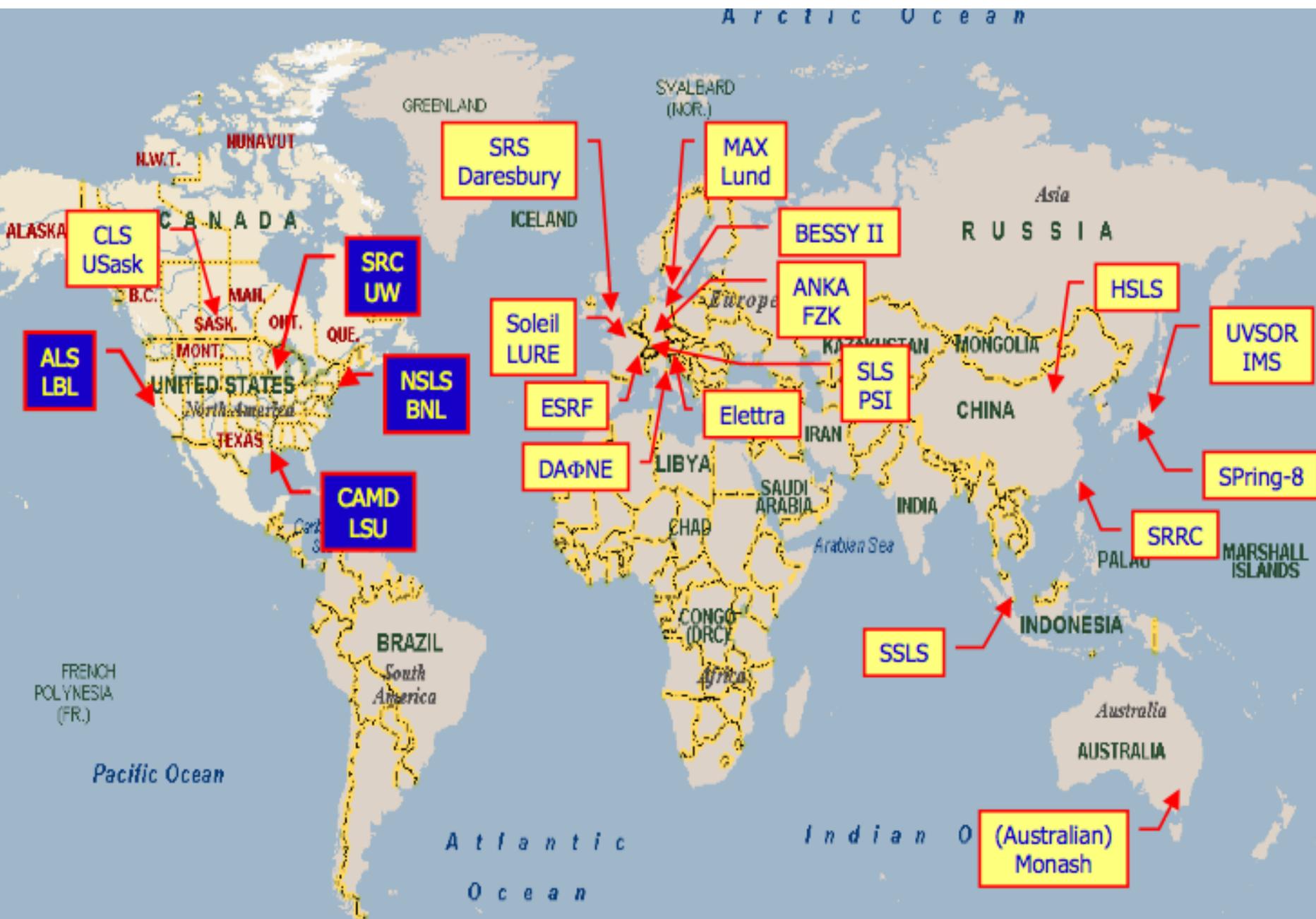
# More recent IR beamline layout: IRENI@Synchrotron Radiation Center, Wisconsin-Madison

The FTIR imaging approach: fast acquisition speed



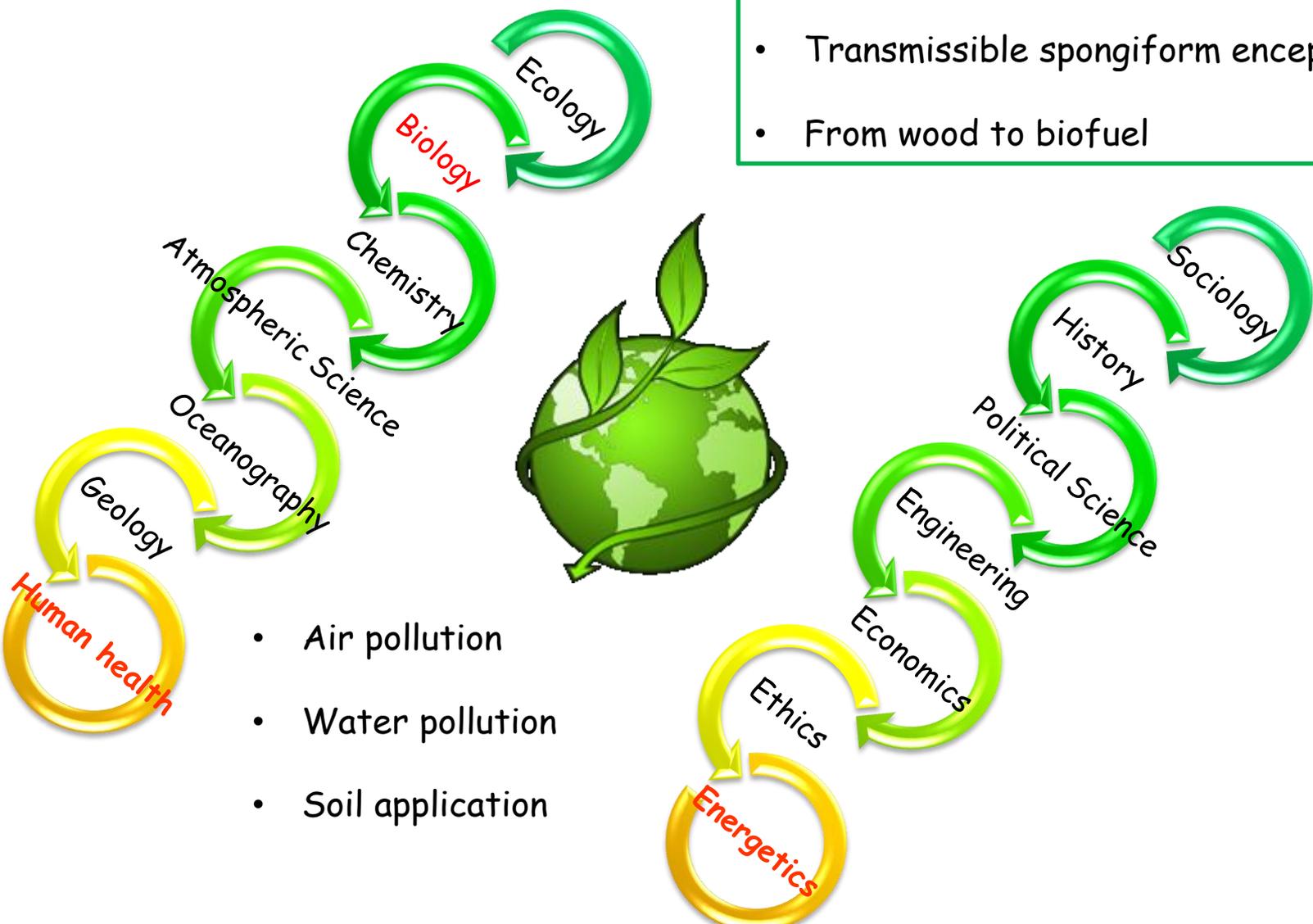
From M.J. Nasse *et al.*, *Nature methods*, 8:413 (2011)

# IRSR Beamlines in the World



# FTIR spectroscopy for environmental sciences

- Bacteria and biofilm characterization
- Transmissible spongiform encephalopathies
- From wood to biofuel

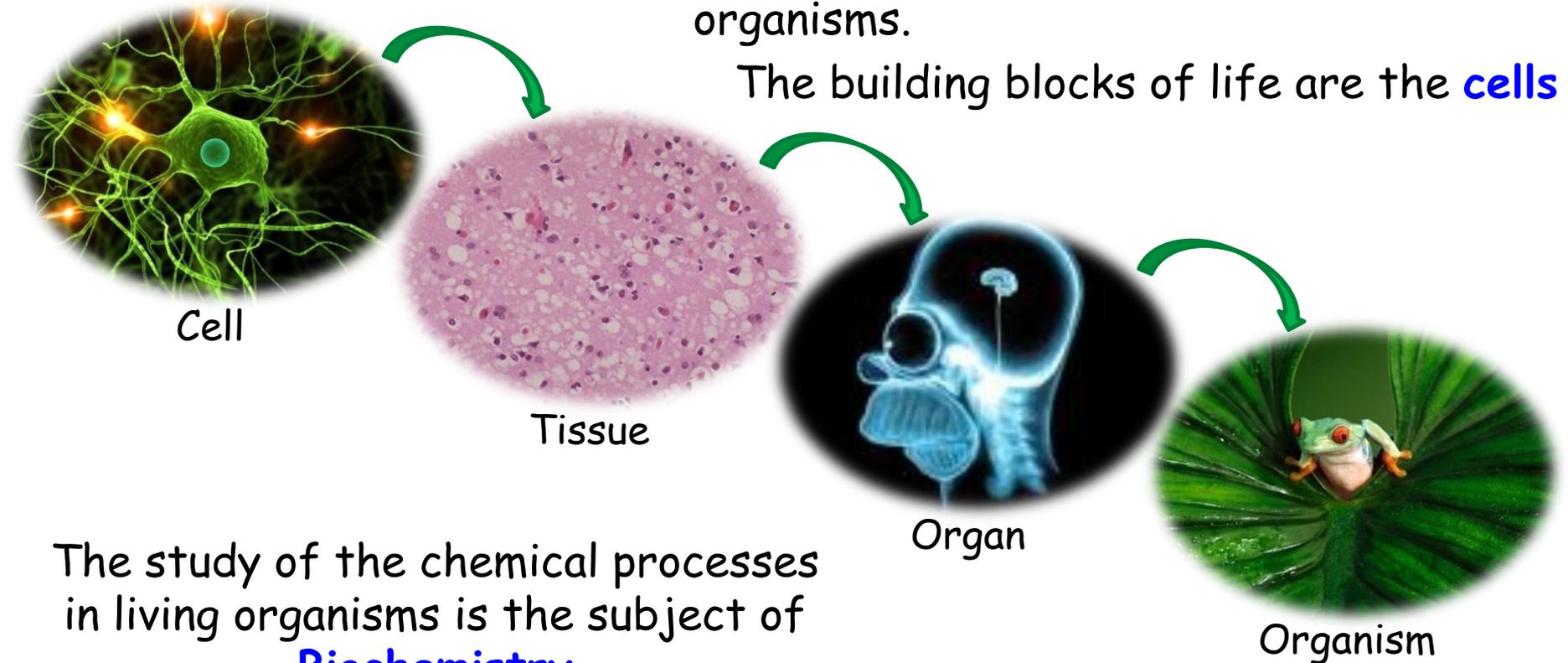


- Air pollution
- Water pollution
- Soil application

# Biology: The spectroscopic point of view

**Biology** is the branch of natural science that studies life and living organisms.

The building blocks of life are the **cells**



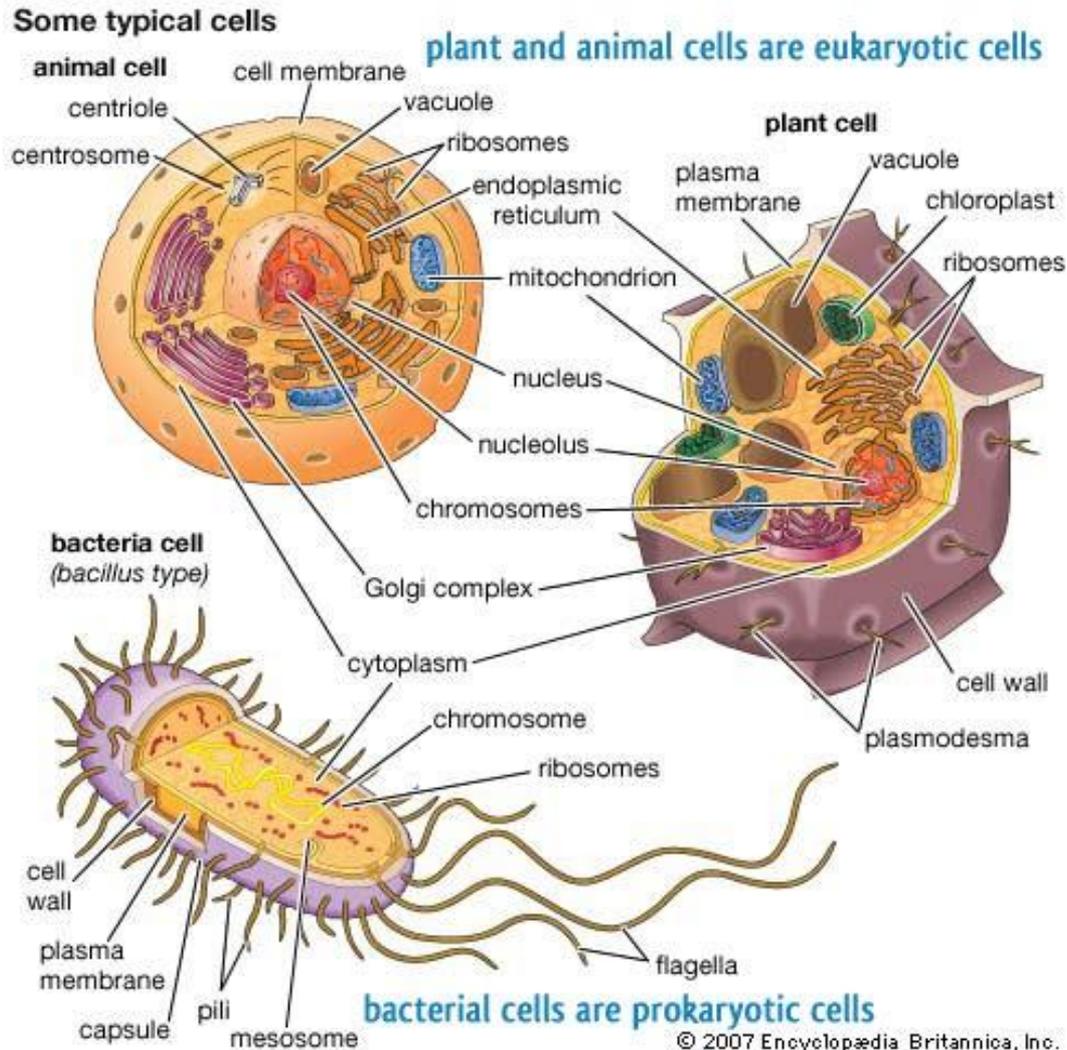
The study of the chemical processes in living organisms is the subject of **Biochemistry**

It deals with the structures and functions of cellular components such as **proteins, carbohydrates, lipids, nucleic acids** and other **biomolecules**

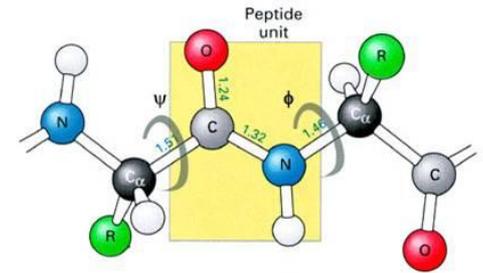
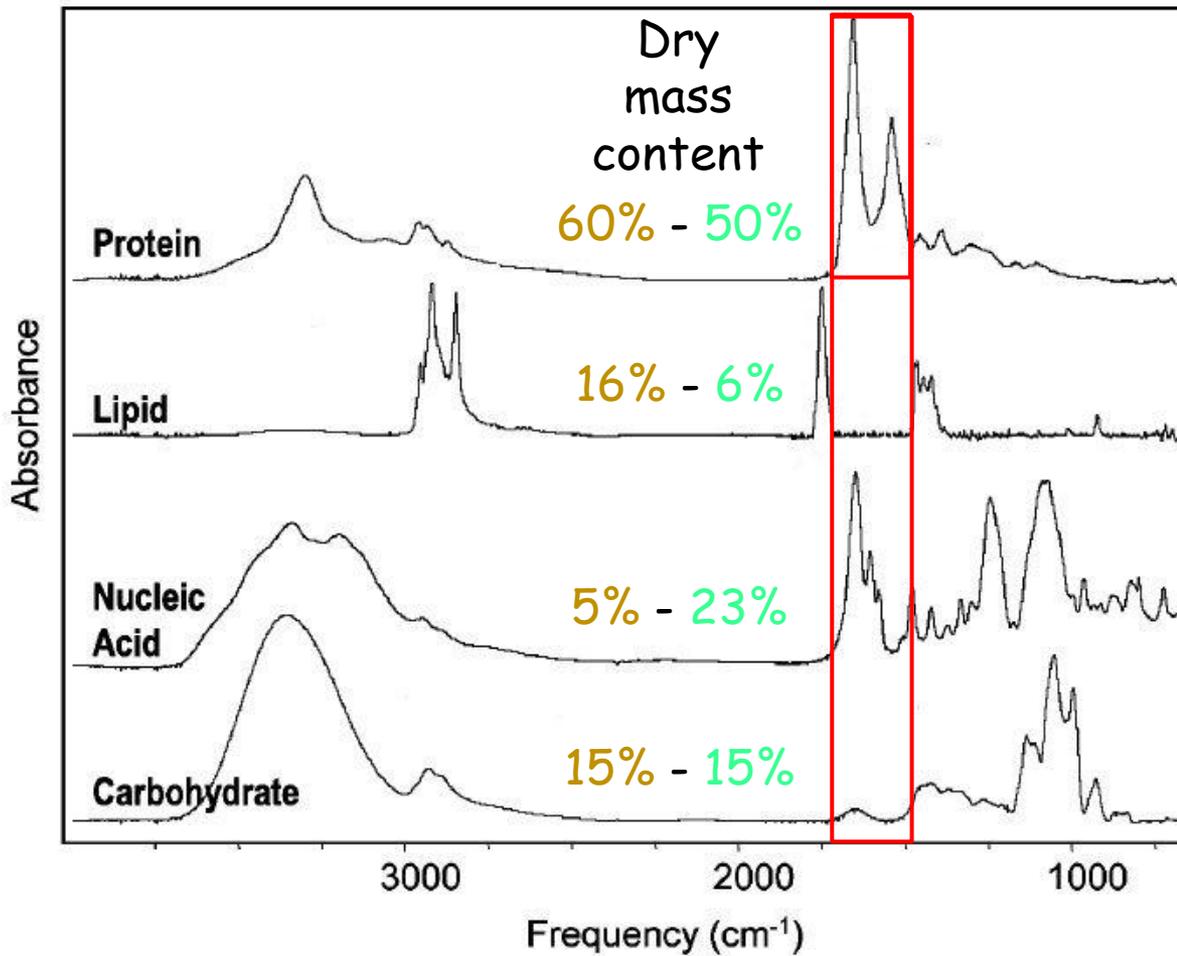
# The cellular architecture

## Prokaryotic versus Eukaryotic

### mammalian versus vegetal



# The eukaryotic/prokaryotic cell spectrum



**1700-1600**

1705-1690 :  $\nu$  C=O RNA

1660-1650 :  $\nu$  C=O DNA

1660-1650 :  $\nu$  C=O RNA

**1700-1600: Amide I**

80%  $\nu$  C=O + 10%  $\nu$  C-N  
+10%  $\delta$  N-H

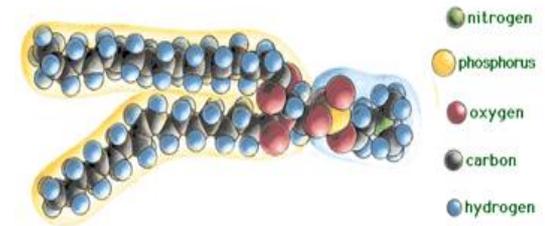
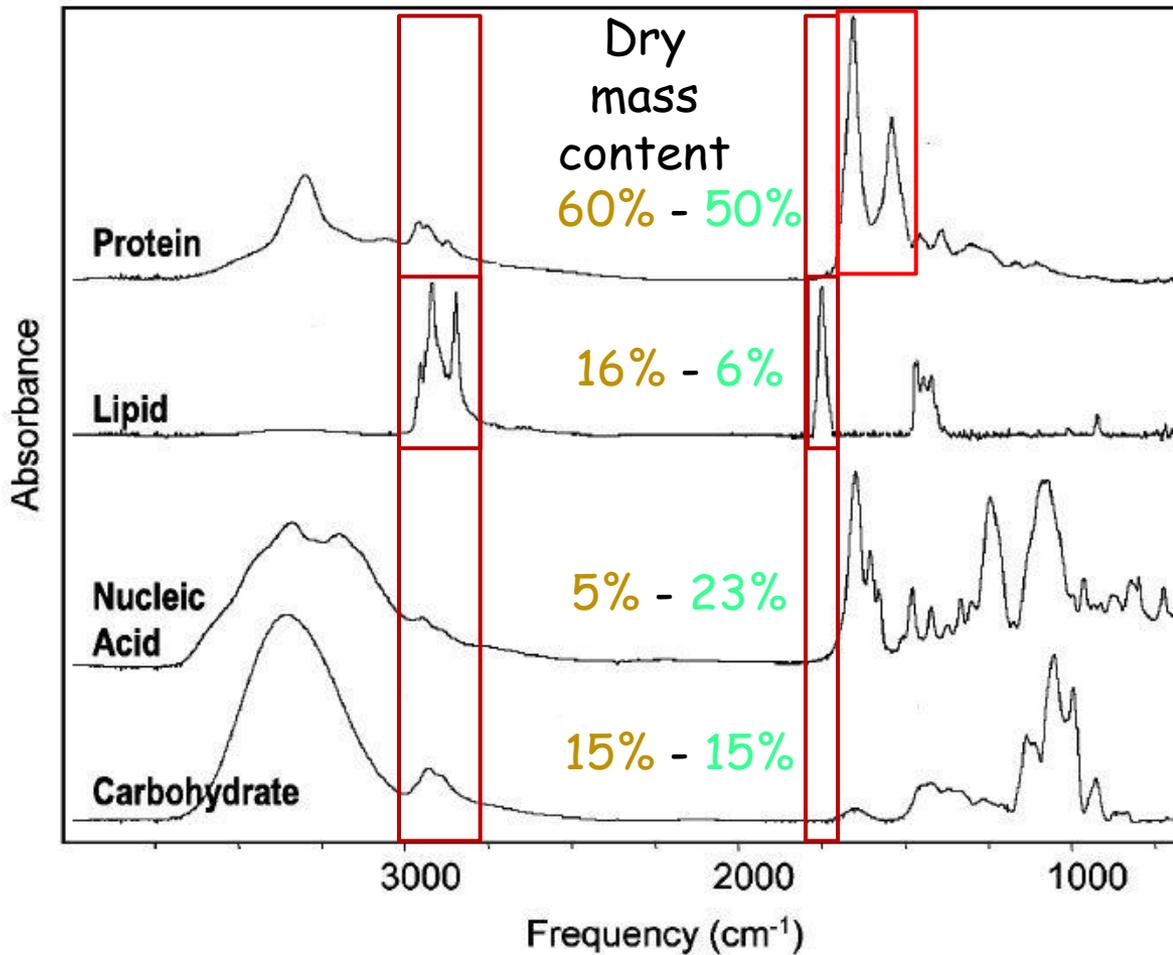
**1600-1500**

**1600-1500: Amide II**

40%  $\nu$  C-N + 60%  $\delta$  N-H

Adapted from: L. M. Miller, G.D. Smith and G. L. Carr, *Journal of Biological Physics*, 29 (2-3), 219-230, 2003

# The eukaryotic cell spectrum



## Saturated Acyl chains

2950-2960 :  $\nu_{as}$  (CH<sub>3</sub>)

2915-2925 :  $\nu_{as}$  (CH<sub>2</sub>)

2867-1877 :  $\nu_s$  (CH<sub>3</sub>)

2845-2855 :  $\nu_s$  (CH<sub>2</sub>)

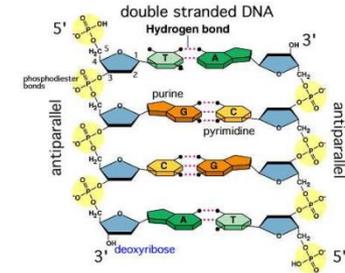
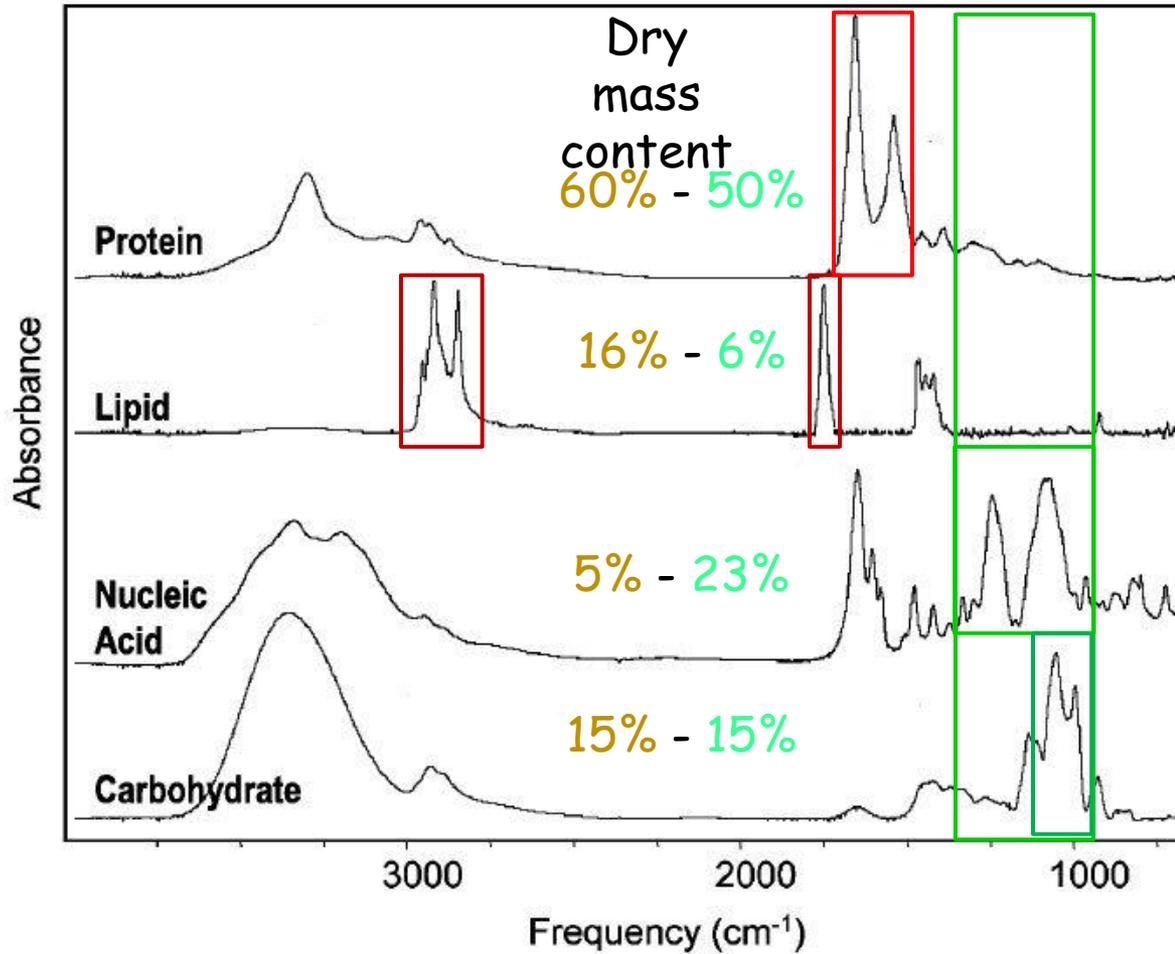
## Unsaturated Acyl chains

> 3000 :  $\nu$  (=CH)

## Ester Band

1700-1750 :  $\nu$  (C=O)

# The eukaryotic cell spectrum



1250-1200

1244 :  $\nu_{as}$  PO<sup>2-</sup> RNA

1230 :  $\nu_{as}$  PO<sup>2-</sup> DNA

1100-1050

1089 :  $\nu_s$  PO<sup>2-</sup> DNA

1084 :  $\nu_s$  PO<sup>2-</sup> RNA

1100-1000

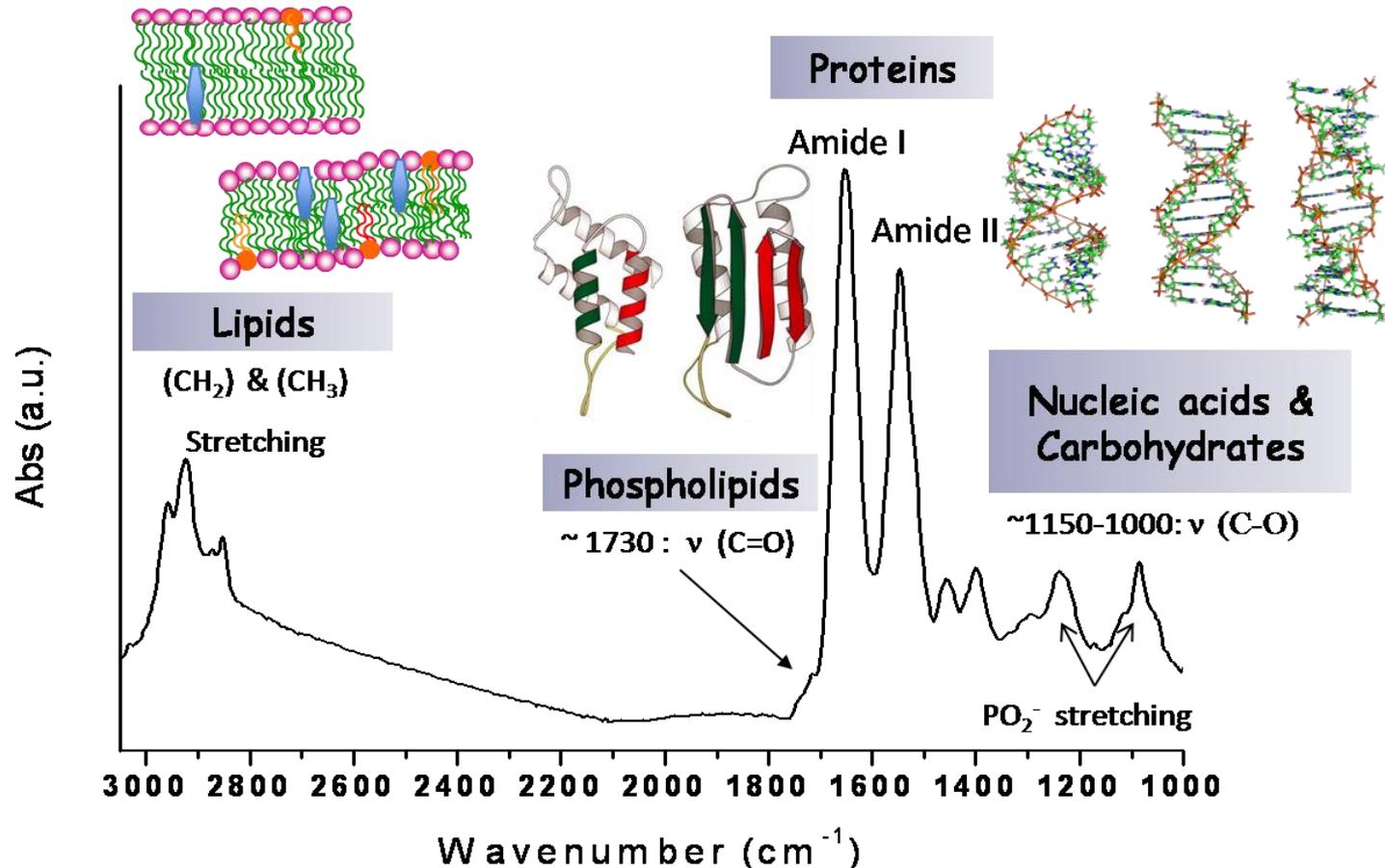
DNA and RNA ribose

$\nu$  C-O

Complex network of carbohydrate bands

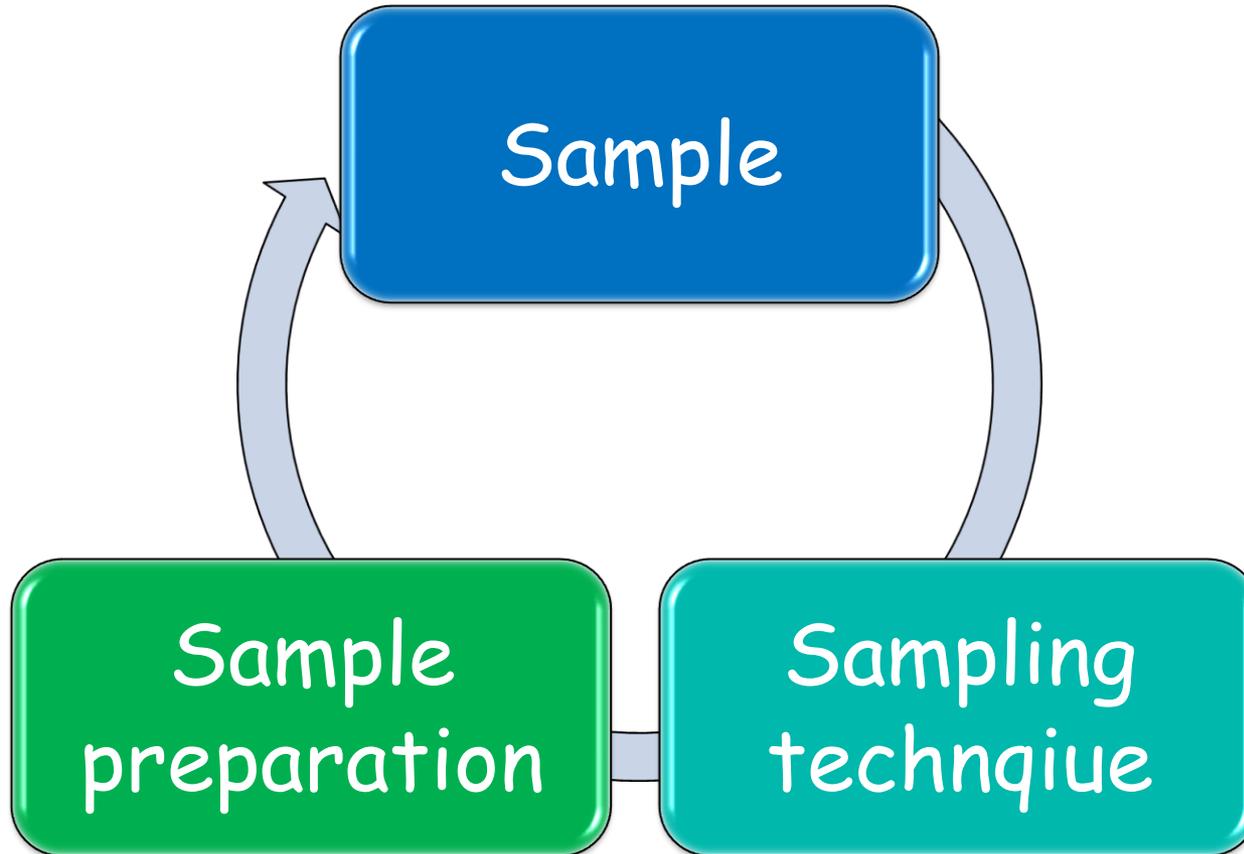
# The eukaryotic cell spectrum

Band intensity, position, width and shape (band components) are sensitive to subtle biochemical changes of bio-specimens.



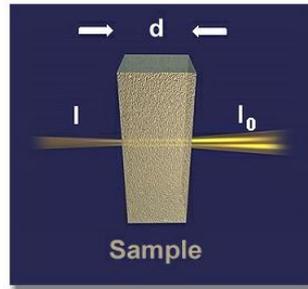
Compositional and structural information at tissue, cellular and sub-cellular level can be achieved by exploiting SR brightness advantage

# From the sample to the biological information



# Sampling techniques

- TRANSMISSION

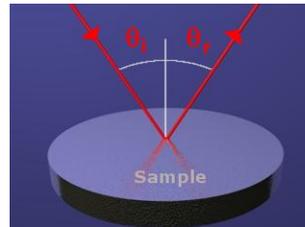


The absorption behavior of the entire sample is investigated

- TRANSFLECTION

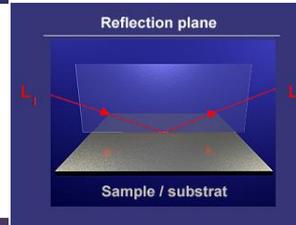
- REFLEXION

- SPECULAR



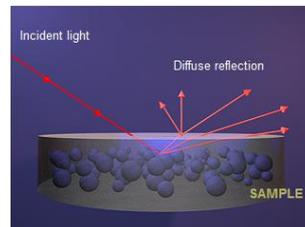
Typical angle of incidence = 10-30°  
The reflection behavior of the bulk sample is investigated

- GRAZING INCIDENCE



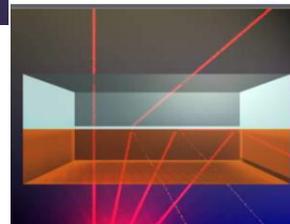
Typical angle of incidence = 50-85°  
The surface properties of the sample are investigated

- DIFFUSIVE



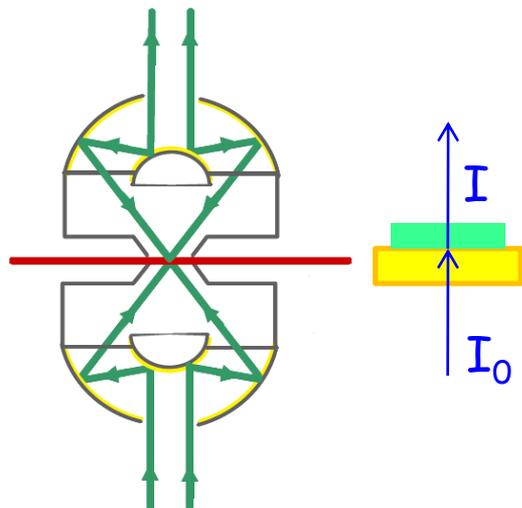
The diffusive-reflection spectrum is defined by the absorption-scattering behavior of the sample

- TOTAL (ATR)



The absorption behavior of the sample surface in contact with the IRE is investigated

# Transmission



+ Quantitative

## Lambert-Beer Law

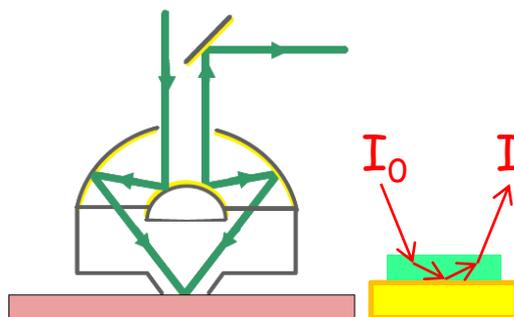
$$A = -\log_{10}(I/I_0) = \epsilon dc$$

$$[\epsilon] = L \cdot \text{mol}^{-1} \cdot \text{cm}^{-1}$$

$$[c] = \text{mol} \cdot L^{-1}, [d] = \text{cm}$$

- Thin samples
  - Limited penetration depth
- Substrate materials must be IR transparent → expensive

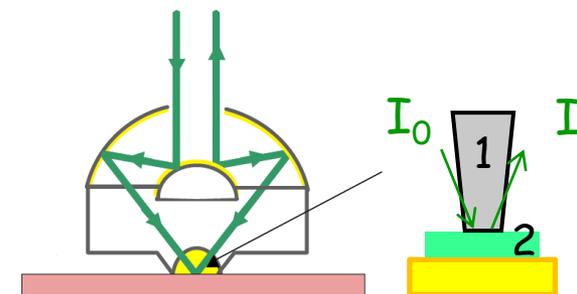
# Transflection



- + Substrate materials must be IR reflective → cheap
- + Improved absorbance signal

- Non quantitative
  - Electric field standing wave
- Thin samples
  - Limited penetration depth

# ATR



- + Surface sensitive technique

$dp$  (penetration depth) =

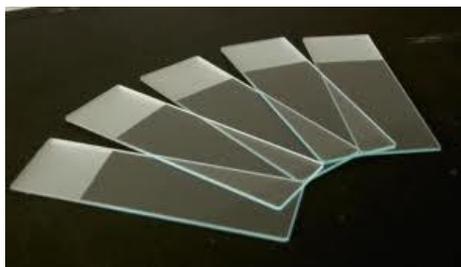
$$\frac{\lambda}{2\pi n_1 \sqrt{(\sin^2 \theta - n_{2/1}^2)}}$$

- + Substrate "unaffected"

- + Sample thickness "unaffected"
  - The absorbance is a function of  $\lambda$  and  $n_{2/1} = n_2/n_1$

# IR substrates

Sampling technique	Material	Vis-Transparent	MIR-Transparent	MIR-Reflective	Biocompatible	Other
Transflection	MirrIR-slides	Berely	No	Totally	Yes	Cheap +
Both	Si, Ge	No	Partially	Partially	Yes	Cheap
Transmission	Diamond	Yes	Partially	No	Yes	Expensive +
	BaF <sub>2</sub>	Yes	Totally	No	No	Expensive
	CaF <sub>2</sub>	Yes	Partially	No	Possibly	Expensive
	ZnSe	Slightly	Partially	No	No	Expensive
	Si <sub>3</sub> N <sub>4</sub>	Slightly	Partially	No	Yes	Fragile
	TEM grids	Yes	Totally	No	Yes	Fragile



MirrIR slides

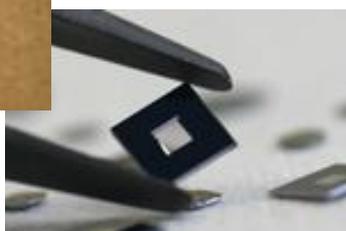
Si windows

ZnSe windows

CaF<sub>2</sub> windows

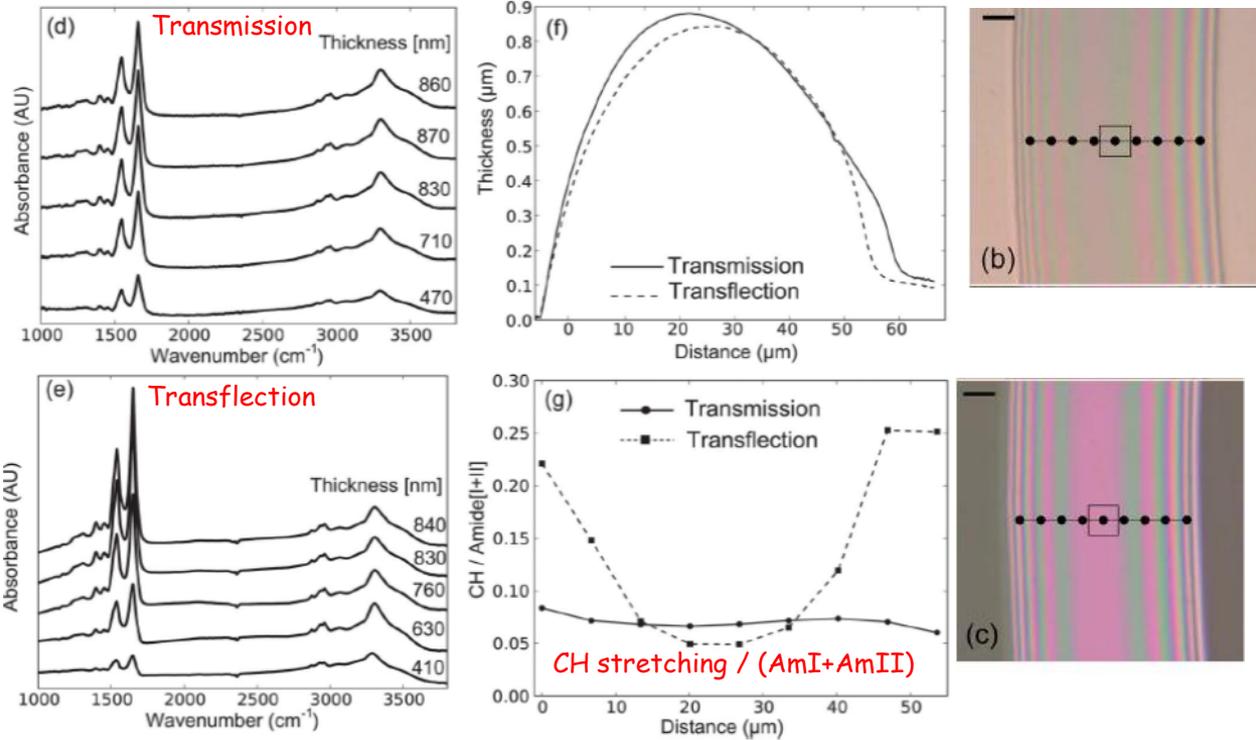


TEM grids



Silicon nitride windows

# Electrical field standing waves

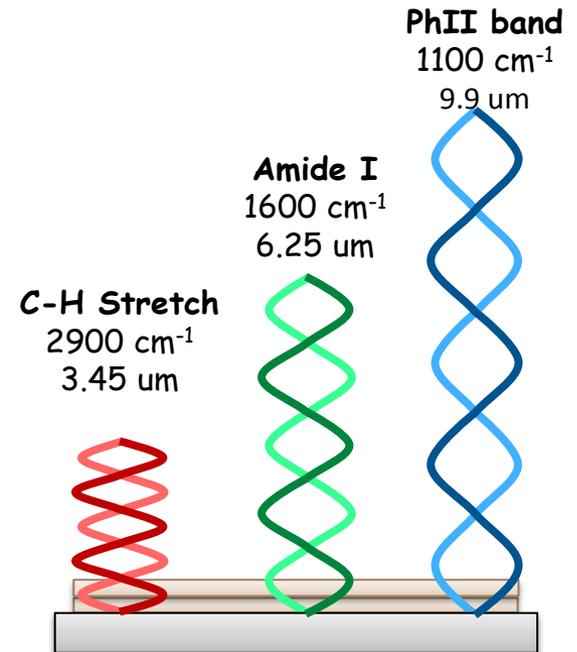


Curved films of BSA (Bovine Serum Albumin) protein have been measured at different profile points in transmission and transfection modes.

Transmission spectra behave as predicted by the Lambert-Beer law while transfection spectra do not.

Adapted from: Jacob Filik, Mark D. Frogley, et al., *Analyst*, 2012, 137, 853-861

- Reflection from a metallic surface induces  $\sim 180^\circ$  phase shifts of the electric field
- Incident and reflected waves interfere each other  $\rightarrow$  electric field standing waves
- Node (destructive interference) and antinode positions (constructive interference) depend on the wavelength (stated constant refractive indexes of substrate, sample and surrounding medium)
- This caused relative band intensity changes with thickness



See also: Paul Bassan, Joe Lee, et al., *Analyst*, 2013, 138, 144-157

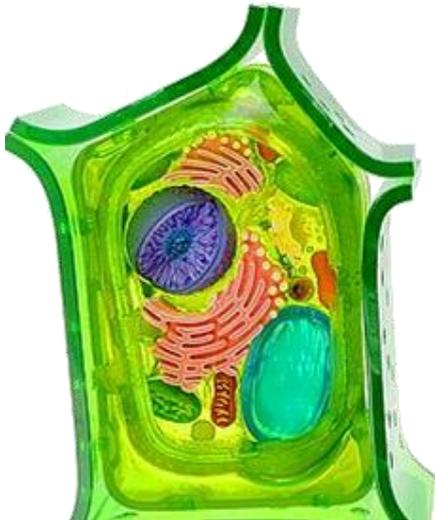
From the sample to biological information

Sampling techniques\_5

# Sample preparation



*Plant leaf*



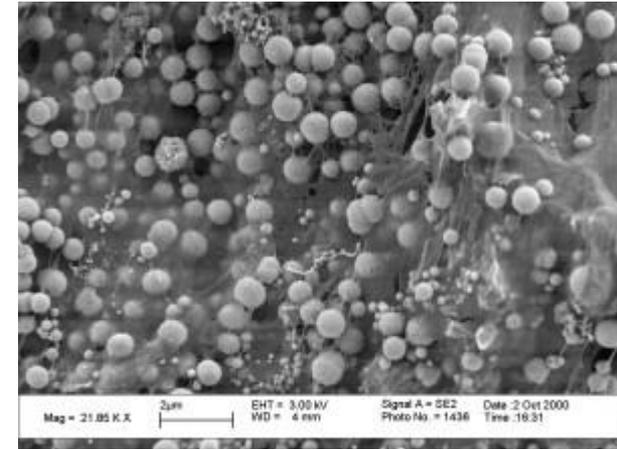
Bio-  
samples

Hydrated

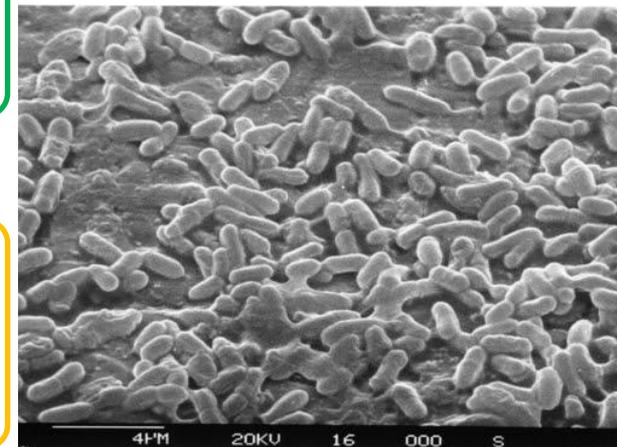
Dried

Fluidic  
devices

Fixed  
samples



Biofilm, a hydrated matrix of polysaccharide and protein formed by aggregates of bacteria.



# Fixed samples

The aim of fixation is to preserve the structural and biochemical constituents of cells in as close to in vivo conditions as possible

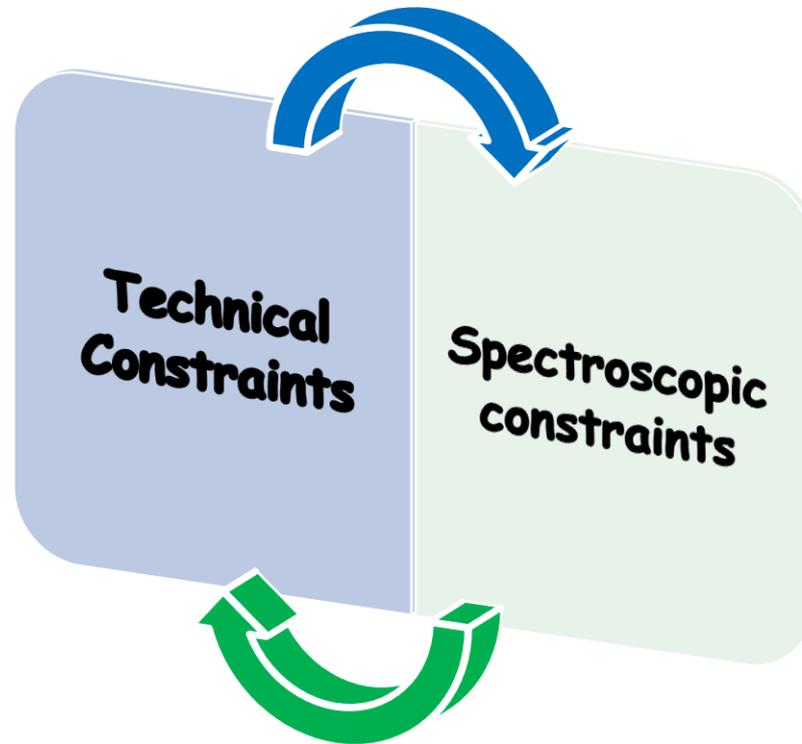
- *Air-drying* can cause collapse of internal cellular structures and activation of cell autolysis (dramatic variation of osmotic pressure within the cells).
- *Flash-freezing* followed by *cell lyophilisation* (freeze-drying) can not be applied to most common FTIR materials, such as  $\text{CaF}_2$  or  $\text{BaF}_2$ , since too brittle and with poor thermal contact.
- *Alcohol fixation* causes a rapid decrease in cellular volume, caused by the extraction of water from cells. Water is displaced from proteinaceous material, resulting in protein denaturation and organelle disruption. Alcohol extracts lipids from cells but has little effect on carbohydrates.
- *Formalin fixative* has bands potentially overlapping with cellular constituents bands (the most intense peak occurs at  $1000\text{ cm}^{-1}$ ); however it preserves most lipids and has little impact on carbohydrates. Formalin also appears to preserve protein secondary structure.

# Hydrated samples

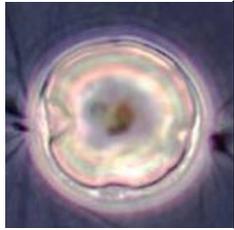
Hydrated sample measurements have been limited up to now by two major constrains

Manufacture of fluidic devices

The water absorption barrier

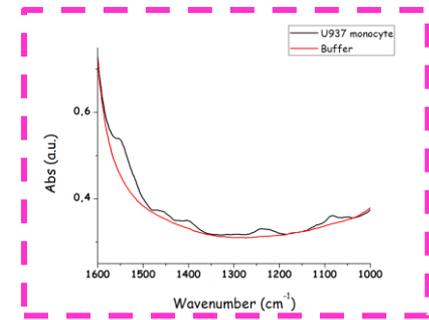
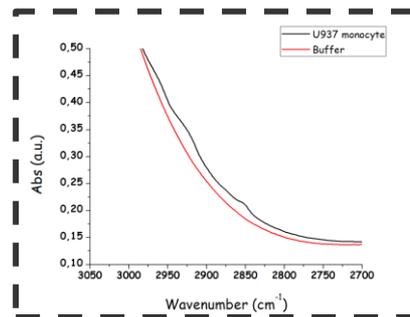
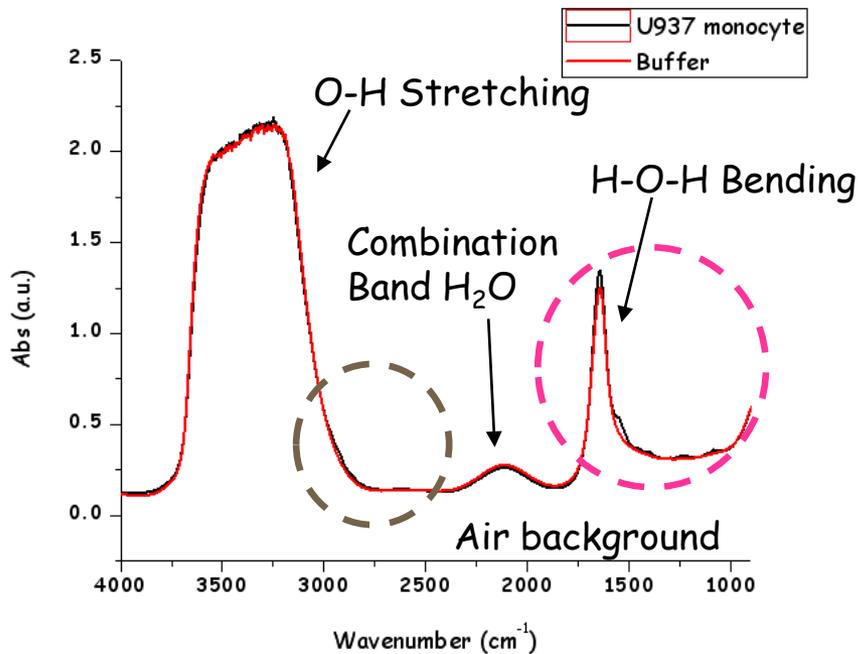
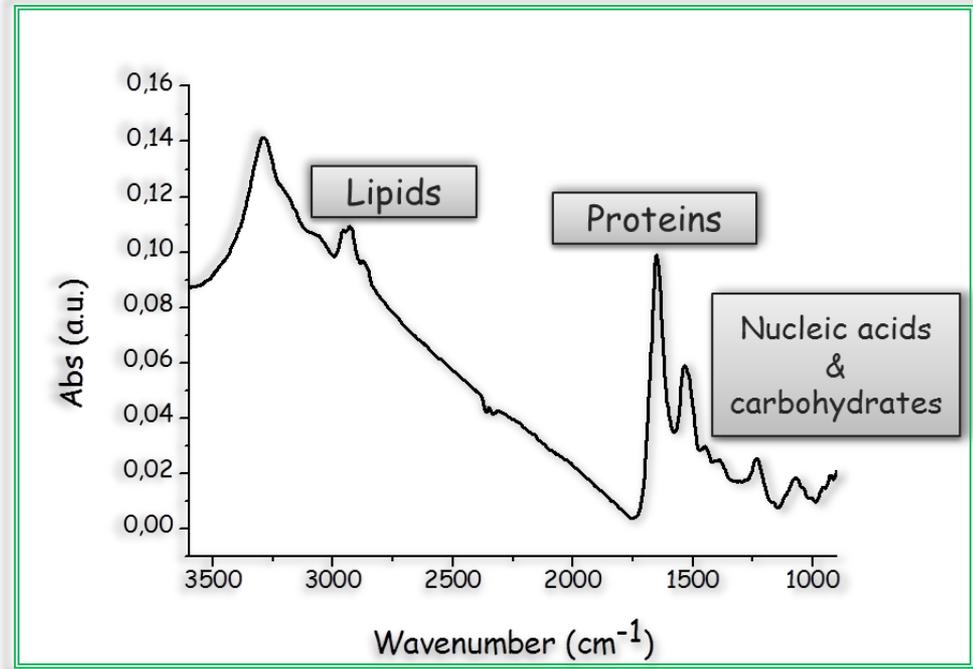


# The water absorption barrier



For the sampled volume:  
 ~ 48% extra cellular water  
 ~ 36% intra cellular water  
**~84% of water**

- Water: ~ 70% of the cell weight
- Dry mass: ~ 50% of proteins
- ~ 15% of carbohydrates
- ~ 15% of nucleic acids
- ~ 10% of lipids
- ~ 15% other molecules



$$\epsilon_{\text{water}} \sim 20 \text{ L} \cdot \text{mol}^{-1} \cdot \text{cm}^{-1}$$

$$\epsilon_{\text{Amide I}} > \epsilon_{\text{water}}$$

$$[C]_{\text{water}} \gg [C]_{\text{protein}}$$

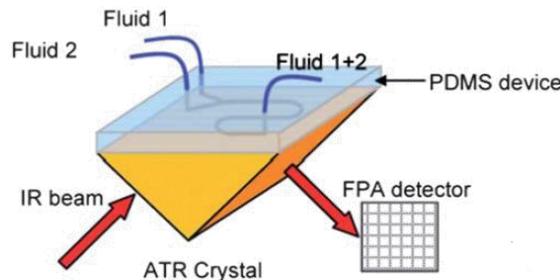
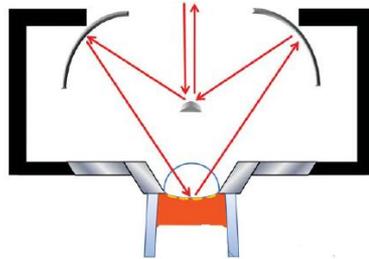
# Strategies for living cell sampling

For disclosing cellular IR features, and in particular protein Amide I band, the spectral contribution of water has to be limited:

- By reducing the sampling depth within the cell in **ATR** mode  
Suitable for prokaryotic cells, very thin adherent cells or special applications where the outermost cell layers are investigated

## Micro ATR geometries

From: Sergei G. Kazarian and K. L. Andrew Chan, Analyst, 2013, 138, 1940



## Demountable liquid cell

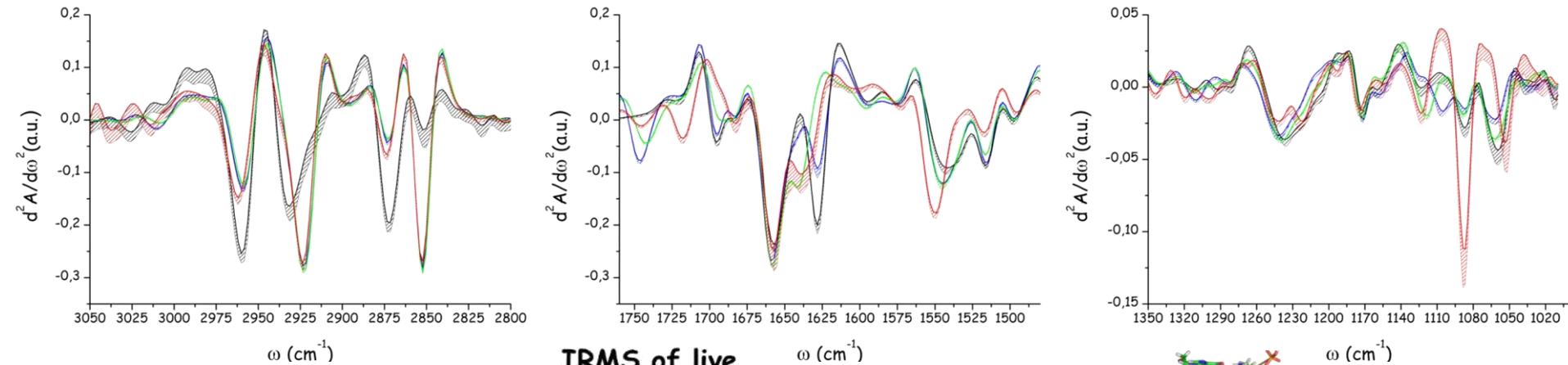
- By recording **transmission** (or **transflection**) spectra of living cells in liquid devices thinner than  $\sim 9$  microns for avoiding bending water band saturation and allowing water subtraction.
  - Demountable liquid cells, fabricated spacing apart two optical windows, do not assure i- the design flexibility needed for the realization of complex experiments; ii- the accurate control of the optical path, both locally and all over the device
  - A **microfabrication** approach is needed for controlling the water layer thickness and microchannel geometry.

Materials conventionally employed for IR transmission measurements are not standard for microfabrication

# Advantages offered by hydrated unfixed samples

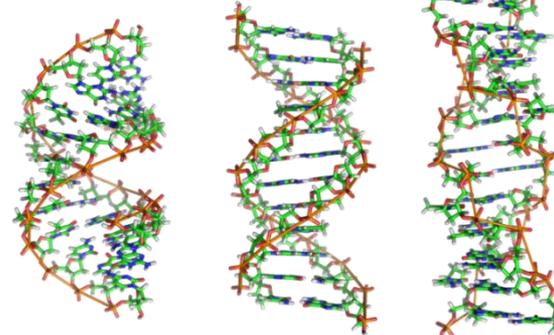
I. Hydrated species, both single cells and tissue section, are closer to the physiological conditions than de-hydrated fixed samples. More relevant biological information can be obtained from their spectra.

Fixatives induce alterations of both content and structure of sample, bio-macromolecules, detectable by IRMS



IRMS of live cells in MD

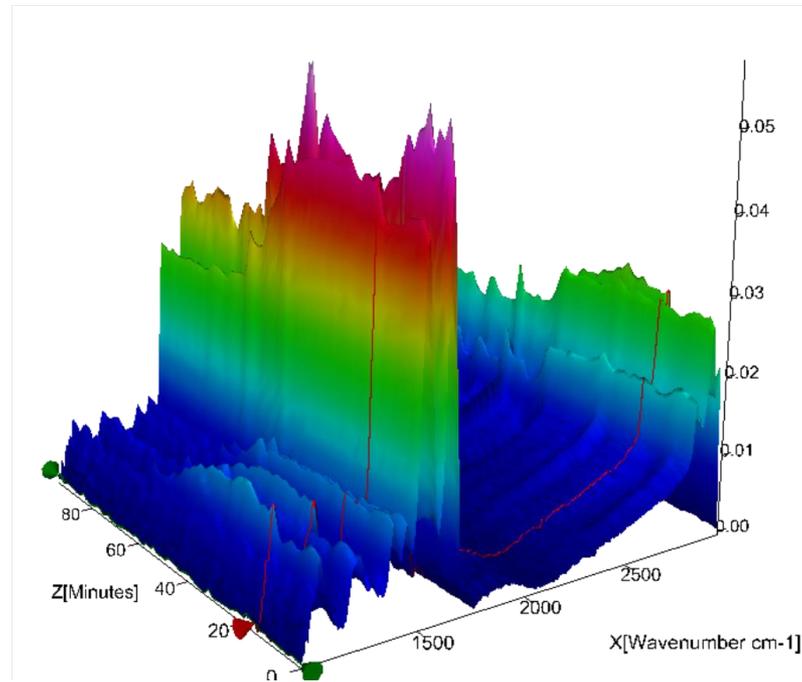
■ Formalin    ■ Air-Dried  
■ Ethanol    ■ Living



A-DNA    B-DNA    Z-DNA

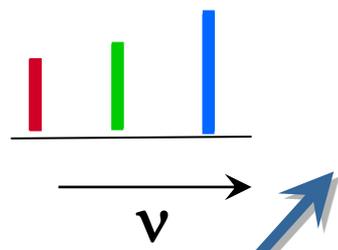
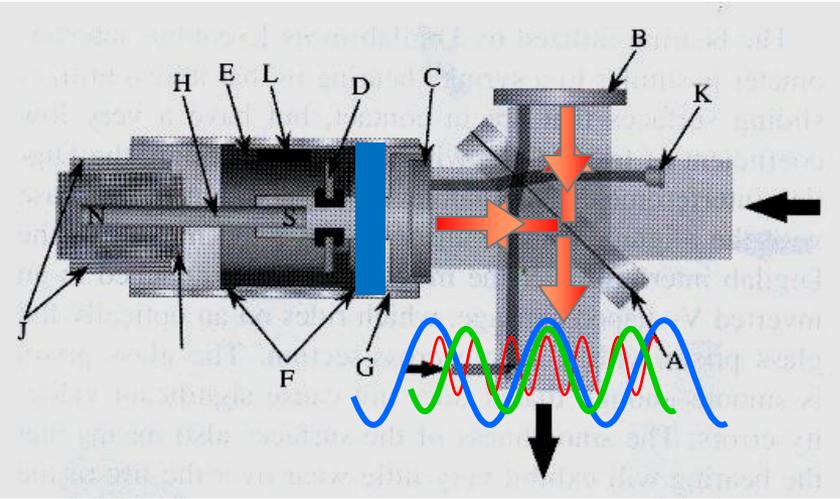
# Advantages offered by hydrated unfixed samples

II. Dynamic experiments can be performed, in order to monitor the response of a live system under physiological conditions toward different kind of stimuli

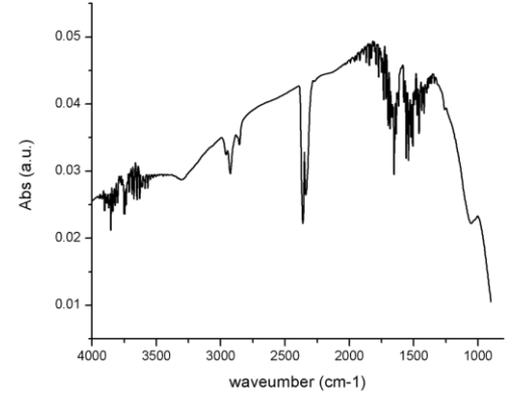


III. For transmission (or transflection) experiments, the closer matching between the refractive index of cell and water produces a great suppression of the scattering effects that dramatically affect single fixed cell spectra

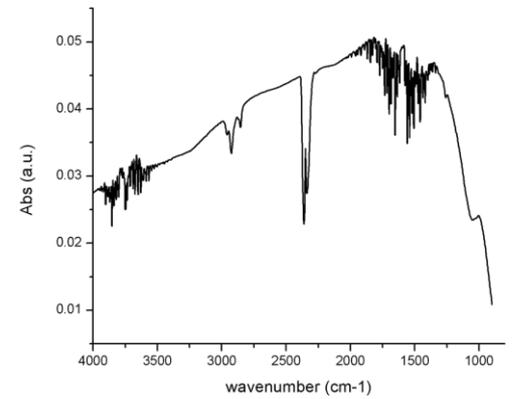
# Data acquisition



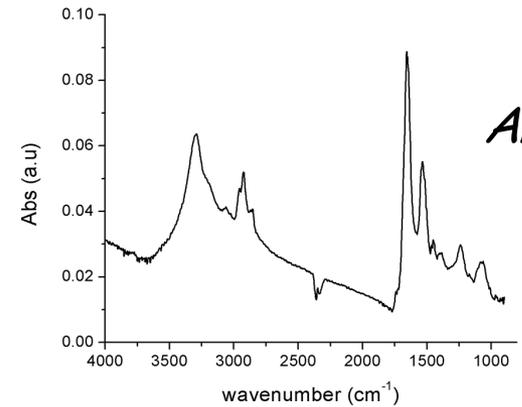
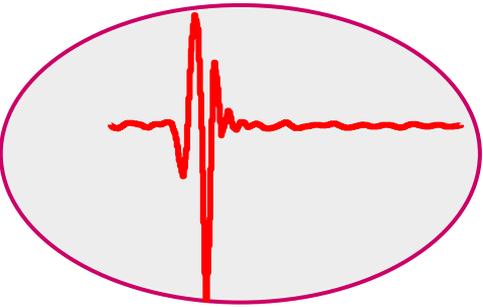
Sample Single channel



Reference Single channel



Fast Fourier Transform

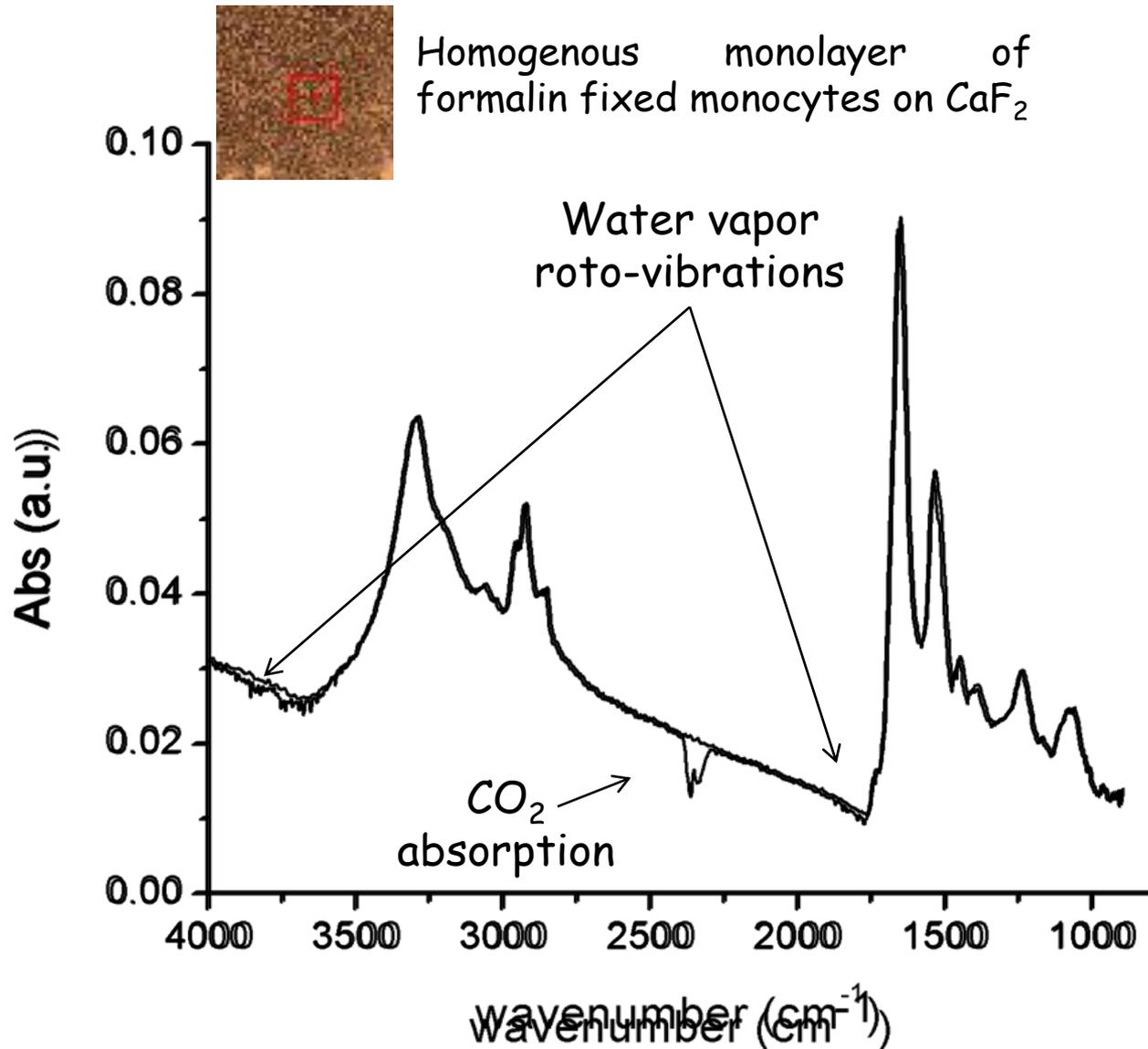


$$Abs = -\log\left(\frac{SSC}{RSC}\right)$$

From the sample to biological information

Data acquisition\_1

# Data preprocessing- Atmospheric compensation



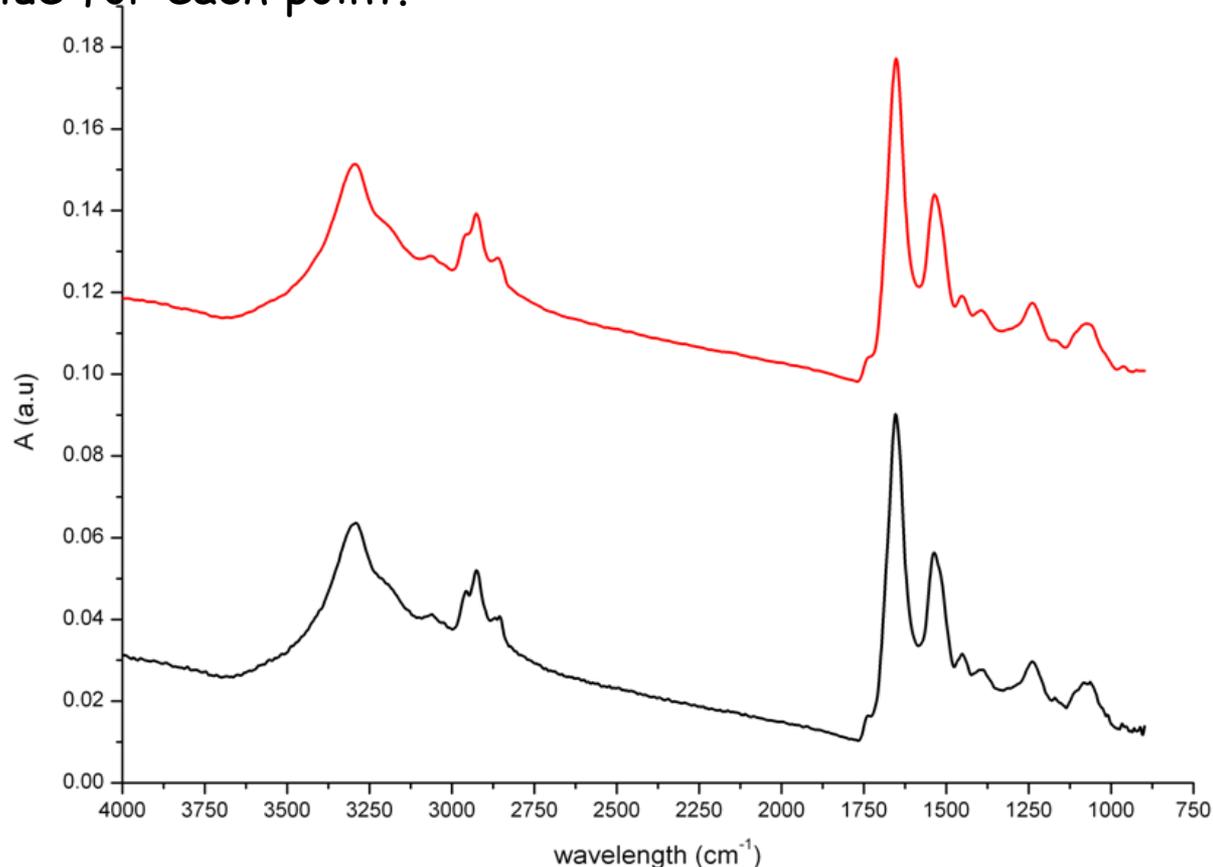
## Atmospheric compensated Spectrum

Spectral regions free of sample bands are used for evaluating water vapor and carbon dioxide content: 3600 to 4000  $\text{cm}^{-1}$  region for  $\text{H}_2\text{O}$  compensation, and 2300 to 2400  $\text{cm}^{-1}$  region for  $\text{CO}_2$ . Correction is extended over the entire spectral range.

# Data preprocessing- Spectral smoothing

**Noise reduction: spectral smoothing**  
**Savitzky-Golay method, K+1 smoothing points**

The method essentially performs a local polynomial regression (of degree  $k$ ) on a series of values ( $k+1$  points, equally spaced in the series) to determine the smoothed value for each point.



# Data preprocessing- Good Laboratory Practice

However, algorithms are NEVER better than GOOD LABORATORY PRACTICE!

Water vapor and carbon dioxide spectral contributions as well as the spectral noise should be reduced:

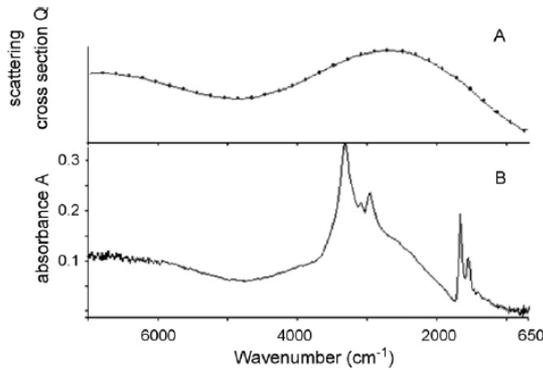
- Purging the interferometer with  $N_2$ /dry air or operating in vacuum
- Purging the microscope stage with  $N_2$ / dry air
- With a good conditioning of the laboratory environment



Too many people around the microscope!

- Implementing the S/N spectral ratio
  - Increasing the number of scans
  - Increasing the signal -> Brighter Sources -> Synchrotron Radiation
  - Reducing any possible source of noise: vibrations, electronic noise, .....

# Data preprocessing- Mie Scattering correction



Single cell spectra often present slow oscillations of the baseline. The origin is related to Mie-type scattering, which takes place when the wavelength of the incident radiation is similar to the one of the scattering particles.

→ **Position and intensity of IR bands can be affected**  
EMSC (Extended multiplicative scattering correction) corrects for these effects modeling the scattering object as a non-absorbing dielectric sphere..

$$Q = 2 - \left(\frac{4}{\rho}\right) \sin \rho + \left(\frac{4}{\rho^2}\right) [1 - \cos \rho]$$

$$\rho = 4\pi d(n - 1)/\lambda$$

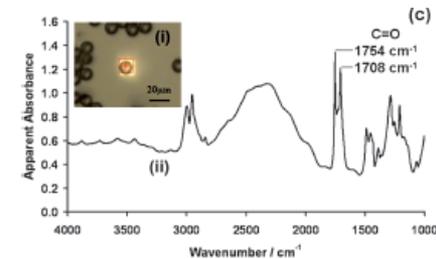
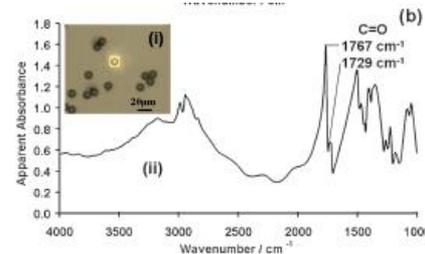
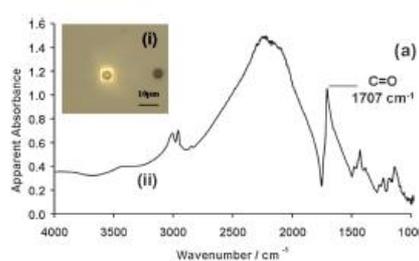
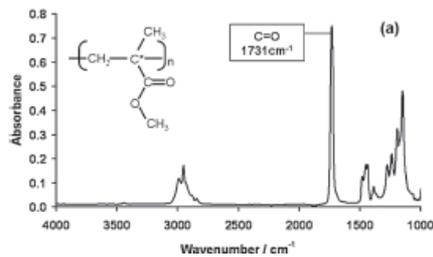
$Q$  Scattering efficiency  
 $d$  diameter of the sphere  
 $n=n_1/n_2$  refractive index of sphere and surrounding medium

B. Mohlenhoff et al, Biophysical Journal, 2005, 88, 3635-3640

## Resonant Mie scattering: scattering with simultaneous absorption

Paul Bassan,, Hugh J. Byrne at al., Analyst, 2009, 134(6), 1171-1175

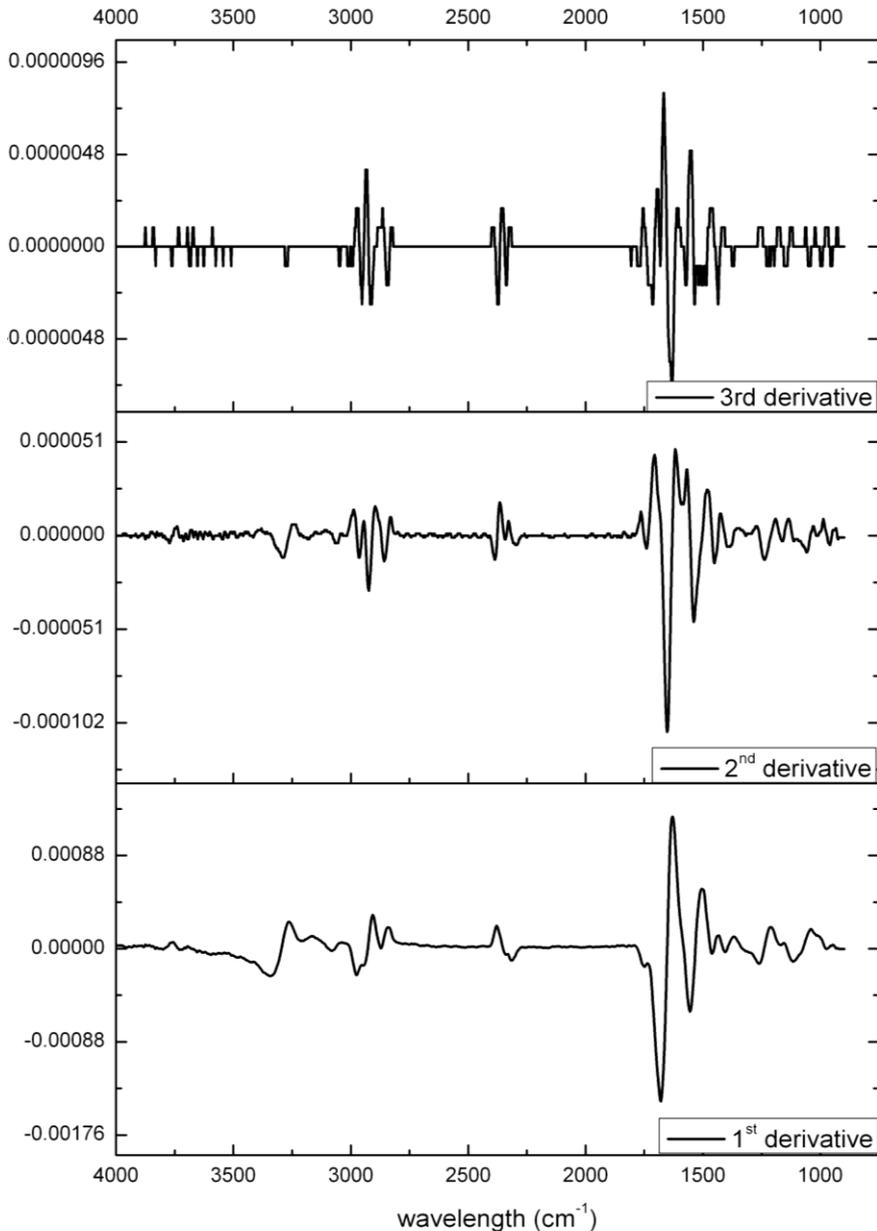
Paul Bassan, Achim Kohler, Analyst, 2010, 135, 68-277



Spectra of isolated single cells often exhibit significant **distortions of the band shapes**, especially a sort of derivative-like distortion on the high wavenumber side of the Amide I band.

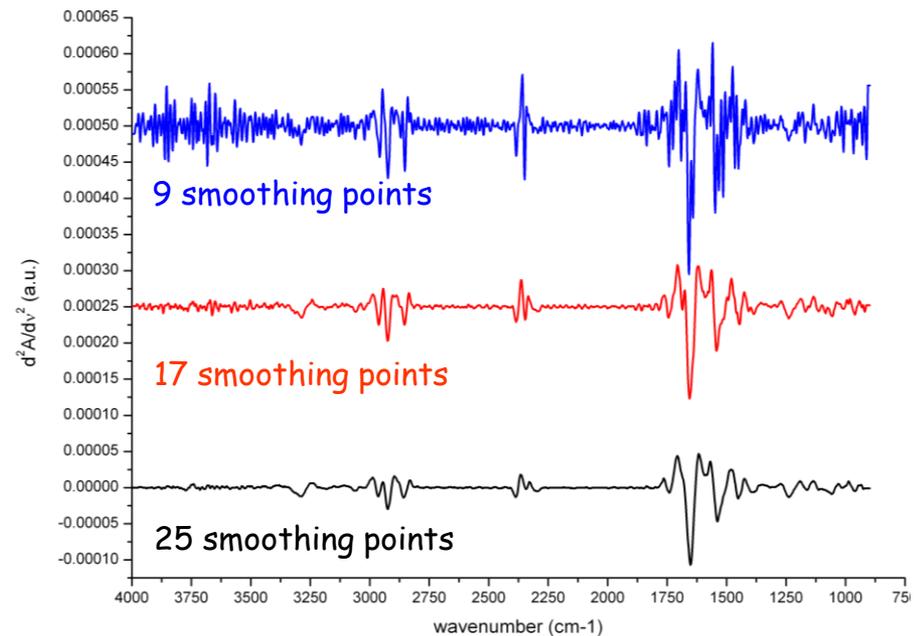
RMieS correction have been developed but still to be fully optimized.

# Data pre-processing- Spectra derivative



By using the derivative spectra, it is possible to minimize baseline variations and maximize spectral resolution. The derivative brings the overlapping peaks apart and the linear background becomes to a constant level.

In parallel, the spectral noise is also magnified, and the spectral complexity as well. Therefore, usually derivative algorithms take place with smoothing, commonly Savitzky-Golay.



# Data pre-processing- Spectra Normalization

The goal of normalization is to perform numerically what was not able to be performed physically during data collection; that is, recover exact replicates when no bio-logical differences exist. Normalization methods preserve the spectral shape, making easier the data analysis, but results depend on the spectral range.

## Constant shift - offset correction

When the replicates exhibit a simple vertical offset between spectra, normalization is achieved by subtracting a sample-specific constant from each.

## Scaling - Vector Normalization

If each spectrum of a data set contains a different amount of energy, large intensities are magnified more than low intensities. The scaling procedure produces a set of unit-length vectors in the wavelength space. For example, normalization allows to remove spectral intensity variations due to variation in sample thickness.

$$A_i(\text{vector normalizaed}) = \frac{A_i}{\|A_i\|}$$

## Standard Normal Variate (SNV)

SNV allows both spectral centering, since it produces mean-zero spectra, and their scaling

$$A_{ik}(SVN) = \frac{A_{ik} - \bar{A}_i}{\sqrt{\frac{\sum_{k=1}^p (A_{ik} - \bar{A}_i)^2}{p - 1}}}$$

$A_{ik}$  = SNV absorbance of the i-spectrum at  $\lambda_k$   
p = number of spectral points  
 $A_i$  = Average absorbance for all the  $\lambda_k$  (spectrum mean)

# Data Analysis

## Spectra comparison

### Spectral analysis "by visual inspection"

For small data sets (few spectra), spectra can be compared "visually" in order to highlight spectral similarities and/or differences affecting:

- Band position (band shifts), width (band broadening), shape (band components) and intensity.
- Ratios of peak areas (different proportion of most fundamental tissue-cell constituents).

**The reliability of biological conclusions drawn out from an experiment relies on the measurements of a statistically relevant number of samples.**

### Statistical analysis

Univariate and multivariate statistical analysis methods allow to compare a huge amount of spectra simultaneously, classifying them on the base of spectral similarities, affinities.

Univariate Methods: Average, standard deviation, regression techniques (PLS,....)

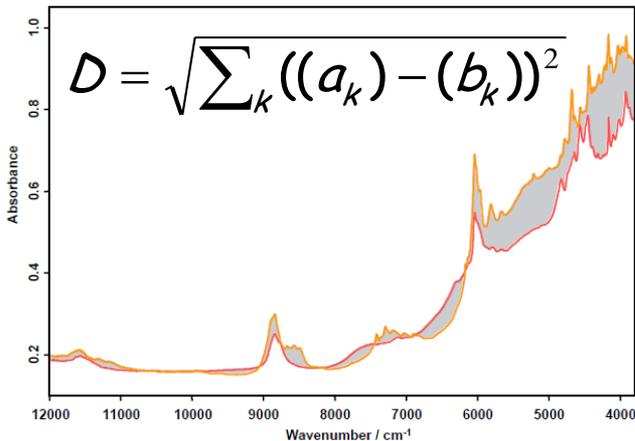
Multivariate Methods: **Cluster Analysis, Principal Component analysis (PCA), ...**

# 1- Cluster Analysis

## 1. Spectral distance calculation

Distance between spectra  $a$  and  $b$  can be calculated with many algorithms.

Euclidean spectral distance between  $a$  and  $b$  spectra is calculated over the all sampled  $k$  points.



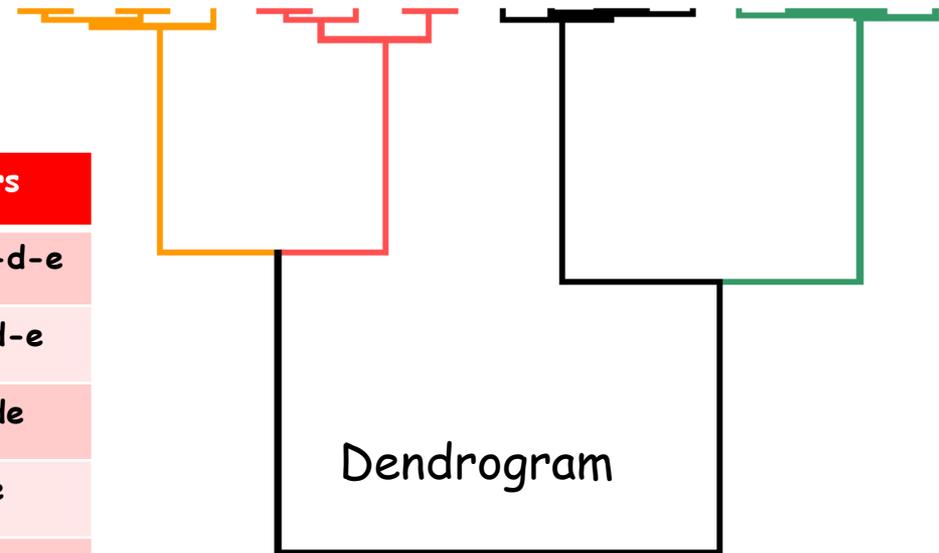
## 2. Spectral distance matrix

	a	b	c	d	e
a	0				
b	44	0			
c	11	54	0		
d	100	68	97	0	
e	120	92	115	21	0



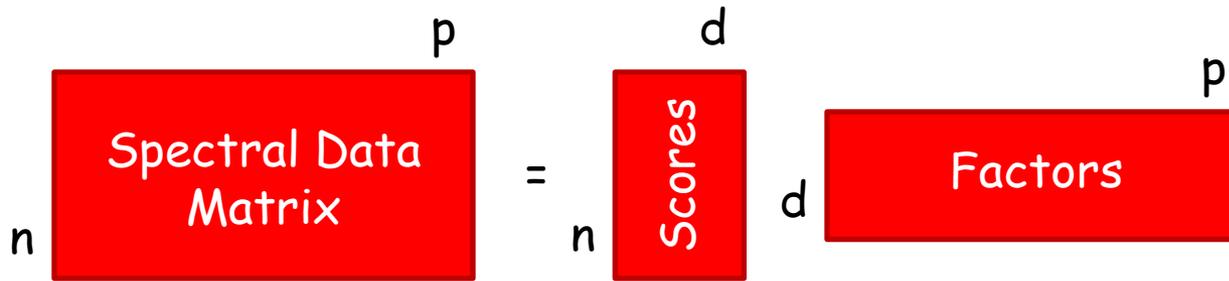
N	Clusters
1	a-b-c-d-e
2	ac-b-d-e
3	ac-b-de
4	abc-de
5	abcde

## 3. Spectra clustering

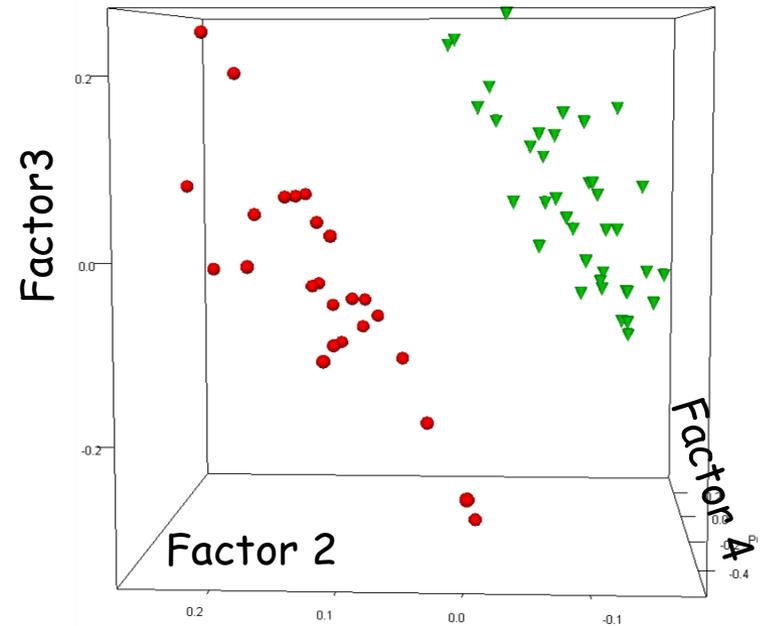
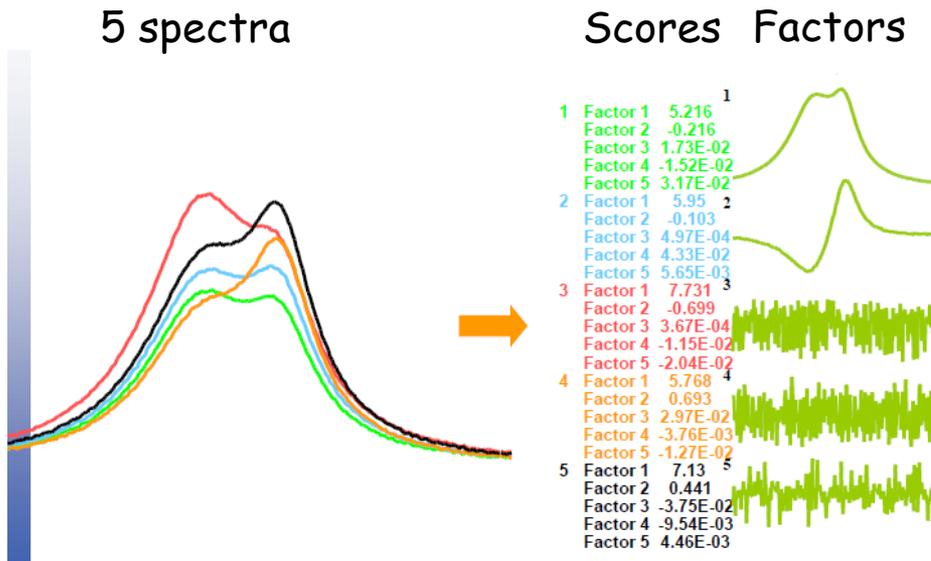


There are many methods available to calculate spectral distances between a newly-created cluster and all the other spectra or clusters.

# 2- PCA - Principal component Analysis



$n$  spectra with  $p$  data points;  $d$  scores for each spectrum ( $d < n$ );  $d$  factors with  $p$  data points ( $d < n$ )



# In Situ FT-IR Microscopic Study on Enzymatic Treatment of poplar Wood Cross-Sections

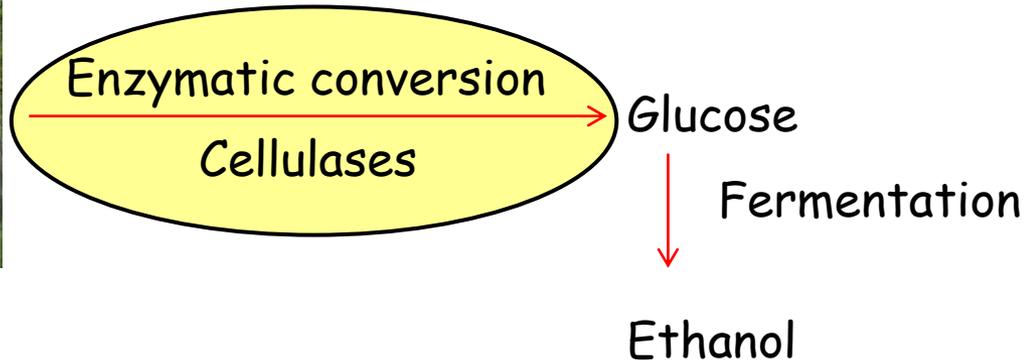
N. Gierlinger et al., *Biomacromolecules* 2008, 9, 2194-2201

Cellulose is the major polymeric component of the plant matter and is the most abundant polysaccharide in Earth.

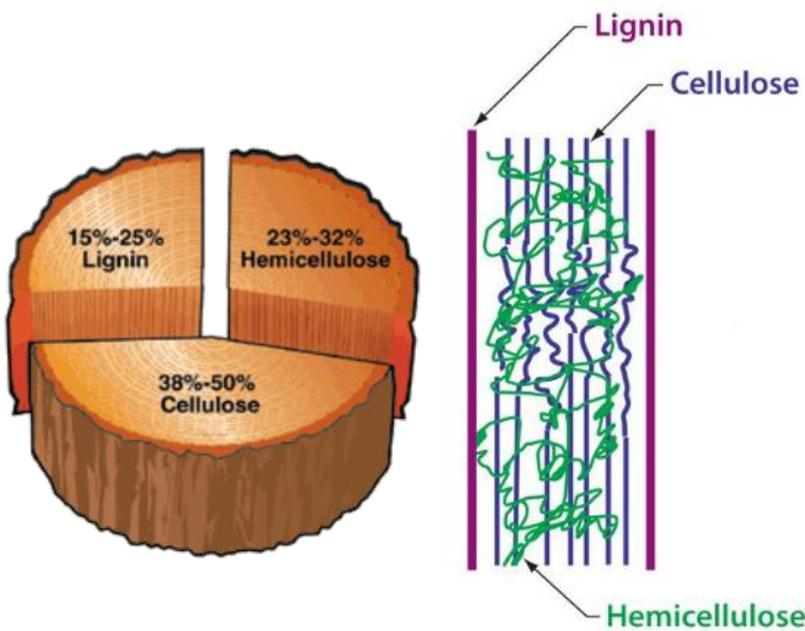
## Biofuels from Cellulosic Biomass

Purpose is to convert cellulosic biomass to fuels such as ethanol, methanol, dimethyl ether, or gasoline.

Poplar are a common source of clean biomass

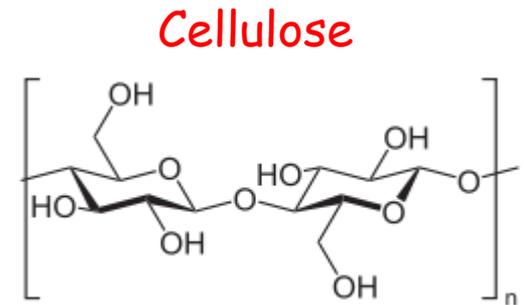


The rate of efficiency of cellulose hydrolysis is affected by many factors: enzymatic system, substrate characteristics (degree of polymerization, crystallinity, pore size) and cellulose association with hemicellulose and lignin.



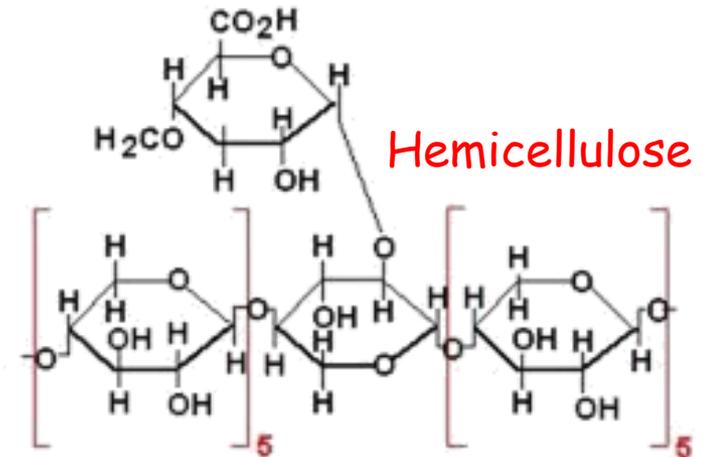
Within a plant cell wall, cellulose is embedded in a matrix of other polysaccharides (hemicellulose) and lignin

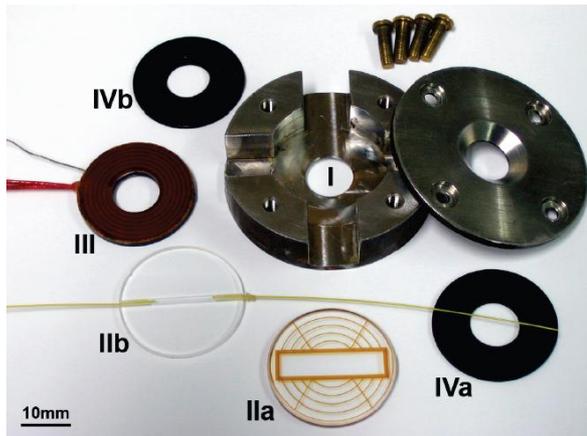
**Cellulose** is a homopolymer of glucose consisting of  $\beta(1-4)$  bonds. It differs from another polymer of glucose, starch that consists of  $\alpha(1-4)$  bonds



**Lignin** is a complex chemical compound most commonly derived from wood. It is a racemic, heteropolymer consisting of three hydroxycinnamyl alcohol monomers differing in their degree of methoxylation

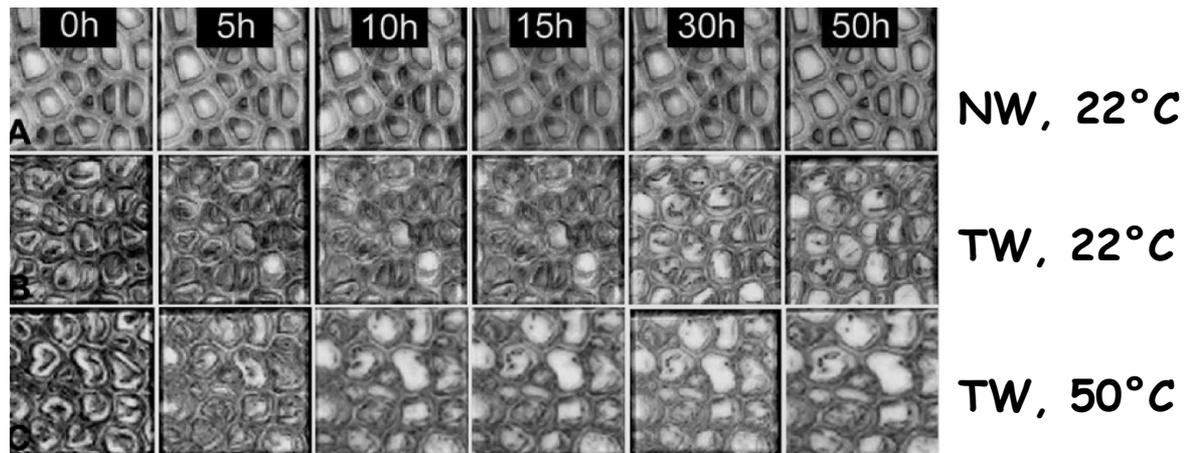
**Hemicellulose:** Heteropolymer of pentoses (xylose and arabinose) and hexoses (glucose, galactose, mannose) and sugar acids (acetic).

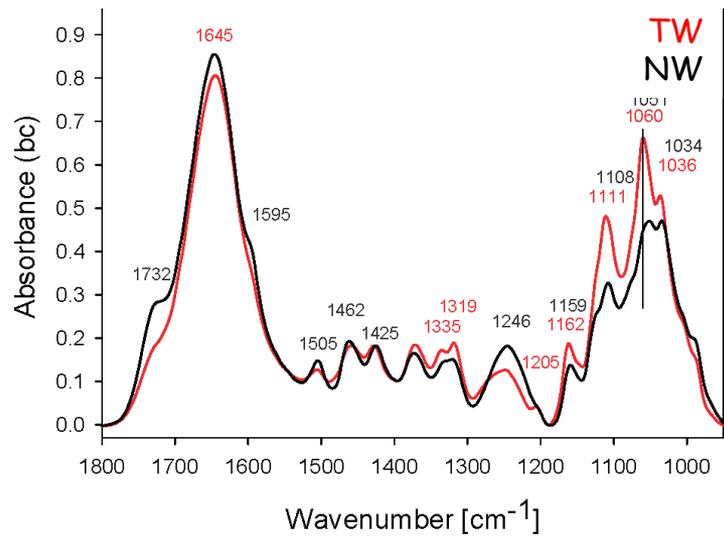




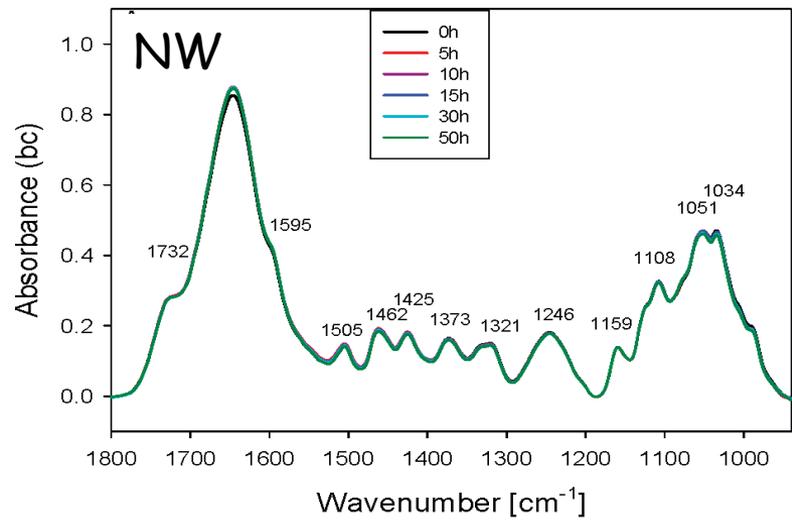
The enzymatic degradation of polar wood samples induced by a multicomponent commercial enzyme has been studied. A fluidic cell with 10 micron optical path has been used that allows i- T control and ii- waste removal. Both normal (N) and tension (T) wood have been considered.

Tension wood is a type of wood which forms in angiosperms in response to environmental stresses. It is characterized by a gelatinous cellulose layer, *G*-layer, and has an extra 50% cellulose content with respect to normal wood. For the tree, the purpose of tension wood is to help the tree stay stable, and to keep the tree upright. This type of wood is not useful for people who work with wood to make flooring, furniture, and other products, because it has an irregular texture, but it should be ideal for biomass synthesis.

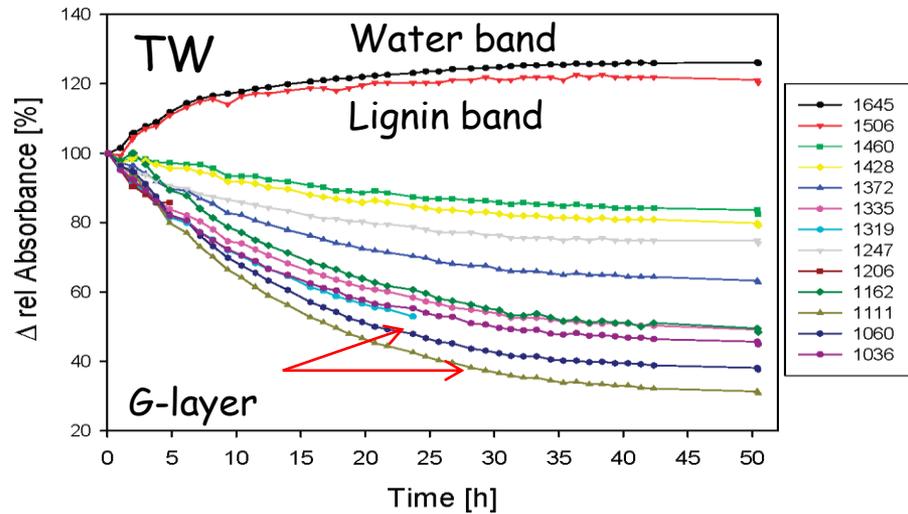




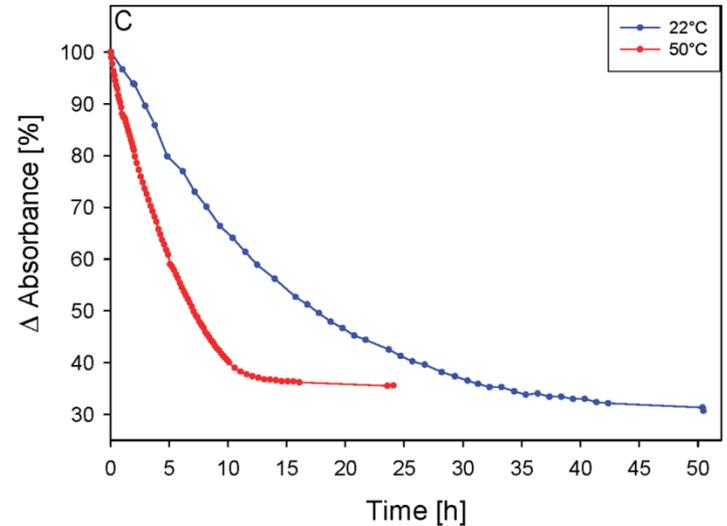
TW and NW have peculiar spectral patterns



NW spectra does not change in time. The enzyme is barely efficient toward cellulose of cell walls



TW bands mostly affected by cellulase enzymes are related to G-layers

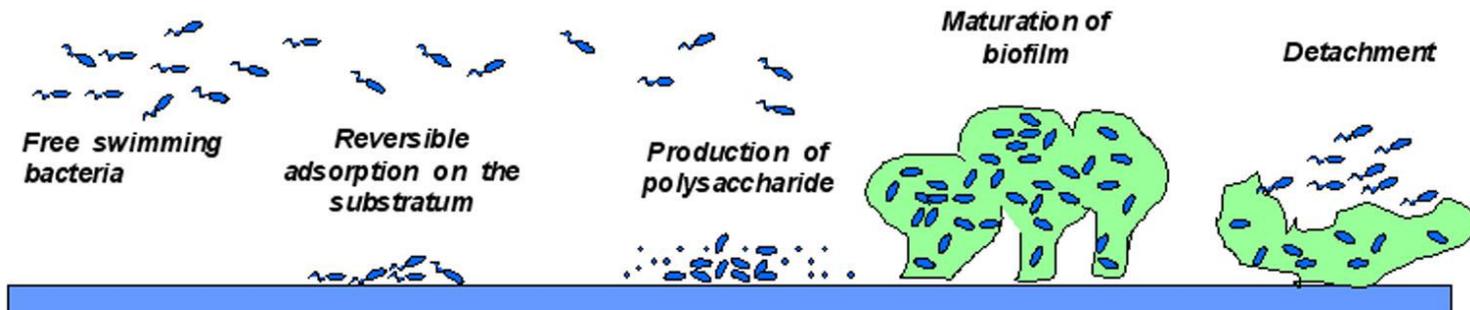


Temperature is reducing to almost 1/3 the degradation time of G-layer in TW

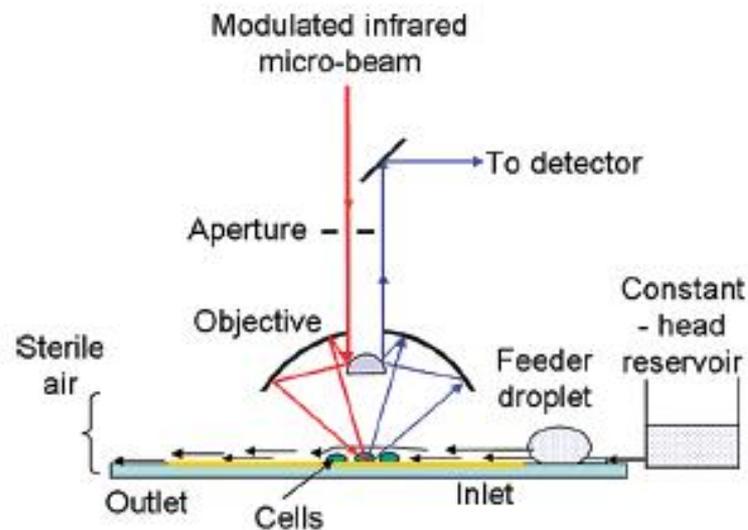
# Real-time Chemical imaging of bacterial activity in biofilms

From H-Y H. Holman et al., *Analytical Chemistry*, 81:8564 (2009)

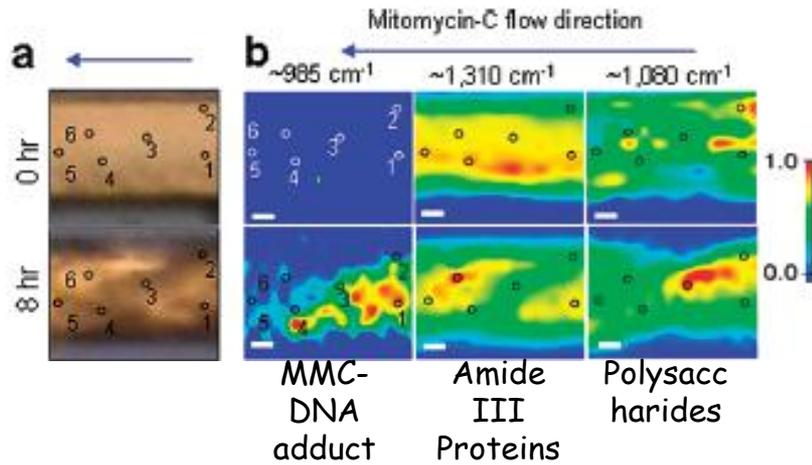
"Bacterial biofilms are structured dynamic communities of aggregated cells enclosed in a self-produced polymeric matrix that adheres to both inert and living surfaces in aqueous environments." [http://www.lcs.syr.edu/academic/biochem\\_engineering/research\\_areas/biomaterials\\_tissue\\_engineering.aspx](http://www.lcs.syr.edu/academic/biochem_engineering/research_areas/biomaterials_tissue_engineering.aspx)



"When opportunistic pathogenic bacteria develop biofilms, they can become up to 1000 times more resistant to antibiotics. The formation of biofilms and their resistance to antibiotics and host immune attacks are at the root of many persistent and chronic bacterial infections. Real time monitoring of bacterial activity at a chemical level during biofilm initiation, growth, release and bacteria-drug interactions in real time as the processes are happening could lead to new preventive and curative treatments."



# Uptake of mitomycin-C (MMC) by Escherichia coli within a biofilm



In points 1 and 2 (closer to MMC source) as well as 3 and 4 (Amide III richer) → Higher concentrations of MMC-DNA adduct  
 In points 2 and 3, increased polysaccharides and decreased Amide III  
 In point 6 (lower concentration of MMC-DNA adduct) the Amide II signal increases.

Cellular diversification processes in response to MMC toxicity

## Dynamics of biofilm formation in micro-channels

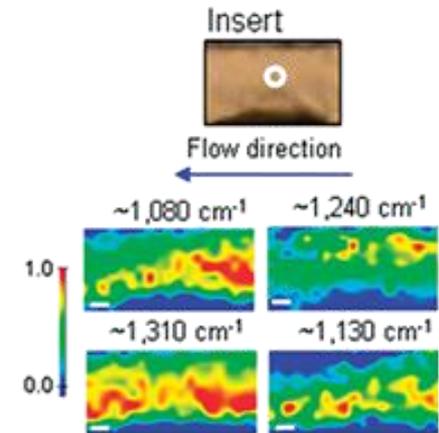
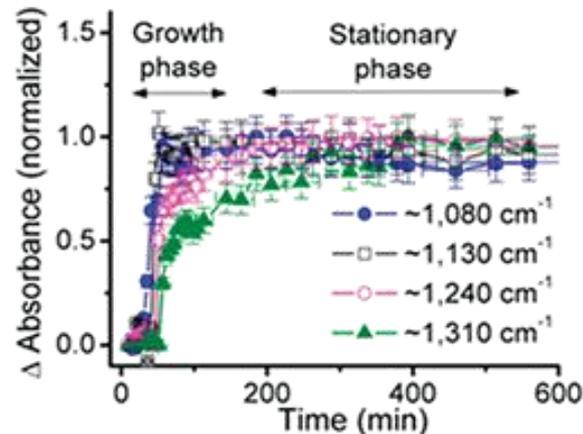
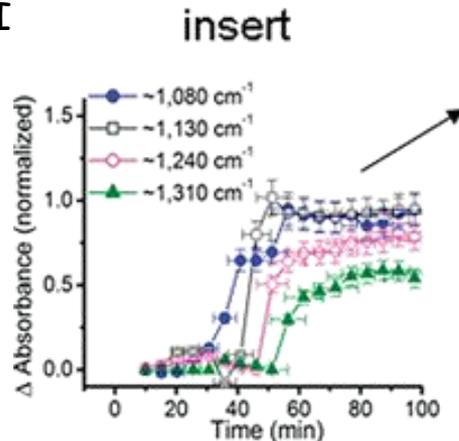
1080  $\text{cm}^{-1}$  polysaccharide

1130  $\text{cm}^{-1}$  glycocalyx

**Glycocalyx** refers to extracellular glycoprotein produced by some bacteria

1240  $\text{cm}^{-1}$  DNA/RNA

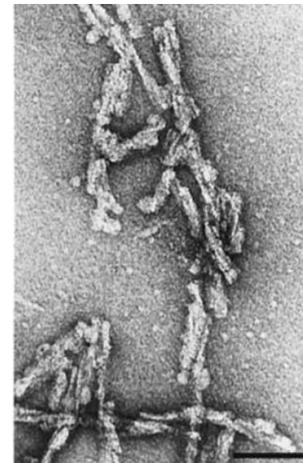
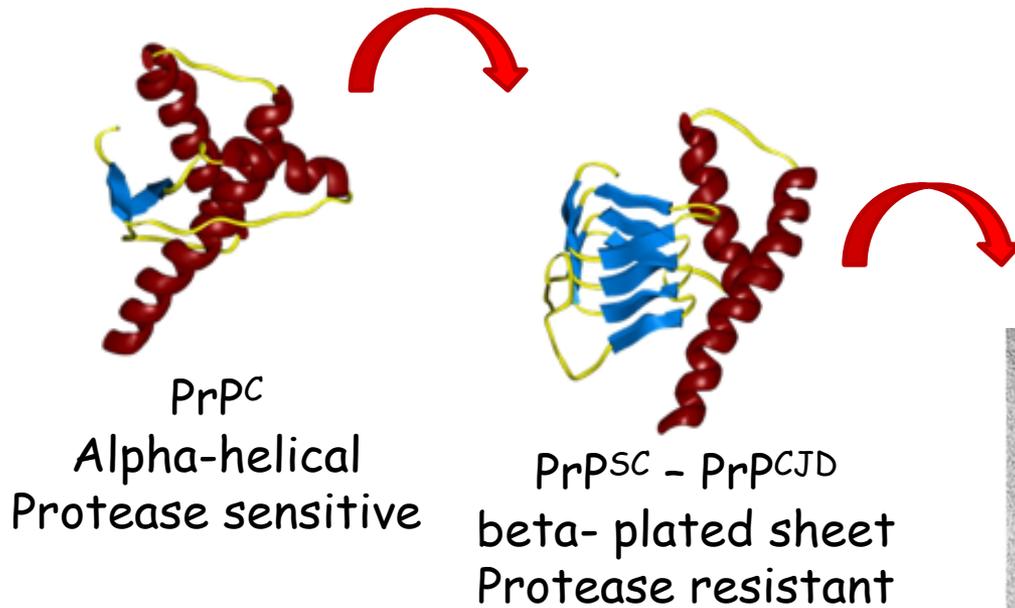
1310  $\text{cm}^{-1}$  Amide III



Strong correlation among different carbohydrates, glycocalyx and Amide III bands. Glycocalyx synthesis is prerequisite for the formation of stable bacterial biofilms

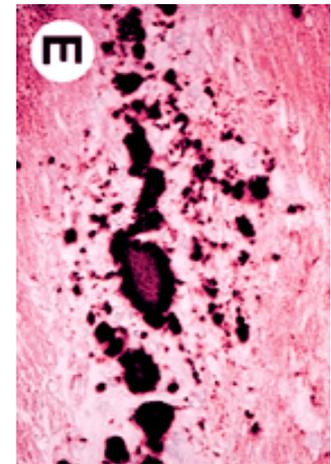
# Prion disorders

## Aberrant metabolism of the Prion Protein (PrP)



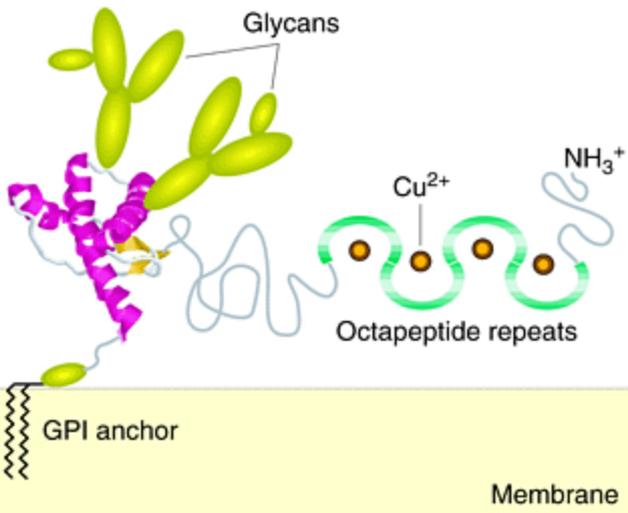
Prion rods

PNAS 1999 vol. 96 no. 26 15137-15142



PNAS 1998 vol. 95 no. 23 13363-13383

PrP amyloid plaques

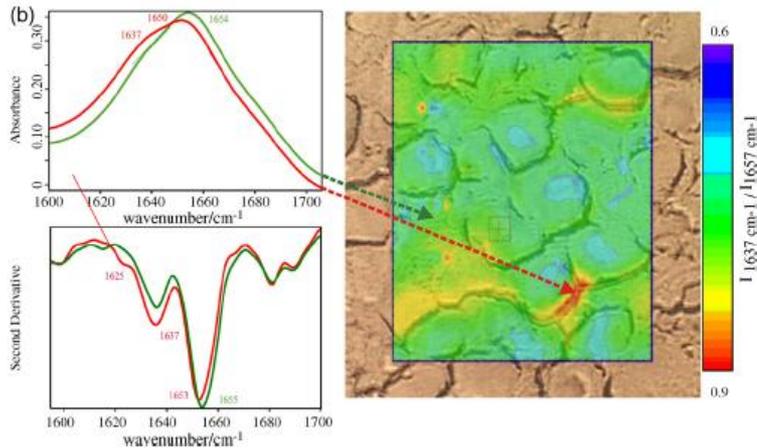


# SR-IRMS and prion research

✓ FTIR spectroscopy has been largely employed for studying the conformational changes associated to the conversion of PrP<sup>C</sup> into PrP<sup>Sc</sup>

PrP<sup>C</sup> 42%  $\alpha$ -helix; 3%  $\beta$ -sheet // PrP<sup>Sc</sup> <  $\alpha$ -helix; >  $\beta$ -sheet Phenotype dependent

✓ IRMS is able to distinguish between scrapie (S) and normal (N) brain tissues but the spectroscopic differences between different cerebellar substructures are much pronounced than disease alterations.



Differentiation is based on the superimposition of multiple contribution more than on the identification of structural protein alteration, evidenced only but not always in terminally ill animals (Dilution effect)

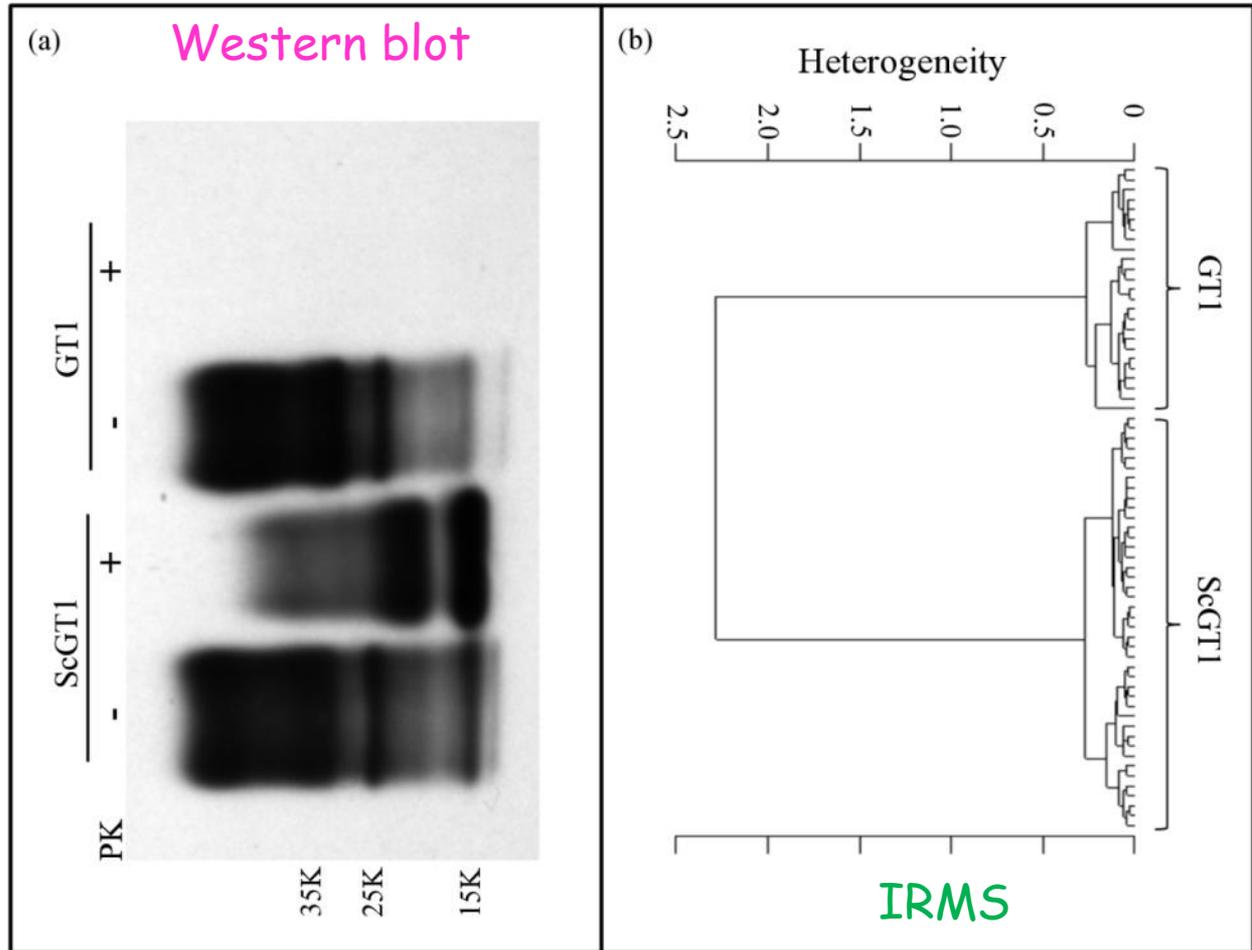
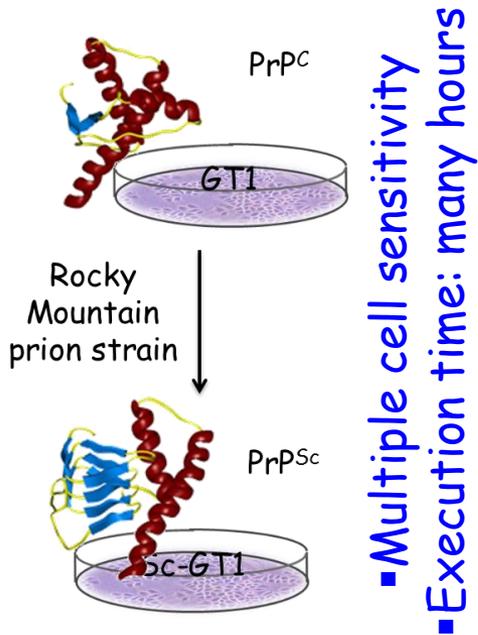
Tissue architecture complexity is limiting the understanding of cellular bases of disease

A. Kretlow wt al., FTIR-microspectroscopy of prion-infected nervous tissue, *Biochimica et Biophysica Acta (BBA) - Biomembranes* (2006), 7:948-959

M. Beekes et al., Analytical applications of Fourier transform-infrared (FT-IR) spectroscopy in microbiology and Prion research, *Veterinary microbiology* (2007), 123:305-319

# SR-IRMS and prion research: a cellular study

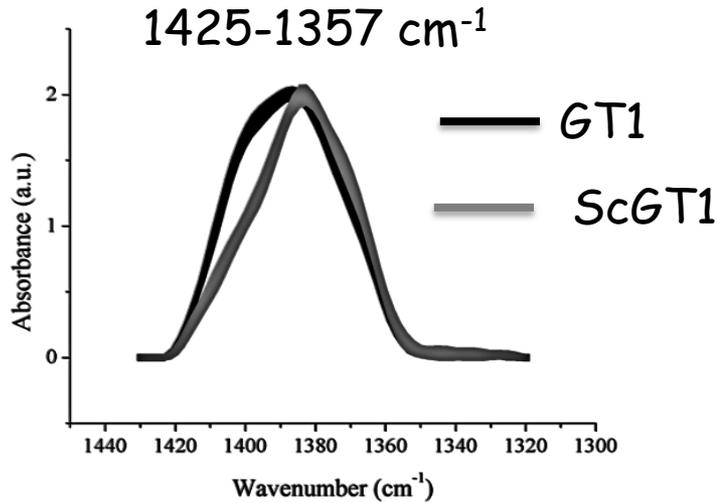
IRMS is a sensitive single-cell diagnostic tool for testing prion infection, faster than conventional Western blot PK digestion assay



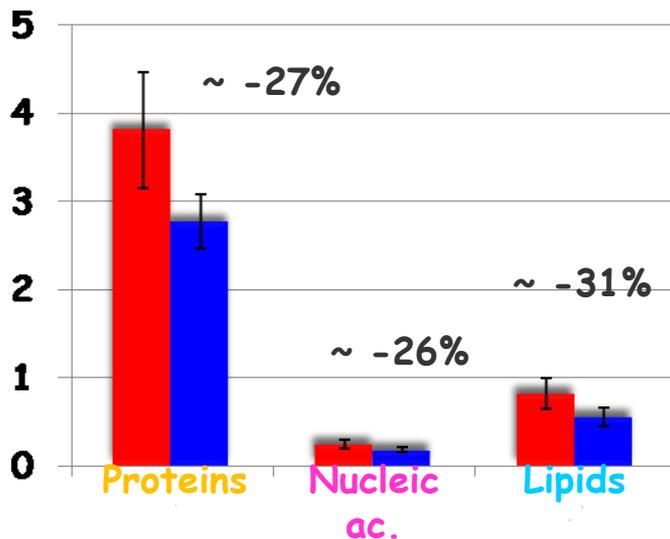
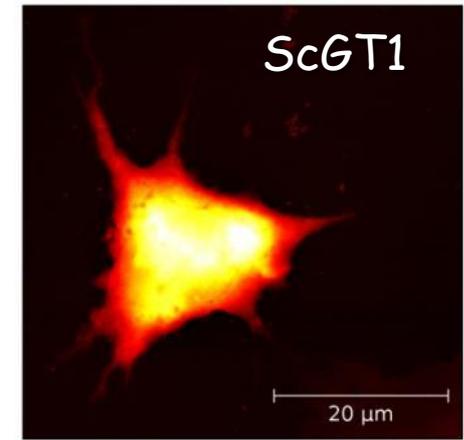
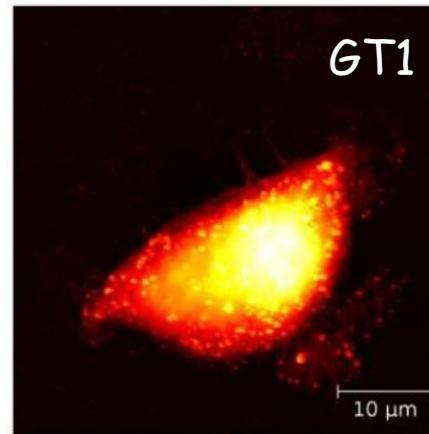
- Single cell sensitivity
- Execution time: few minutes

# IRMS revealed the biochemical reasons of classification

- Increase Glu and Asp protonated aminoacid

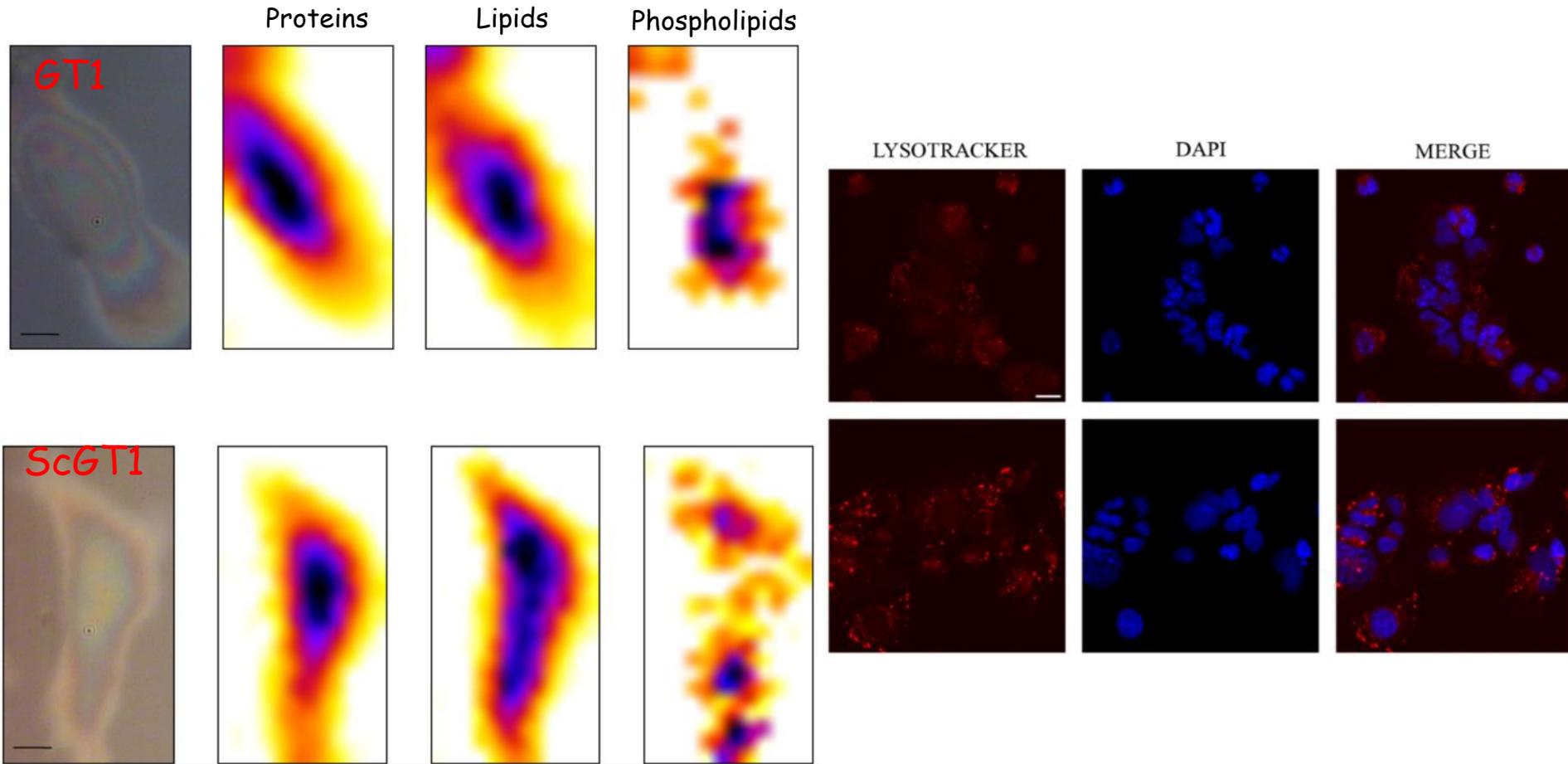


- Down-regulation in protein and lipid synthesis upon prion infection was elicited by semi-quantitative analysis



AFM: GT1 and Sc-GT1 have comparable pyramid like shape and effective cell height

# IRMS revealed an increase in both number and dimension of lysosomal compartments upon prion infection



The synergic matching of IRMS with complementary investigation tools is a winning strategy for shining a light on cellular phenomena behind prion infection

# YOUR EXPERIMENT

## The sample

Root thin sample slides deposited on TEM grids

## Data Acquisition

FTIR imaging with conventional source and FPA detector

IRSR with single point MCT detector (Mapping of small areas)

## Data pre-treatment

## Data Analysis

Univariate data analysis (band integration)

HCA

PCA

Sample regions exposed and non to X-rays will be monitored. The radiation damage effects will be considered

